

Novel regulators of prostate cancer stem cells and tumor aggressiveness $\mathsf{Zoni}, \mathsf{E}.$

Citation

Zoni, E. (2016, June 2). *Novel regulators of prostate cancer stem cells and tumor aggressiveness*. Retrieved from https://hdl.handle.net/1887/39840

Version:	Not Applicable (or Unknown)
License:	<u>Licence agreement concerning inclusion of doctoral thesis in the</u> <u>Institutional Repository of the University of Leiden</u>
Downloaded from:	https://hdl.handle.net/1887/39840

Note: To cite this publication please use the final published version (if applicable).

Cover Page



Universiteit Leiden



The handle <u>http://hdl.handle.net/1887/39840</u> holds various files of this Leiden University dissertation.

Author: Zoni, E. Title: Novel regulators of prostate cancer stem cells and tumor aggressiveness Issue Date: 2016-06-02

1

General Introduction

1. General Properties of Cancer

Cancer is considered one of the leading cause of morbidity and mortality worldwide, with 14 million new cases and 8.2 million cancer-related deaths registered in 2012 (1). The 5 most common cancers diagnosed in women in 2012 were breast, colorectal, lung, cervix and stomach cancer while in men these include lung, prostate, colorectal, stomach and liver cancer.

Carcinogenesis is a complex multi-step process that usually proceeds over several years and starts from one single cell. It progressively drives normal cell evolution into a cell with an increasingly abnormal neoplastic phenotype. This process is the result of a combination of genetic and epigenetic factors determined by individual variability caused by hereditary predisposition, life style and other variables like environmental influences, infectious agents, nutritional factors, hormonal and reproductive factors, and exposure to physical, chemical and biological carcinogens (2). Tumor formation and progression is driven by a sequence of essential alterations in cell physiology, cell homeostasis, randomly occurring mutations and epigenetic alterations of DNA. These events affect genes controlling different processes, such as cell proliferation, differentiation and survival, and bring cancer to acquire different malignant capabilities that together lead to malignant growth. The genetic abnormalities that contribute to cancer pathogenesis generally involve two main mechanisms: the inactivation of negative mediators of cell proliferation (tumor suppressor genes) and the activation of positive mediators of cell proliferation (protooncogenes) (3).

The definition of cancer, as established nowadays by the last advances in the tumor biology, includes multiple characteristics and aspects, that surprisingly are already present in the etymology of the word itself. The word "Cancer" originates from the ancient Greek word "καρκίνος" (Karkinos, "crab") credited to the Greek physician Hippocrates (460-370 BC). This word was probably chosen to describe the similarities of solid tumors with swollen veins and spreading projections reminding the shape of a "crab". Strikingly, this description includes exactly all the elements that the modern biology of tumors ascribe to cancer; a primary tumor mass, the presence of new vessels and the spreading and invasion of the neighboring tissues. Today we learned that the biology of tumors should be investigated not only focusing on the traits of single cancer cells, but also considering the contributions of the tumor microenvironment, the interactions between tumor cells and the supportive stroma, the role of the immune system and the preferential tropism of spreading tumor cells for specific metastatic sites. These interactions are fundamental to understand the mechanisms that lead to the switch from a contained disease to the aggressive spreading and metastatic phase of tumors. These events and characteristics have been systematically outlined in 2000

by Hanahan and Weinberg as "Hallmarks of Cancer" (4) and more recently updated to include the supportive cellular and non-cellular microenvironment (5,6).

1.1. Hallmarks of Cancer

There is a certain series of events that have to occur to drive the transformation from a normal cell to a cancer cell. These events are part of a multistep process and all the steps involved in this process contribute progressively to the generation and development of cancer. The fundamental characteristics of cancer or the hallmarks of cancer represent the set of properties that a cell or a group of cells in general have to acquire to become a tumor and to interact with the surrounding stroma (5,6) (**Fig. 1**):

Sustaining Proliferative Signaling: normal cells are constantly proliferating as part of the physiological turnover present in every normal tissue. However, their proliferation is finely tuned and regulated by multiple growth factors to maintain a proliferative rate appropriate for the maintenance of the homeostasis of the tissues where they home. These growth factors are part of a paracrine signaling and their availability or signaling efficacy depends also on the capability of the "receiving" normal or tumor cells to properly react to these stimuli. In cancer, the tumor cells can instruct the supportive tumor-stroma to supply growth factors (7) or acquire a "self-sustained proliferative signaling" resulting in an autocrine and abnormal proliferative stimulation.

Evading Growth Suppressors: part of the regulation of the maintenance of the homeostasis of cells and tissue is determined by the suppression of the proliferation. Normal cells have to proliferate to generate new tissues and maintain the tissue integrity but also have to stop their growth to prevent abnormal hyper-proliferation. In cancer, tumor cells have to escape these suppressive mechanisms and have to circumvent the programs that negatively regulates cell proliferation. Typical alterations in tumor suppressor genes include the loss of function of critical "gatekeeper" of cell-cycle progression such as pRb (Retinoblastoma 1) and p53 which regulates apoptosis and is a stress responsive sensor.

Resisting to Cell Death: Programmed cell death, apoptosis, is one of the mechanisms in normal physiology that prevents cancer development and the afore mentioned p53 is one of the key player in this process. There are two main circuits that orchestrate apoptosis: one receives and process extracellular death-inducing signals (e.g. Fas and Fas ligand mechanism) and the other sense intracellular signals (intrinsic program). Both the machineries converge on the activation of a cascade of proteolytic cleavage events involving latent and effector caspases. Cancer cells have evolved a variety of

strategies to avoid this programmed death. The most common feature of tumor cells evading apoptosis is the loss of p53; other strategies include the suppression of proapoptotic factors (e.g. Bax, Puma, Bim) or the upregulation of anti-apoptotic genes (e.g. Bcl-2, Bcl-X_L).

Enabling Replicative Immortality: normal cells in healthy tissues are capable of pass through only a limited number of division cycles. The mechanisms that limit the number of growth-and-division cycles are essentially two: senescence (nonproliferative but viable state) and crisis (cell death). Both are linked to the length of telomeric DNA that in a cell dictates how many cycles of division are still available before the cell enters in a phase of DNA instability (i.e. crisis). It is a remarkable properties of cancer cells their ability to proliferate indefinitely, escaping from this control. One of the mechanisms that drive immortalization in tumor cells is the presence of telomerase activity, responsible for the integrity of telomeric DNA, detected in up to 90% of spontaneously immortalized cells.

Inducing Angiogenesis: in every normal tissue, nutrients and oxygen are provided by a fully functional network of vessels, responsible also for the elimination of wastes and other products of the metabolism. In normal physiology, in an adult organism these vessels remain mostly stable and quiescent. In malignant tumors, vessel remodelling and new vessel formation occurs after the so-called "angiogenic switch" which cause the normal quiescent vasculature to sprout and produce new branches (neovascularization). This abnormal angiogenesis produces vessels that are histologically different from those formed during the physiological process. Moreover, highly invasive tumor cells can form fluid-conducting channels in a process defined as "vasculogenic mimicry" (8). The new vessels in the tumor have an aberrant morphology and are characterized by abnormal level of endothelial cells proliferation and apoptosis. In addition, the leakiness that characterize these vessels is one of the major effectors for the low efficiency in the delivery of therapies specifically to the tumor.

Activating Invasion and Metastasis: the invasion of the surrounding tissues, the intravasation in blood and lymphatic vessels and the formation of distant metastasis represent one of the critical problem in tumor progression. In this context transformed epithelial cells acquire a motile mesenchymal phenotype in a process referred to as "epithelial to mesenchymal transition" (EMT) (9). However, cancer cell migration is not restricted to singly migrating cells. Different patterns of cell migration include single-cell migration, multicellular streaming and collective cell migration, (reviewed in (10)). The impact of this process during the onset of metastatic spreading and its relevance in the

establishment of therapy resistance will be discussed more in details in the next paragraphs.

Interaction with the tumor stroma: tumor cells do not behave independently from the rich microenvironment where they are localized and that represents an important component during tumor initiation, growth and progression. During cancer progression the stroma co-evolves with the tumor and create a dynamic signaling network of paracrine signals that promotes cancer. The different stromal components involved in this process include: cancer-associated fibroblasts (CAFs), pericytes, endothelial cells, immune cells and the extracellular matrix (reviewed in (6)).

Metabolic reprogramming: In addition to the above mentioned specific characteristics, tumor cells can also adapt their metabolism and switch to the so called "aerobic glycolysis" converging their metabolism largely to glycolysis (i.e. Warburg-effect) (11). This is one of the basis of the non-invasive visualization of tumors based on positron emission tomography (PET) with radiolabelled analog of glucose as reporter. In proliferating (cancer) cells the mitochondrial metabolism is reprogrammed toward macromolecular synthesis to sustain multiple cell divisions, (reviewed in (12)). Moreover, oncogenic mutations in metabolic enzymes such as the cytosolic NADP⁺-dependent isocitrate dehydrogenase 1 gene (IDH1) and the mitochondrial homolog IDH2 responsible for converting α -ketoglutarate to 2-hydroxyglutarate (2HG), a metabolite found only in reduced amounts in mammalian cells under normal conditions have been reported (12,13). Interestingly this has also effects on epigenetic mechanisms, resulting in altered histone methylation marks, hypermethylation at CpG islands and dysregulated cell differentiation(12).

Moreover, it is important to note the pro-inflammatory and immunosuppressive properties of cancer cells. Inflammation can sustain proliferative signaling and inhibiting cell death, activate extracellular matrix-modifying enzymes and support invasion and angiogenesis (14,15). Tumor cells can also secrete immunosuppressive factors or recruit immunosuppressive cells, blocking the action of cytotoxic lymphocytes or recruit tumor associated macrophages that can enhance tumor progression and metastasis and suppress antitumor immunity (16-18).

All the aspects discussed above depend, to a large extent, on genomic alterations in neoplastic cells. Different cells can gain different alterations and the combination of several alteration together will produce a cancer cell, capable of outgrow and gain a local dominance over other neoplastic and/or normal cells. In this perspective, tumor progression is characterized by the expansion of different heterogeneous clones. In this process, only those clones capable of gaining all the hallmarks of cancer will succeed in generating a malignant tumor. The issue of intra-tumor and inter-tumor heterogeneity and its impact on resistance to current therapies will be discussed in the next paragraphs.



Figure 1. Schematic representation of the hallmarks of cancer, their correlation with cancer progression and reciprocal interaction with the tumor stroma.

1.2. Tumor Heterogeneity

There are two main levels of complexity in tumor heterogeneity. If we consider the tumor mass as an independent entity, one level consists of the differences between different cancer types or different patients affected by the same cancer and is defined as inter-tumor heterogeneity (Fig. 2). Another level of heterogeneity encompasses the cellular differences within the same tumor (e.g. multiple cell clones with different properties, dispersed within the same tumor mass of the same patient) and is defined as *intra-tumor heterogeneity* (Fig. 2). Despite the fact that, overall, the evolution and progression of these tumor can be similar (e.g. onset of primary tumor, progression from benign to malignant growth, neo-angiogenesis, invasion of surrounding tissues, and formation of distant metastasis) there are intrinsic differences that distinguish one cancer from the other and, within the same cancer family, one cancer subtype to another (e.g. hormone-naïve or androgen-independent prostate cancer). These differences are part of the so-called inter-tumor heterogeneity and reflect the differential responsiveness to specific therapeutics and not to others. Additionally, the tropism for specific metastatic sites (e.g. osteotropism in prostate cancer), is also characteristic of certain malignancies and can be ascribed to the inter-tumor heterogeneity.

The second level of complexity comprises intra-tumor heterogeneity. As already discussed, cancer formation is a multistep process that starts from one single cell; on the other hand, tumors are tissues and therefore are constituted by a variety of different tumor- and other-cell types. As established in the last decade by high-resolution genome-wide studies, the formation and progression of tumors is characterized by a continuous "Darwinian-like" evolution of branches of specific clones (19). This process of "clonal evolution" results also in the construction of a supportive tumor microenvironment, which is continuously being remodelled during the tumor progression. Different cell types contribute to increase the complexity and the heterogeneity of this environment. As previously mentioned, these cell types include, among the others, also non-malignant cells, such as immune and inflammatory cells, endothelial cells, pericytes, and cancer-associated fibroblast. For the purpose of this thesis we will mainly focus on the cancer cells and discuss the different cell types and subpopulation that are represented within the tumor. Macroscopic tumors are constituted of different subpopulation of malignant cells. Depending on environmental stimuli and stochastic processes, or depending on their alterations, such as mutations and epigenetic changes, these clones can acquire a dominant phenotype with clinically relevant characteristics (e.g. resistance to therapy). In this respect, the intra-tumor heterogeneity is one of the relevant problems in the identification of therapeutic strategies capable to eradicate completely all the different cancer cell subpopulations

and clones that maintain the cancer. Similarly, this also has an impact on studies that approach tumor biology without distinguishing the different cell types dispersed within the tumor.



Figure 2. Schematic representation of inter-tumor and intra-tumor heterogeneity. Adapted from [19].

Molecular- and genetic-profiles of cancerous "bulk" tissues indeed cannot discriminate between the aggressive subsets of cancer cells responsible for tumor maintenance, growth & the development of therapy resistance and less aggressive, more differentiated cancer cell subpopulations. This raises the question whether the different clones and subpopulations present in the tumor are properly represented also in a transcriptional analysis between "bulk" tumor and "normal" tissues. One of the aspects of tumor heterogeneity, that has revolutionized the tumor biology in the last years, is indeed the discovery of a subpopulation of cancer cells with tissue stem-like properties, the cancer stem cells (CSCs). The contribution of these cells to the tumor formation and maintenance, metastasis, therapy resistance and their clinical relevance for the identification of new therapeutic strategies will be discussed later.

2. Anatomy of the Prostate

The prostate is a walnut-sized exocrine gland, well encapsulated and positioned in the pelvic cavity inferior of the bladder (it surrounds the first tract of the urethra) and anterior of the rectum (20) (**Fig. 3**). The function of the prostate is to secrete a slightly alkaline milky white fluid, that constitutes about the 30% of the volume of the semen and that contains carbohydrates, phospholipids, and enzymes (e.g. prostate specific acid phosphatase (PAP) and prostate specific antigen (PSA)) (21).



Figure 3. Anatomy of the prostate within the male reproductive system.

Source: http://www.aboutcancer.com/prostate

The prostate can be divided in four "zones" (mainly used in pathology, (22,23)) (**Fig. 4**) or in four lobes (mainly used in anatomy):

- Central zone: it surrounds the ejaculatory ducts and constitutes about 20% of the whole gland and it presents large and irregular ducts. Approximately 1 – 5% of prostate cancer originates from this region and tend to invade the seminal vesicles (24). This part roughly corresponds to the median lobe.
- 2) *Peripheral zone*: accounts for the majority of the gland and it originates from the mesoderm. Up to 70% of prostate cancer originates from this part. Roughly corresponds to the posterior lobe.
- 3) *Transition zone*: it surrounds the prostatic urethra and it originates from the endoderm. About 20% of prostate cancer originates from this zone which is also responsible for the formation of benign prostatic hyperplasia (BPH, discussed later (25)). Roughly corresponds to the anterior lobe.
- 4) Anterior fibromuscular stroma: it consists of muscular and fibrous tissue.

The "fourth lobe" is named lateral and spans all zones.



Figure 4. Structure of the prostate. The four zones are indicated: transition zone, central zone, peripheral zone and anterior fibromuscular stroma. From Baylor College of Medicine 1990.

The growth of the prostate is regulated by androgens like testosterone. Testosterone is mainly produced by Leydig cells in the testis and to lower extent in the adrenal cortex

and its synthesis is controlled by luteinizing hormone (LH) and the follicle stimulating hormone (FSH). The secretion of LH from the pituitary gland is regulated by the hypothalamic luteinizing hormone-releasing hormone (LHRH) (26). When testosterone is converted to 5 α -dihydrotestosterone (DHT) by 5 α -reductase, it can stimulate the growth of the prostate (for example during puberty) (27). DHT in the blood is associated with the sex hormone-binding protein (SHB), responsible for its transportation into the vasculature and to the target cells, where androgens bind and activate the androgen receptor (AR). Activation of the AR by androgens results in the transcription of androgen-responsive genes like PSA or the prostate-specific gene TMPRSS2 (28,29).

2.1. Architecture of the prostate

The prostate has a glandular structure characterized by several ducts constituted by three major cell types: luminal, basal and neuroendocrine cells (30) (**Fig. 5**). These cells are different in morphology, function and significance for tumorigenesis.

Luminal cells: these cells are located along the glandular lumen and have a secretory function. They are terminally differentiated and express specific differentiation markers such as AR and cytokeratin 8/18 (31,32). Additionally, they are androgen regulated and produce PSA and PAP. Cells with self-renewal properties have been identified within the luminal compartment in mice and humans (33,34) and proposed as the cell-of-origin of castration resistant prostate cancer (CRPC, discussed later).

Basal cells: these cells are located between the luminal cells and the basal layer that separates the epithelium from the stroma, which consists of fibroblasts, blood vessels, nerve cells, smooth muscles cells, infiltrating immune cells and connective tissue. The contribution of these cells and components to prostate cancer is crucial, especially during the progression of the disease (35,36). Basal cells are proliferative and characterized by the expression of cytokeratin 5 and 14 (37,38). Experimental evidence have shown the presence of stem-like cells within the basal compartment (39,40) which could maintain basal cells or differentiate into luminal cells and neuroendocrine cells (41,42).

Neuroendocrine cells: these cells are dispersed within the basal layer and are androgen independent; they express different neuropeptides like serotonin and chromogranin A (43). It is hypothesized that their function is to participate in the differentiation of the normal prostate and they also play a role in tumorigenesis (44,45).



Figure 5. Histology of the prostate. The epithelial layer that characterizes the prostate consists of basal cells (Cytokeratin 5 positive, bottom left), separated from the stroma by the basal lamina; neuroendocrine cells, dispersed within the basal cells (Chromogranin A positive cells, top right) and luminal cells (Cytokeratin 18 positive cells, bottom right). Source: www.proteinatlas.org.

3. Diseases of the Prostate

Due to its high blood perfusion and connection with the urethra, the prostate gland is susceptible to acute and chronic bacterial infection (i.e. prostatitis) typically treated with antibiotics (46). Moreover, during aging, the prostate increases physiologically in size and this can result in benign prostatic hyperplasia (BPH), classified in two types: histologic BPH, characterized by microscopic evidence of epithelial and stromal hyperplasia, and macroscopic BPH, characterized by an enlargement of the prostate (25). Three main theories have been proposed to explain the etiology of BPH (47): 1) the enlargement of the prostate could be caused by a shift in the prostatic androgen metabolism occurring with age, which lead to abnormal accumulation of dihydrotestosterone; 2) changes in epithelial-stromal interaction induce prostatic growth; 3) an expansion in epithelial stem cells. Clinical manifestations of an enlargement of the prostate include lower urinary tracts symptoms such as bladder outlet obstruction and chronic urinary retention which results in additional complications (e.g. infections). Treatment options include pharmacologic agents employed to relax the prostatic smooth muscle (alpha-blockers, 5-alpha reductase

inhibitors) or transurethral resection (48,49). Additionally, there are some similarities between BPH and prostate cancer as both require androgens for growth and development and therefore might respond to antiandrogen treatments (50).

4. Prostate Cancer

4.1. Epidemiology

Prostate cancer is the second leading cause of death from cancer in males in western countries with 220,800 new cases estimated for 2015 (51). The incidence of prostate cancer increases with age (prostate cancer is a rare event in men under the age of 50), and it is higher in the western world compared to less developed countries, due to differences in life-style, eating habits, environmental agents and ethnicity (52). However, there is homogeneity in the age-dependent prevalence of prostate cancer in different countries.

4.2. Prostate Cancer Initiation

Prostate cancer is considered a multifocal disease. The primary tumor often presents multiple histologically independent foci that can be genetically identified for their properties and are relevant for understanding the distinction between latent and clinical disease (30). Although prostate cancer is commonly considered a disease of older men, analysis of specimen collected from younger healthy individuals revealed the presence of histologic foci of prostate cancer also in men in their 20s to 40s, suggesting an early onset of cancer (53). In the majority of the cases, these multifocal lesions will result in a latent disease that will not progress to clinically detectable and relevant prostate cancer. This can be explained by two hypothesis: there is a critical difference between the initiation of the pathogenic program of latent and clinical prostate cancer, or the critical events that are needed to generate a clinical disease do not occur in the latent foci. In the initiation phase, the normal prostate gland present a ductal-acinar histology, characterized by an organized epithelium with luminal secretory cells, basal and neuroendocrine cells and a basal lamina (Fig. 5). This organized structure is altered during the "initiation stage", where histological changes of the luminal epithelium occur and lead to luminal epithelial hyperplasia defined as prostatic intra-epithelial neoplasia (PIN) (54). PIN lesions are classified between lowgrade and high-grade, they are multifocal and, at this stage, the cancer is contained by the intact basal layer which prevents the invasion of the surrounding tissues. The morphological integrity of the glandular structures prevents also an increased release of PSA into the blood stream. For this reason PIN lesions are usually detected by biopsy and not by blood test as they don't produce increased PSA levels (55). High-grade PIN

lesions are characterized by high expression of proliferation markers (56) and by the histological presence of basal cells (30).

A number of genomic alterations such as copy number variations and chromosomal rearrangements (insertions, deletions) associated with prostate cancer and hereditary prostate cancer have been identified with multiple genome analysis studies (30).

Among the most common copy number alterations, those occurring at 8p21 (Nkx3.1), at 10q23 (PTEN) and at 8q24 (MYC) involve key regulatory genes (30,57-59). In addition, genome-wide association studies have shown the involvement of HPC1 and HPC2 (mapped in 1q24-25 and 17p11 respectively) in hereditary prostate cancer (60,61). The down regulation of Nkx3.1 is one of the critical events in prostate cancer initiation and is detected in up to 80% of the prostate tumors (also in PIN lesions and early invasive carcinoma). Nkx3.1 has a critical role in prostate morphogenesis and differentiation and mutant mice for Nkx3.1 develop PIN lesions that resemble closely those detected in human.

Another frequent chromosomal loss detected in a high percentage of prostate cancer cases is represented by PTEN (tumor suppressor gene). Loss of PTEN leads to hyperactive PI3K-AKT-mTOR signaling, which results in aberrant cell proliferation and metabolism (62). Recent studies have shown that the allelic loss of PTEN represents an early event in prostate carcinogenesis and correlates with progression of the disease (59). As for Nkx3.1, also loss of PTEN in mice results in PIN lesions and/or adenocarcinoma (63).

Besides the aforementioned chromosomal losses, genetic studies have identified also gene amplifications in prostate tumors. The oncogene MYC is amplified during initiation of prostate tumors and upregulation of MYC has been registered in PIN lesions (64). Similarly, transgenic mice overexpressing human MYC undergo rapid formation of PIN lesions, followed by progression to invasive adenocarcinoma (65). Another set of genetic alterations is represented by chromosomal rearrangements. Among these, the most common alteration regards the family of transcription factors (ERG, ETV1 and ETV4) and the prostate specific gene TMPRSS2 (66). The most frequent rearrangement produces the fusion gene TMPRSS2-ERG, where the N-terminally truncated ERG protein is expressed under the control of the promoter of the androgen-regulated TMPRSS2 gene (66). This alteration usually occurs in cancer initiation and is also detected as early event during cancer progression.

4.3. Prostate Cancer Detection

The oldest known case of prostate cancer diagnosed reliably by morphological and biochemical techniques dates back to 2,700 years ago (67). Schultz and co-workers

described that a well preserved skeleton of a 40 to 50-year-old Schythian king who lived during the Iron Age in the Southern Siberia (Arzhan) suffered from macroscopically visible osteoblastic and osteolytic lesions throughout the entire skeleton (67). This diagnosis is based on microscopic imaging of the lesions and detection of prostatespecific-antigen (PSA) complexed with α 1-antichymotrypsin (PSA/ACT) in the extracellular matrix (ECM) proteins extracted from the compact cortical bone of the skeleton from Arzhan (68,69).

The blood test for PSA, nowadays routinely used in the clinic, has revolutionized the clinical practice over the past four decades and has represented the standard for prostate cancer detection and monitoring. PSA is a glandular kallikrenin-related peptidase produced by the gene KLK3 and its transcription is regulated by androgens which make its expression a main characteristic of the prostate epithelium (70). PSA is continuously produced by the healthy prostate. In the normal prostate, the morphological structure of the glands contains PSA tightly confined and only a reduced amount is released into the blood (0.6 ng/ml in a healthy adult male) where it exits in multiple forms: as pro-protein or mature protein and free or associated with different protease inhibitors (70). In BPH or prostate cancer, the disruption of the normal prostate architecture often results in a massive release of PSA into the blood (>100 ng/ml) that is measured almost exclusively in males with advanced prostate cancer (70). Altered PSA levels in blood are also commonly detected during the occurrence of other alterations of the prostate such as inflammation (prostatitis) and its' levels are also influenced by age. For this reason, PSA is considered as a prostate-specific marker but not a cancer-specific marker. In this perspective, two more specific and clinically-promising markers for prostate cancer detection are represented by the non-coding messenger RNA for the so called Prostate Cancer Antigen 3 (PCA3), identified in 1999 (71) and the fusion gene TMPRSS2-ERG described in 2005 (66). However, although PCA3 and TMPRSS2-ERG (also as combined biomarkers) displayed higher specificity and diagnostic accuracy for prostate cancer outcome, PSA is still the most widely used biomarker in prostate cancer diagnosis (72). Importantly, the expression of KLK3 at molecular level in the prostate epithelium and the increase of PSA level in the blood of men affected by prostate cancer are not directly correlated (73). The detection of augmented PSA level into the blood is indeed determined by an increased release of PSA from the prostatic gland as a consequence of disruption of normal prostate architecture and not by an increase of its transcription. This leads to the documented paradox that during development and progression of prostate cancer, KLK3 expression might slightly decrease (74).

The typical clinical practice for men with high PSA levels schedules biopsy to assess the possible presence of prostate cancer. The prostate tissue collected is then graded according to the Gleason scoring system introduced for the first time in the clinic

in the 1960s and recently updated in 2005 (75-77). The Gleason scale describes the primary and secondary architectural pattern of the tissue obtained from prostate biopsies and classifies tumors according to their differentiation, from 1 to 5, based on the morphological architecture of the prostate (76,78). Briefly, Gleason 1 corresponds to a transformed prostate epithelium that resembles a normal prostatic epithelial tissue; from Gleason 2 to Gleason 4, the infiltration of cells at the margin of the gland is progressively increasing; Gleason 5 corresponds to a cancerous prostate which has completely lost its epithelial structure and is filled with invading mesenchyme-like cancer cells. The final Gleason score is obtained upon mathematical addition of primary and secondary score and can range from 2 to 10 (79). In addition, the status of the primary tumor is also graded, from organ-confined to fully invasive (T1-4), with or without involvement of lymph nodes (N0 or N1) and with or without presence of distant metastasis (M0 or M1 a-c) (80). These together constitute the so called Tumor-Node-Metastasis (TNM) system of grading.

4.4. Treatment of Localized Disease

There are several options for treating prostate cancer patients with localized disease, depending on the stage and the patient condition.

As previously mentioned, due to improved screening methods, prostate cancer can be detected already at the very initial stage. Active surveillance is considered a logical approach for those men with localized prostate cancer and associated low-risk to prevent overtreatment (81-83). The clinical criteria to define an active surveillance strategy are: confined disease with T1-T2 stage, maximum PSA level of 10 ng/mL and Gleason score <7 (84). Additionally, watchful waiting is considered an alternative for old men with less aggressive disease (85).

Surgical approaches like the removal of the entire prostate are applied to men with high life expectancy and localized disease with the aim to completely eradicate the tumor (86). In these patients, given the low risk of lymph node involvement, the removal of pelvic lymph nodes remains controversial (86,87).

Another therapeutic approach for the localized disease is radiotherapy which employs x- and gamma-rays or alpha emitting radio isotopes (88) to kill tumor cells by causing DNA damage. Two applications are possible: external beam radiotherapy and internal radiotherapy (also called brachytherapy, which consists of implantation near the cancerous region of radioactive plugs which will release slowly the radiation). Recently, also image guided intensity-modulated radiotherapy has been developed to deliver high dose particles to specific regions reducing the impact on the surrounding tissues (89). It is important to consider that although surgical removal and radiotherapy produce a

similar outcome, they have a different impact on the quality of life of the patient (e.g. urinary and sexual function) (90).

4.5. Prostate Cancer Progression and Bone metastasis

When the cancer enters into the "progression phase", the loss of the basal lamina occurs and results in the switch from high-grade PIN to adenocarcinoma with an invasive phenotype, macroscopically characterized by the lack of basal cells as shown by p63 and cytokeratin 5/14 staining (91). However, whether prostate cancer is originated from luminal or basal prostate cancer stem cells is still under debate (33,92). The majority of prostate adenocarcinomas present with an acinar morphology while ductal and mucinous adenocarcinomas are more rare. In less than 2% of the cases the adenocarcinomas are classified as neuroendocrine variants and mainly occur during recurrence after androgen deprivation therapy (93). This can partially be explained by the fact that neuroendocrine cells lacking of AR expression survive ADT and prevail producing relapse (94).

The terminal phase of prostate cancer progression encompasses systemic metastasis, which coincides with the development of therapy resistance, e.g castration and chemotherapy resistance (95). The majority of aggressive prostate cancers is characterized by their osteotropism leading to the development of predominantly osteoblastic/osteosclerotic lesions and, thus, represent one of the major clinical challenges in uro-oncology. The first explanation for the bone tropism of prostate cancer metastasis was provided in 1940 when Oscar Batson suggested that the venous network that drains the prostate and connect the pelvic veins to the paravertebral venous plexus could explain the dissemination (96). However, another study demonstrated that the venous network does not represent the major driver in the dissemination of prostate cancer cells to the bones (97). Alternatively the interactions between cancer cells and the endothelium was also suggested to underlie organspecific dissemination (98). Furthermore, the interaction between the chemokine (C-X-C motif) receptor (CXCR) 4 (CXCR4) and its ligand stromal derived factor 1 (SFD1, also known as CXCL12) may be critically important (99). Prostate cancer cells express CXCR4 and experimental evidence has shown that neutralization of CXCR4 reduces prostate cancer bone metastasis in preclinical models (100). Moreover, prostate cancer cells also express various integrins, e.g. integrin $\alpha v\beta 3$, which correlates with prostate cancer bone metastasis and is responsible for the interaction with fibronectin, vitronectin and osteopontin (101-104). The notion that molecular factors might be involved in the specific bone tropism of certain cancer cells was for the first time postulated by Sir Stephen Paget who introduced the "Seed and Soil" hypothesis in which he compared the bone metastatic breast cancer cells to the seed of plants, capable of growing only in

a fertile soil, the bone marrow (105). Today we know that the formation of distant metastasis is a complex process, characterized by multiple bi-directional interactions between the tumor cells and the supportive stroma (106). This process starts at the level of the confined primary tumor where factors systemically released contribute to the conditioning of the metastatic "soil" and provide the establishment of the so called "pre-metastatic niche" (107). The "pre-metastatic niche" is defined as a fertile microenvironment induced in the metastatic target organs that facilitates the future invasion, colonization and the proliferation of metastatic tumor cells (107). During the establishment of the "pre-metastatic niche", bone marrow-derived hematopoietic progenitor cells expressing VEGF receptor 1 (VEGFR1) are recruited to metastatic target organs by specific factors released by the primary tumor (108). Among these factors, LOXL enzymes, VEGFA, VEGFC, TNF α and TGF- β produced by the primary tumor stimulate inflammation, attachment and recruitment of, for example, myeloid cells and the expansion of lymphatic vessels in the proximity of the sentinel lymph nodes (109-112). Interestingly it has been proposed that extracellular vesicles and exosomes released from the primary tumor represent the mechanism of communication between the primary cancer cells and the metastatic sites during the induction of the "premetastatic niche" (113) also in prostate cancer (114).

Additionally, in primary and metastatic cancers, tumor cells interact with different cell types that constitute the stroma. Such cells include tumor-associated macrophages (TAMs), cancer associated fibroblasts (CAFs), endothelial cells, pericytes and mesenchymal stem cells (MSCs) reviewed in (6,115). Tumor cells produce several factors that "activate" the surrounding stromal cells and induce remodelling of the EMC. These factors include fibroblast growth factor 2 (FGF2), vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), epidermal growth factor (EGF), interleukins colony-stimulating factors and TGF- β (116) and proteolytic enzymes (117) that remodel ECM, enabling cell migration. In the progression and castration resistant phase of prostate cancer, cancerous polarized-epithelial cells localized at the site of the primary tumor undergo biochemical changes and acquire an invasive and often mesenchymal phenotype (118,119) which confers them enhanced migratory ability, invasiveness, resistance to apoptosis and resistance to therapy, which are all properties resulting in a clinically-relevant phenotype (120,121). Together these events result in the invasion of the surrounding stroma and in the intravasation and circulation of cancerous cells in the blood stream. Tumor cells which possess stem cell-like characteristics, that survive in the circulation can extravasate at those distant sites where the "pre-metastatic niche" has previously prepared a fertile "soil" for future colonization. Recent research has highlighted the clinically relevant properties of the so called circulating tumor cells (CTCs) capable of surviving into the blood stream and in

distant metastatic sites (122). Once that these CTCs have colonized the metastatic site (e.g. the bone), they may activate a reverse program of mesenchymal-to-epithelial transition (MET) and remain dormant for years (123). Therefore it appears that these disseminating tumor cells (DTCs) can perpetuate in the bone the malignant progression and establish a "metastatic niche".

Typically, the "metastatic niche" is located at perivascular locations (124). CTCs and DTCs may, potentially, establish a metastatic niche through competition with hematopoietic stem cells (HSCs) for their niche at a perivascular location (124-127). Moreover it has been hypothesized that tumor cells can also create their own niche (125,128). PCa cells amplify the existing hematopoietic niche and induce de novo an ectopic epithelial tissue-of-origin niche which together with the amplified hematopoietic niche generates a hybrid niche, supportive for cancer cell growth (106) and reviewed in (129). DTCs can survive in the bone microenvironment as nonproliferating (dormant) cells that originate microscopic lesions (classified as micrometastasis) (130,131). The mechanisms that induce exit from dormancy are still largely unknown (131). However, it has been shown that a collagen-I enriched fibrotic environment plays a crucial role in the cytoskeletal reorganization in dormant cells and in their awakening from dormancy (132). Once that these cells escape from dormancy, they induce local inflammation, followed by vascular and bone remodelling and establishment of a distant secondary tumor (bone metastasis) (120,133). Recently, it was revealed that the molecular signature of the stroma response in prostate cancerinduced osteoblastic bone metastasis highlights the amplification of hematopoietic and prostate epithelial stem cell niche (106). This observation supports the notion that angiogenesis and osteogenesis are crucial processes involved in the formation and growth of osteoblastic bone metastasis. Moreover, a recent report described the presence of two different type of microvessels: type "H" (CD31^{high} and endomucin^{high}) and type "L" (CD31^{low} and endomucin^{low}) (134). Interestingly, angiogenesis and osteogenesis have been coupled to the type "H" vessels, that provide also signals for HSCs and where osteoblasts also reside (135). Moreover, the kinetic of type "H" vessels in mice shows a peak at week 4 and loss of type "H" endothelium during ageing has been documented (134). Together this support the involvement of angiogenesis in the homing of metastatic cells in the bones in preclinical mouse models.

The bone remodelling induced by metastatic cancer cells results in either bone formation (osteoblastic bone metastasis) or bone resorption (osteolytic bone metastasis) and interferes with hematopoiesis (133). In prostate cancer, the bone lesions are typically osteoblastic (133,136), however the co-existence of osteobastic and osteolytic response have been documented (137).

Factors inducing osteoblast recruitment and activity in prostate cancer are: BMP6 (138), and BMP modulators, such as Noggin (NOG) (139); IGF1 (140) VEGFs (141), wnt signaling (142) and modulators of Wnt signaling such as dickkopf (DKK) and Sclerostin (SOST) (139). On the other hand, factors modulating osteoclast recruitment and activity in prostate cancer are: MMP-7, which promotes osteolysis via cleavage of RANKL that stimulates osteoclastogenesis (143); Noggin which antagonizes bone morphogenetic proteins (BMPs) and impairs bone formation (139,144);

It has been hypothesised that osteolytic cancer cells produce PTHrP that stimulates osteoblasts to secrete RANKL. This in turn stimulates ostecolasts progenitor cells and leads to osteoclastogenesis therefore bone resorption. During this process, many factors such as TGF- β , IGF-1 and calcium are released from the mineralised matrix to further feed cancer cell growth, thus perpetuating this "vicious cycle" (133,145). In prostate cancer for example, the expression of the calcium sensing receptor by tumor cells makes them responsive to the release of calcium during bone resorption and leads to increased proliferation and PTHrP release (146,147). However, the inhibition of bone resorption as strategy to impair bone metastasis with agents such as bisphosphonates revealed no effect on cancer cell proliferation in animal studies (148,149) and clinical trials also in prostate cancer (150) suggesting that other mechanisms support tumor cell growth in the bone. In this perspective, the recent identification of the molecular stroma response in osteoblastic prostate cancer (106) supports the coupling of angiogenesis and ostegenesis in bone metastasis (134) and suggest that anti angiogenesis might impact on the growth of osteotropic prostate cancer cells in the bone.

4.6. Treatment of Advanced Disease

As previously described, PSA testing allows an early detection of many cases of the disease when the cancer is still confined and may therefore be successfully resolved by surgery or radiotherapy. However, after local treatment, 20-40% of the cases, biochemical relapse will occur (PSA > 0.2 ng/ml) (**Fig. 6**) (95). Typically these patients will be treated with androgen deprivation therapy (ADT, which consists of chemical or surgical castration and/or treatment with anti-androgens) which will lead to regression of prostate tumors (151).

A strategy consists in the modulation of the testosterone biosynthesis via interference with LH and LHRH. This can be achieved in two ways: employment of LH agonists to produce in the long term a downregulation of the LH receptor thus resulting in a decrease of the testosterone biosynthesis (152); employment of LH antagonists which result in a rapid decrease of testosterone levels (152). Another strategy consists of treatment with anti-androgens such as bicalutamide and enzalutamide (153).

Despite these therapies, 30-70% of the patients treated with androgen deprivation therapy will inevitably display increased PSA levels, acquire resistance to androgen suppression and develop incurable metastatic disease (154). This situation is commonly defined as castration resistant prostate cancer (CRPC) or hormone refractory prostate cancer (HRPC). Although similar, these two terms refer actually to two different clinical situations. Patients who are traditionally identified as HRPC are highly heterogeneous depending on: 1) the clinical status, 2) the level of PSA, 3) the applicability of hormone therapy and 4) the eventual presence of metastasis (95).



Disease Progression

Figure 6. Overview of prostate cancer progression combined with diagnosis and treatment options. Prostate cancer is initially treated with prostatectomy or radiotherapy and in almost 80% of the cases, patients will be cured. In 20-30% of the cases, prostate cancer relapses and these patients will be typically treated with androgen deprivation therapy. However, the development of castration resistance prostate cancer (CRPC) will inevitably occur. Although these patients will be treated with therapies such as docetaxel, cabazitaxel, enzalutamide and abiraterone, the development of incurable metastasis, typically in the bone will occur.

Interestingly, there are documented cases in which the androgen receptor (AR) signaling remains active after androgen deprivation therapy probably through escape mechanisms (30). Such mechanisms include amplification of the AR gene (155-157), gain-of-function mutations of AR (158-162), expression of alternative splice variants (163-165) and endogenous expression of enzymes involved in DHT synthesis by tumor tissue (166-169). For this reason, the term CRPC has been progressively introduced into

the clinic to indicate a condition where response to hormonal therapy is still possible, therefore reveals a different condition from HRPC (95).

Once that tumor acquires resistance to androgen suppression and patients develop metastasis mainly in the bones, treatment options are limited and include symptomatic care with analgesics or radiotherapy to reduce bone pain, treatment with bone-seeking isotopes (e.g. Strontium-89 and the recently FDA-approved Radium-223 chloride) and chemotherapy (170). Typical therapeutic treatments consist of agents targeting the androgen pathway (abiraterone acetate and enzalutamide) and taxanes (docetaxel and cabazitaxel), which target microtubules and result in the arrest of the cell cycle (170-174). Current first-line treatments consist of combination therapy with docetaxel and prednisone, while second line combination treatments are cabazitaxel and prednisone, abiraterone acetate and enzalutamide (171-174). Recent studies revealed that simultaneous treatment of ADT and docetaxel significantly increases patient survival (175,176). However, longer follow up of these studies is needed to assess whether this benefit translates also into metastatic-free survival.

4.7. Prostate Cancer Stem Cells

According to what is commonly known as the "cancer stem cell hypothesis", CSCs appear to be strongly involved in tumor formation, therapy resistance, recurrence and metastasis. As we already mentioned in a previous paragraph, cancer is a disease that originates from a single normal cell after a series of specific genomic and non-genomic alterations. As a result it was hypothesized that cells with self-renewal ability represent good candidates for oncogenic transformation and cancer formation (47). There are two putative sources of cells with self-renewal properties that are believed to generate cancer: adult stem cells (SCs) and non-stem cells that acquire self-renewal properties after de-differentiation and transformation. The majority of prostate cancer have a luminal phenotype and the absence of basal cells is a diagnostic feature of prostate adenocarcinoma (91,177). One could, therefore, speculate that prostate tumors originate from luminal progenitor cells or stem cells within the basal layer that after transformation differentiate into a luminal progeny. However, the histological compartment where the putative cell of origin of prostate cancer resides is still under debate. In hormone-naïve cancer, experimental evidence in rodents and humans support the existence of cells with self-renewal properties and tumorigenic ability in the basal compartment of the prostate ($\alpha_2\beta_1$ integrin^{hi} and CD133⁺ cells (178,179)) (**Fig. 7**). Other common markers include ALDH^{high}, CD44⁺ and CD24⁻ (39,40,180). On the other hand, in CRPC, cells with self-renewal properties exhibit a luminal phenotype. These cells have been identified in castrated mice and are known as castration-resistant Nkx3.1-expressing cells, named CARNS (33) and later the stem-like cells with luminal

phenotype (CARN-like cells) were also identified in humans (34). CARNs are characterized by low AR expression, and display the stem-cell marker ALDH1A1 or NANOG and express the luminal marker NKX3.1 and CK18 (34). Interestingly, the experimental observation in favour of the luminal hypothesis suggest the presence of a residual and dormant subpopulation of cancer cells which are castration-resistant for survival but castration-sensitive for growth (34). Recently the field has been additionally complicated by the experimental evidence that murine luminal (CD49f positive (181)) and basal (CD49 positive (182)) cells and human luminal (CD26 positive (182)) and basal (CD49f positive (182)) cells are capable of generating prostate organoids (183). However, the debate about the localization of the cell of origin of human prostate cancer and its role in the progression to a castration resistant phase is still controversial.



Figure 7. Hierachical model of tumorigenesis: role of normal and transformed tissue stem/progenitor cells. Cells within the different epithelial compartments can be distinguished by their phenotypic characteristics.

In addition to haematological malignancies, the presence of a subpopulation of epithelial cells with self-renewal properties is generally recognized in solid tumors including those of the human prostate. Furthermore accumulating experimental and clinical evidence suggest that such cells are highly tumorigenic and may play a key role in distant metastasis in preclinical models (33,178,183) and in clinical reality (184,185). The assumed role of CSCs in tumor maintenance represents one of the major problems for the identification of new, targeted therapies capable of eradicating the disease. Current therapies are indeed very effective in the treatment of the primary tumor mass (186). However, the relapse that is commonly observed (even after many years) in patients, suggests the presence of subpopulation of cells, resistant to therapy, which probably remain dormant for long time and that are capable of producing a new tumor (186,187).

5. Pathways Involved in Prostate Cancer Progression and Bone Metastasis

The notion that genes involved in developmental process are also likely to be altered in cancer is known and established. Molecular analysis revealed that a wide range of genes, commonly expressed during prostate organogenesis and developmental processes, are also abnormally expressed in prostate cancer.

Wnt pathway: the Wnt signaling pathway is an evolutionarily conserved pathway that regulates crucial aspects of development and cell behaviour, such as differentiation, migration and cell polarity. The Wnt signaling is characterized by two branches: a canonical pathway (Wnt/ β -catenin dependent) and a non-canonical pathway (β -catenin independent).

The **canonical Wnt signaling** is activated upon the interaction between a ligand (Wnt) and its receptor (Frizzled, Fz) and co-receptor (low-density-lipoprotein-related protein 5/6, LRP5 and LRP6) (**Fig. 8A**).



Figure 8. Schematic representation of canonical (A) and non-canonical (B) Wnt signaling.

In the absence of Wnt, a complex of Axin, APC, GSK3- β , CK1 and β -catenin is localized in the cytoplasm. CK1 and GSK3- β phosphorylate β -catenin which is subsequently degraded by the proteasomal machinery. In presence of Wnt, LRP6 is phosphorylated by CK1 and GSK3- β , thus recruiting to the plasma membrane a complex containing Axin and Dishevelled (Dsh), which is sequestered or degraded. This results in stabilization of β -catenin which subsequently translocates into the nucleus and mediates transcription of downstream target genes via interaction with LEF/TCF family members. Canonical Wnt signaling regulates process such as cell fate decision and anteroposterior organization in embryogenesis, as well as important function in organogenesis and stem cell renewal.

Many studies have documented alterations of the Wnt signaling pathway during prostate cancer progression, reviewed in (30,188-191). More specifically, elevated canonical Wnt signaling seems to play a role in the onset of castration resistance in prostate cancer (192). Additionally, alterations or interferences with canonical Wnt signaling, such as modulation of DKK (142) or mutation in sclerostin (SOST), which inhibits LRP5, contribute to disrupt bone formation, a process also regulated by Wnt signaling (193). In addition to the well-established effects of Wnt-signaling on enhanced osteogenesis, Wnt-signaling also induces bone-active factors, such as OPG which prevents the binding of RANKL to RANK thereby inhibiting osteoclast function (194).

The **non-canonical Wnt signaling**, comprise two branches of signaling transduction: the Wnt/Ca²⁺ signaling and the Wnt/planar cell polarity (PCP) pathway (195) (**Fig. 8B**). In Wnt/Ca²⁺ signaling, the interaction between Wnt and Fz activates phospholipase C via G proteins and lead to increase intracellular Ca²⁺. This can induce, for example, EMT and invasion, therefore promoting cancer progression. In Wnt/PCP, the non-canonical Wnt (Wnt5a and Wnt11) bind their receptor Fz which recruits Dsh at the plasma membrane. This lead to a cascade of interactions which converge on common regulators of cytoskeletal remodelling and actin organization such as RhoA, Rac1 and JNK, which also impact on cell motility.

Wnt/PCP and canonical-Wnt signaling are both part of a negative feedback-loop where Wnt/PCP negatively regulates canonical-Wnt signaling and *vice versa* (196). In cancer, due to aberrant alterations in tumor cells, cancer cells can escape from these control mechanisms and as tumors progress, Wnt/PCP gets activated and promote cell motility, invasion and metastasis (195). Interestingly, β -catenin and GSK3- β have indeed been shown to be decreased in prostate cancer cell lines with high invasive and metastatic potential, such as PC3 (197). Therefore one could speculate that there is a misbalance in these cells between canonical and non-canonical Wnt signaling which results in mesenchymal phenotype and invasive properties. **Notch pathway:** the Notch signaling pathways exerts also a crucial role during embryogenesis and organogenesis. In cancer, an aberrant activation of this pathway produces abnormal cell proliferation, increase in self-renewal properties and induction of therapy resistance (198,199). Conversely from Wnt signaling, the Notch signaling pathways requires a direct cell-to-cell contact for its activation. Typically, a signalsending cell expressing on its plasma membrane the ligand (JAG1/2 or Delta-like 1, 3 and 4, in mammals) stimulates a signal-receiving cell expressing on its membrane the receptor (Notch1/2/3/4). This interaction produces a series of proteolytic cleavages operated by ADAM10 and γ -secretases that convert the full-length transmembrane Notch receptor into a transcriptional activator (Notch intracellular domain, NICD). Subsequently, NICD translocates into the nucleus, where it interacts with RBPjk/CBP transcription factors, resulting in the transcription of downstream target genes (e.g. Hairy and enhancer of split, HES and Hairy/enhancer-of-split related with YRPW motif, HEY) (**Fig. 9**).



Figure 9. Schematic representation of Notch signaling.

Notably, Notch plays a crucial role during prostate organogenesis and is involved in its regeneration (198). Importantly, and relevant for the purpose of this thesis, the Notch

signaling pathway is characterized by multiple cross-talk with other major signaling pathways involved in prostate cancer progression and bone metastasis formation (e.g. TGF- β , AR and PI3K/AKT) (**Fig. 9**) (200-205). Members of TGF- β superfamily can control Notch signaling: for example, Bone morphogenetic protein 7 (BMP7) can inhibit the branching morphogenesis of the prostate during development via down-regulation of Notch signaling (206). Notch can also suppress AR signaling which is crucially involved in prostate growth and disease. Upon binding of androgen to the AR, the receptor undergoes a homodimerization and traslocates into the nucleus where it can recruit coactivators such as p300/CBP and steroid receptor coactivator 1 (SRC1). The downstream target of Notch, HEY1 can directly bind the N-terminal activation domain of AR thus preventing and rogen signaling, supporting a role of Notch in the acquisition of a castration resistant phenotype (204). Finally, Notch can also suppress the PI3K/AKT pathway that is fundamental for prostate growth and cell migration (207). The activation of this pathway triggers a cascade of sequential phosphorylation that can be suppressed by PTEN (198). It appears that NICD contributes to induction of PTEN expression, therefore suppressing indirectly PI3K/AKT pathway (208). This has led to the paradox that Notch signaling (particularly when triggered by Notch1) can exert a tumor suppressive role in the prostate. The complexity of the interaction between the Notch signaling and AR and PI3K/AKT pathways is further increased by a reciprocal feedback mechanism between PI3K/AKT and AR signaling: recently it was indeed demonstrated that inactivation of PI3K/AKT induces activity in AR signaling, while suppression of AR pathway induces increase in PI3K/AKT (209). Given the established increased in PI3K/AKT in advanced prostate cancer (59,210), one could speculate that an increase in Notch signaling (as documented during prostate cancer progression), through its downstream target HEY1, produces a decrease in the AR pathway (castration resistant phase) which results in increased PI3K/AKT signaling (increase migration and metastasis). Notch signaling has also been shown to critically be involved in prostate cancer progression and bone metastasis formation and JAG1 has been found to be elevated in metastatic prostate cancer compared to primary tumor (211). Additionally, in the bone microenvironment, tumour-derived JAG1 activates the Notch pathway in osteoblasts and osteoclasts and the activation of the pathway in osteoblasts results in a growth advantage to bone metastatic tumour cells (212). Interestingly, mechanistic studies showed that the proliferative effect was dependent on osteoblast-secreted IL-6, which was transcriptionally regulated by the Notch signaling in breast cancer (213). The Notch signaling pathways represents a promising target for therapy against tumor growth (214) and bone metastasis (213). However, the presence of studies addressing the possible application of y-secretase inhibitors in breast but not prostate cancer is remarkable.

TGF-β superfamily signaling pathways: during the early phase of prostate tumor growth, TGF-β acts as tumor suppressor by reducing proliferation and inducing apoptosis (215). However, during tumor progression, TGF-β switch gains a tumor promoter role and facilitates EMT and therefore metastasis (9,216). The transforming growth factor (TGF)-β superfamily of ligands includes more than 30 factors such as Bone morphogenetic proteins (BMPs), Growth and differentiation factors (GDFs), activins, inhibins, nodal and Anti-müllerian hormone (AMH). For the purpose of this thesis we will mainly focus on TGF-β members and BMPs. TGF-β is a pleiotropic cytokine that regulates many biological processes such as tissue growth and morphogenesis, cell proliferation and apoptosis, adhesion, differentiation, migration and metastasis (217). The TGF-β cytokine family consists in different members (TGF-β1, β2 and β3) whose bioactive cytokine molecule is a dimer consisting in a polypeptide chain which is cleaved from a latent precursor into the biologically active product (218).

BMPs include approximately twenty members and these are less homologous compared to the TGF- β isoforms (219). They are functionally involved in skeletal and joint morphogenesis, bone remodeling and in different cellular processes including osteogenesis, cell differentiation, anterior/posterior axis specification, growth, and homeostasis (220). In normal tissues, basal release of TGF- β by local sources is enough for the maintenance of homeostasis. In case of tissue injury, TGF- β is abundantly released by blood platelets and stromal components to prevent aberrant regenerative cell proliferation and inflammation. This occurs also in tumor microenvironment, where TGF- β is frequently present initially as factor to prevent premalignant progression, and eventually as factor that cancer cells may use to their advantage (218).

TGF- β superfamily members bind to type I and type II serine/threonine kinase receptors (**Fig. 10**). In human, seven different human type I receptors have been identified (ALK1-7) and five type II receptors, namely, TGF- β receptor II (T β RII), BMP receptor II (BMPRII), activin receptor II (ActRII), ActRIIB and AMH receptor type II. TGF- β binds T β RII and ALK5 and in endothelial cells it signals also via ALK1. BMP signaling occurs via BMPRII, ActRII and ActRIIB in association with ALK1,2,3 or 6 depending on the molecular context (221-231).

Binding of TGF- β and BMPs to their heterodimeric transmembrane receptors induces phosphorylation of type I receptor threonine/serine kinases. The signal is then transduced via Smad intracellular proteins, which later translocate into the nucleus and regulate transcription. The Smad pathway is also named **canonical signaling pathway**. TGF- β type I receptor propagates the signal by phosphorylating receptor-regulated Smad proteins (R-Smads) Smad-2 and -3. On the other hand, BMPs induce phosphorylation of Smad-1, -5 and -8 (218,232) (**Fig. 10**). This phosphorylation operated by activated type I receptor occurs at the C-terminal SXS domain that is shared by all Smad proteins and that represent a nuclear localization signal (218,233).



Figure 10. Schematic representation of the TGF- β and BMP signaling.

Depending on their phosphorylation state, Smad-2 and Smad-3 linked to Smad-4 undergo constant nucleo-cytoplasmic shuttling, in a sort of rapid activation-deactivation cycle, determined by repeated cycles of dephosphorylation and rephosphorylation, involving direct interactions with both nuclear pore proteins and importins and exportins, a protein family of transport factors (233).

TGF- β also regulates alternative pathways via Smad-independent signaling (**non-canonical Smad signaling**). These signaling include the extracellular-signal- regulated kinase (ERK1 and ERK2), p38, MAPKs, c-Jun N-terminal Kinase (JNK), PI3K-Akt and small GTPases. The non-canonical Smad signaling pathways have been extensively reviewd in (234-236).

Each step in the TGF- β signaling pathways is controlled by specialized factors. These factors include encapsulation of the extracellular ligand by binding proteins, inhibition of activation of latent TGF- β , receptor-interacting partners (BAMBI, SARA and FKBP12), inhibitory Smads (Smad6 and Smad7) and post-translational modification by E3 ubiquitin ligases, co-repressors and phosphatases, reviewed in (237).

Integrins: integrins belong to a family of heterodimeric transmembrane glycoprotein receptors which consist of an α and a β subunit and which play important roles in tissue development and cancer (238,239). To date 18 α and 8 β subunits have been identified from which 24 different functional heterodimers can be generated (240). Integrins regulate many processes such as cell adhesion, migration, proliferation, neo angiogenesis (241) and have been shown to undergo changes in their expression during the transition to neoplastic phase (242,243). Integrins establish the connection between the cell and the extracellular environment (mostly the extracellular matrix molecules) and the cytoskeleton and transduce signals from the outside and into the cells and vice versa (reviewed in (238,240,244-246)) (Fig. 11). In addition, integrins can modulate the signaling cascade of multiple growth factor receptors via RasGTP, such as the epidermal growth factor (EGF), platelet-derived growth factor (PDGF) and fibroblast growth factor (FGF) receptors, thereby lowering the threshold level in different signaling pathways (247). Moreover, ligation and clustering of integrins can lead to the activation of focal adhesion kinase (FAK) and extracellular signal-related kinase kinase (MEK) (248) which have been implicated in prostate cancer progression and metastasis (249,250). Previous studies have shown that specific integrins (such as $\alpha\nu\beta3$ (101,102)) correlate with poor survival and are involved in the formation of bone metastasis (251-254). Furthermore, targeting of α_v integrin (knockdown or selective drug targeting) in human prostate cancer cells, abolished the formation of bone metastasis in preclinical mouse model (255,256).



Figure 11. Schematic representation of the multiple roles of integrins.

Additionally, α_v integrins appear to be up-regulated in tumor- and metastasis-initiating prostate cancer cells (ALDH^{high} (39,257)) and these integrins are involved in the activation of latent TGF- β , thereby modulating TGF- β signaling (and *vice versa* in a feedforward loop (244,258-260)) (**Fig. 8**). As previously outlined, high bone turnover provides a significant contribution to the development and the relapse of bone metastasis (261). Interestingly, it has been reporter that increased expression of integrin alpha-v enhanced the TGF- β mediated osteoclastogenesis (262). Therefore it appears that changes in integrin expression play an important role in malignat disease and impact not only on primary tumor growth and invasion but also on the bone microenvironment.

Cripto pathway: Cripto (TDGF1, CRIPTO-1) is a small, GPI-anchored/secreted fetal-oncoprotein that plays important roles in regulating stem cell differentiation, embryogenesis, tissue growth and remodelling (263). An essential mediator for the Cripto signaling is the Glucose-regulated protein 78 (GRP78) (264). As for Wnt and Notch, Cripto represents one of those embryonic signaling pathways that when corrupted can drive tumor initiation and progression. The Cripto pathway modulates the signaling of multiple TGF- β ligand that transduce the signal via Smad2 and 3 such as Nodal, GDF1 and GDF3 (265-267) (Fig. 12). Interestingly, Cripto has also been shown to negatively regulate the activation of Smad by Activin-A (268,269), Activin-B (270) and TGFB-1 (269,271) leading to suppression of the cytostatic effect of these ligands (271,272). These cross-talk with the TGF β pathway, also crucially involved in prostate cancer bone metastasis, highlight the interest of elucidating the role of Cripto signaling in the contest of bone metastasis. Additionally, even though the soluble vs. secreted effects regulated by Cripto are not yet been entirely elucidated (273), Cripto has also signaling activities that are independent from the TGF- β pathway. Relevant for the purpose of this thesis, soluble Cripto can activate and promote signaling routes of extreme relevance in prostate cancer formation and progression, such as the already discussed PI3K/AKT. In this context, blockade of Cripto binding to cell surface GRP78 inhibits oncogenic Cripto signaling via MAPK/PI3K and Smad2/3 signaling routes (270). Moreover, Cripto is also known to modulate Wnt and Notch signaling pathways (274-277). The interaction between Cripto and PI3K/AKT pathway is mediated via Glypican-1, a GPI-anchored heparan sulphate proteoglycan, that activates a cascade of phosphorylation in which MAP Kinase are involved. This lead to the subsequent activation of PI3K/AKT pathway which promotes proliferation and motility (278). Cripto has also been shown to crosstalk with Wnt signaling; it can bind LRP5/6 facilitating the interaction with Wnt3a, therefore stimulating Wnt pathway through cytoplasmic stabilization of β -catenin (279). Cripto is also involved in the processing of the Notch receptors by enhancing its

cleavage from the plasma membrane, thereby potentiating Notch signaling (277). Finally, Notch signaling can also modulate the expression of Nodal, further complicating the cross-talk between Notch and Cripto /Nodal signaling (280). As we previously mentioned, one of the key processes that characterize the switch from non-invasive to invasive disease in prostate cancer is represented by EMT. Interestingly, Cripto exerts an important role in this process in prostate cancer, where its overexpression produces increase in the mesenchymal marker Vimentine, decrease in the epithelial marker E-Cadherin and augment PI3K/AKT and FGFR1 activity, thus inducing migration (281).



Figure 12. Schematic representation of the multiple interactions of Cripto signaling.

The genetic alterations and the signaling pathways discussed in this paragraph obviously do not cover the entire complexity of aberrant genetic events and abnormalities that characterize multiple pathways and molecules from the onset to the progression of prostate cancer. Among these, we have focused our interest on the alteration and the role of a class of small non coding RNA, namely microRNA, that regulate gene expression. The properties of these molecules, the mechanism of action and their functional relevance in the maintenance of aggressive subpopulation of cancer cells during prostate cancer progression and metastasis are presented in the next paragraphs.

5.1. The involvement of microRNA in prostate cancer

microRNAs (miRs) are a class of small non-coding RNAs that derive from larger precursor (pri-miRNA) folded into a stem-loop configuration. miRs are transcribed by RNA polymerase II (Pol II) and subsequently processed into the ~70-nucleotide precursors (pre-miR) (282,283). The pre-miR is then cleaved to generate a ~21-25-nucleotide mature miR. miRs localized within Alu-repetitive element, can be transcribed by RNA polymerase III (284). miRs can be positioned at different genomic locations; for example, they can map within introns of both protein-coding or non-coding genes (285). These are transcriptionally regulated through the promoters of these genes (286,287). The transcription of miRs held in the same cluster is regulated by the same promoter and all the miRs from that cluster are transcribed at the same time.

The processing of miR is catalyzed by different multiprotein complexes (**Fig. 13**) (reviewed in (288)). A complex localized in the nucleus and composed by an RNase III enzyme Drosha and the double-stranded RNA-binding domain (dsRBD) protein DGCR8/Pasha, process the pri-miR (289). This enzymatic reaction produces a 2-nucleotide-long 3' overhangs at the cleavage site. The processing of the pri-miR into ~70-bp pre-miRs by Drosha depends on the terminal loop size and the flanking sequence of the Drosha cleavage site. Shortening of the terminal loop, disruption of complementariety within the sequence, or mutations of flanking sequence at the Drosha cleavage site, can significantly reduce, if not abolish the processing of the pri-miR.

After the pri-miR is cleaved by Drosha, the resuting pre-miR is exported from the nucleus into the cytoplasm by Exportin 5 (Exp5), a nucleo/cytoplasmic cargo transporter Ran-GTP dependent (290-292). In the cytoplasm, another RNase III enzyme (Dicer) cleaves the hairpin into a small imperfect dsRNA duplex that contains both the mature miR strand and its complementary strand (293-295). The ability of Dicer to recognize the pre-miR molecules is due to the presence of a PAZ (Piwi-Argonaute-Zwille) domain that allows a low-affinity interaction with the 3' end of ssRNAs (296-298). For this reason, the pre-miR that presents 2-nucleotide 3' overhangs resulting from Drosha cleavage, can be easily recognized and processed by Dicer.

Dicer cleavage generates mature miRs ~21-25-nucleotide long. After the dsRNA duplex is formed, the target specificity and the functional efficiency of a miR, requires that the mature miR strand is incorporated into the RNA-induced silencing complex (RISC) (reviewed in (288)). In human cells, transactivating response (TAR) RNA-binding protein (TRBP), recruits the Argonaute protein Ago2 and together with Dicer they form a trimeric complex that initiates the assembly of the RISC complex (reviewed in (288)).

The mechanism by which the RISC complex incorporates the mature miR strand of the dsRNA duplex is driven by the different stability of the miR duplex. Potentially the

mature miR strand can reside on either strand of the hairpin, but, because of thermodynamic reasons, it mostly derives from the strand with the less stable 5'.



Figure 13. Schematic representation of the microRNA processing pathway.

The detection of microRNA in blood and urines represent an interesting and noninvasive approach to diagnose prostate cancer (299). Independent studies have shown that miR-141 and miR-375 are significantly elevated in the blood of prostate cancer patients with bone metastasis and in the respective exosomes (300,301). Interestingly elevated levels of the two microRNAs are also associated with higher Gleason score, positive lymph nodes and were also detected in the urine of prostate cancer patients (302,303). miR-375 has also been identified as prognostic marker in castrationresistance prostate cancer together with miR-1290 in exosomes (304). Additional microRNA that have been measured in the urine and associated with prostate cancer are miR-107, miR-574-3p and miR-200b (303). Interestingly, the last has also been associated by an independent study with docetaxel resistance (305). In the same study, miR-429, miR-200a, miR-21, miR-200c, miR-375, miR-132 and miR-20 have been associated with lower survival (305). A recent report investigated the expression of microRNAs in patient-derived stem like cells (CD133⁺, $\alpha 2\beta 1$ integrin^{high}) enriched from benign prostatic hyperplasia, Gleason 7 treatment-naive prostate cancer, and CRPC and identified miR-548c-3p as functional biomarker involved in prostate cancer progression (306).

5.2. Mechanisms of microRNA Post-Transcriptional Repression

After incorporation into the RISC complex, the miR interact with its target mRNA by base-pairing interactions. If mRNA/miR complementarity is perfect or near-perfect, the target mRNA can be cleaved and degraded; otherwise the translation is repressed (294).

The target complementarity is determined by base-pairing of nucleotides in the so called "seed sequence" of the miR (307). This sequence is essential for the binding of the miR to the mRNA. The seed sequence is an heptametrical sequence located at positions 2-7 from the miR 5⁻ end and has to be perfectly complementary to the target mRNA complementary sequence. The miR seed sequence is exploited to develop computational approaches for target prediction.

The microRNA target site is positioned at the 3'UTR region, probably because the movement of ribosomes that occur during translation will contrast RISC binding and interaction (308). Different and "non-canonical" miR-mediated mechanisms of mRNA expression modulation are also emerging. In fact some miRs can bind to the open reading frame (ORF) sequences or to the 5'UTR region of the target genes, determining gene activation rather than repression (309). The RISC action on target mRNA is modulated by the Ago protein that is incorporated in the complex and by the grade of complementarity between the miR strand and its mRNA target. Ago2, for example, is able to cleave RNA, but this event requires extensive base pairing between the miR strand and the mRNA target (310,311).

To date, six models have been proposed for the miR translational repression:

- 1) the RISC complex induces de-adenylation determining a decrease of translational efficiency by blocking target mRNA circularization (312);
- RISC complex blocks cap function by interacting with both the cap or eIF4E (313);
- 3) Argonaute proteins recruit eIF6, which blocks the recruitment of 60s ribosomal subunit (314);
- 4) RISC complex blocks the translation elongation or promotes premature dissociation of ribosomes (ribosome drop-off) (315);
- 5) RISC complex induces the proteolysis of nascent peptides during translation (in this model the translation is not inhibited) (316);

6) RISC complex recruits target mRNAs to processing bodies, where the mRNA is degraded or stored in an inactive state for translation (317,318).

5.3. microRNA and Cancer Stem Cells

microRNAs regulate multiple biological process, such development and cell growth and have been proposed as one of the important players during pathogenesis and cancer. In human prostate cancer, several studies in patient samples and xenografts have revealed their characteristic pattern in benign vs. aggressive disease and highlighted their role in castration-resistant prostate cancer and their implication in bone metastasis formation (see microRNAs described in paragraph 3 and reviewed in (299,300,302)).

However, the number of studies addressing the role of specific miRs in the regulation of stem-like properties in bulk prostate cancer cells lines is limited. Remarkably, there is an even lower number of studies, that investigated the expression of miRs directly in selected subpopulation of cells, characterized by stem-like properties and capable of maintaining the tumor and producing metastasis. In these studies new miRs have been identified using expression profiling of subpopulation of cells enriched for cancer stem cells isolated from bulk prostate cancer cell lines. For example, prostaspheres from PC3 cells have been compared to adherent PC3 cells and miR-143 has been identified as promoter of prostate cancer metastasis (319). In other approaches, a fraction of prostate cancer stem cells (PCSCs, described as CD44⁺/CD133⁺) has been isolated by viable cell sorting from culturedLNCaP cells and miR-101 has been found to inhibit cell growth and promote apoptosis in PCSCs (320). With a similar approach, miR-409-3p/5p has been identified in embryonic stem cells and then studied in prostate cancer, where it was found to promote bone metastasis (321). Other studies have also employed cell sorting of various stem/progenitor cell population, including CD44⁺, CD133⁺, integrin $\alpha 2\beta 1^+$ and side population of cells isolate from bulk cell lines and found multiple tumor-suppressive microRNA down-regulated, including miR34a, let-7, miR-106a and miR141 (322). Interestingly, a 'near-patient' approach highlights that a population of CD44⁺ cells isolated from xenografts led to the identification of miR-34a as master regulator of metastasis (323) and, as a consequence, this was subsequently validated in CD44⁺ cells isolated from primary prostate tumors. Moreover, in patientderived stem like cells (CD133⁺, $\alpha 2\beta 1^{high}$), it was recently found that miR-548c-3p can be considered as functional biomarker involved in prostate cancer progression (306) as enforced overexpression of this miR in differentiated cells induced stem-like properties and radioresistance. Finally other studies have used Hoechst 33342-based flow cytometry to isolate a CSC-like side population and confirmed the tumor suppressive role of miR-34a and additionally identified miR-200c as mediator of chemoresistance.

Strikingly, there is lack of microRNA expression profiles of cancer-stem like/progenitor cells obtained from clinical prostate cancer specimens. The molecular characterization of this subpopulation of highly tumorigenic cells, could indeed provide novel insights in tumor progression and facilitate the identification of new therapeutic targets and strategies.

6. Outline of the thesis

Cancer is a heterogeneous disease and the presence of multiple genetically distinct foci in the primary prostate cancer supports this notion. The identification of the molecular properties of highly aggressive and metastatic subclones might facilitate the identification of new targets for therapy and putative markers for monitoring the progression of the disease.

In **Chapter 2** of this thesis, we established a microRNA signature common to three key signaling pathways in prostate cancer progression and bone metastasis formation (i.e. TGF- β , Wnt and Notch). With this approach we identified a signature of validated microRNA targeting the process of epithelial-to-mesenchymal transition (EMT) that may be critically involved in the spreading of many aggressive cells from the primary tumor and the formation of distant metastases.

Chapter 3 is focused on a candidate tumor suppressor microRNA that is downregulated in highly metastatic, stem-like ALDH^{high} cells vs. non-metastatic, more differentiated ALDH^{low} prostate cancer cells. We studied the functional role of miR-25 in the maintenance of aggressive behaviour of ALDH^{high} compared to ALDH^{low} subpopulation of cells *in vitro* and *in vivo*. Our analysis revealed that miR-25 represents an important player in the regulation of invasiveness in human prostate cancer through the interaction with at least three signaling pathways.

Chapter 4 describes a follow-up study for the regulatory role of miR-25 in human prostate cancer biology, in particular its role in the cross-talk between the TGF- β and Wnt signalling in prostate carcinogenesis and progression.

In **Chapter 5** we show that the soluble chimeric protein ALK1Fc reduces BMP-9 induced activation of Notch signaling and proliferation in human prostate cancer cells. Alk1Fc is capable of reducing tumor growth in an orthotopic model of human prostate cancer *in vivo*.

Chapter 6 contains a study about the role of Cripto and GRP78 in the maintenance of an aggressive behaviour in human prostate cancer cells *in vitro* and their role in metastatic dissemination in two different preclinical models of prostate cancer invasion and metastasis *in vivo*. The general conclusions are included in **Chapter 7**, the general discussion of this thesis.

REFERENCES

- 1. 2012 Cancer Statistics. http://www.cancerresearchuk.org/health-professional/cancer-statistics/worldwide-cancer#heading-Zero.
- 2. Marahatta SB, Sharma N, Koju R, Makaju RK, Petmitr P, Petmitr S. Cancer: determinants and progression. Nepal Med Coll J 2005;7(1):65-71.
- 3. Coleman WB, Tsongalis GJ. Molecular mechanisms of human carcinogenesis. EXS 2006(96):321-49.
- 4. Hanahan D, Weinberg RA. The hallmarks of cancer. Cell 2000;100(1):57-70.
- 5. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell 2011;144(5):646-74.
- 6. Pietras K, Ostman A. Hallmarks of cancer: interactions with the tumor stroma. Exp Cell Res 2010;316(8):1324-31.
- 7. Bhowmick NA, Neilson EG, Moses HL. Stromal fibroblasts in cancer initiation and progression. Nature 2004;432(7015):332-7.
- 8. Folberg R, Maniotis AJ. Vasculogenic mimicry. APMIS 2004;112(7-8):508-25.
- 9. Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelial-mesenchymal transitions in development and disease. Cell 2009;139(5):871-90.
- 10. Friedl P, Locker J, Sahai E, Segall JE. Classifying collective cancer cell invasion. Nat Cell Biol 2012;14(8):777-83.
- 11. Warburg O. On respiratory impairment in cancer cells. Science 1956;124(3215):269-70.
- 12. Ward PS, Thompson CB. Metabolic reprogramming: a cancer hallmark even warburg did not anticipate. Cancer Cell 2012;21(3):297-308.
- 13. Dang L, White DW, Gross S, Bennett BD, Bittinger MA, Driggers EM, et al. Cancer-associated IDH1 mutations produce 2-hydroxyglutarate. Nature 2010;465(7300):966.
- 14. DeNardo DG, Andreu P, Coussens LM. Interactions between lymphocytes and myeloid cells regulate pro- versus anti-tumor immunity. Cancer Metastasis Rev 2010;29(2):309-16.
- 15. Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. Cell 2010;140(6):883-99.
- 16. Mougiakakos D, Choudhury A, Lladser A, Kiessling R, Johansson CC. Regulatory T cells in cancer. Adv Cancer Res 2010;107:57-117.
- 17. Ostrand-Rosenberg S, Sinha P. Myeloid-derived suppressor cells: linking inflammation and cancer. J Immunol 2009;182(8):4499-506.
- 18. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. Cell 2010;141(1):39-51.
- 19. Marusyk A, Almendro V, Polyak K. Intra-tumour heterogeneity: a looking glass for cancer? Nat Rev Cancer 2012;12(5):323-34.
- 20. Leissner KH, Tisell LE. The weight of the human prostate. Scand J Urol Nephrol 1979;13(2):137-42.
- 21. Owen DH, Katz DF. A review of the physical and chemical properties of human semen and the formulation of a semen simulant. J Androl 2005;26(4):459-69.
- 22. McNeal JE. Normal and pathologic anatomy of prostate. Urology 1981;17(Suppl 3):11-6.
- 23. McNeal JE. The zonal anatomy of the prostate. Prostate 1981;2(1):35-49.
- 24. Cohen RJ, Shannon BA, Phillips M, Moorin RE, Wheeler TM, Garrett KL. Central zone carcinoma of the prostate gland: a distinct tumor type with poor prognostic features. J Urol 2008;179(5):1762-7; discussion 67.
- 25. Lepor H. Pathophysiology, epidemiology, and natural history of benign prostatic hyperplasia. Rev Urol 2004;6 Suppl 9:S3-S10.
- 26. In: Liverman CT, Blazer DG, editors. Testosterone and Aging: Clinical Research Directions. Washington (DC)2004.
- 27. Marchetti PM, Barth JH. Clinical biochemistry of dihydrotestosterone. Ann Clin Biochem 2013;50(Pt 2):95-107.

- 28. Powell SM, Christiaens V, Voulgaraki D, Waxman J, Claessens F, Bevan CL. Mechanisms of androgen receptor signalling via steroid receptor coactivator-1 in prostate. Endocr Relat Cancer 2004;11(1):117-30.
- 29. Heinlein CA, Chang C. Androgen receptor (AR) coregulators: an overview. Endocr Rev 2002;23(2):175-200.
- 30. Shen MM, Abate-Shen C. Molecular genetics of prostate cancer: new prospects for old challenges. Genes Dev 2010;24(18):1967-2000.
- 31. Brawer MK, Peehl DM, Stamey TA, Bostwick DG. Keratin immunoreactivity in the benign and neoplastic human prostate. Cancer Res 1985;45(8):3663-7.
- 32. Hudson DL, Guy AT, Fry P, O'Hare MJ, Watt FM, Masters JR. Epithelial cell differentiation pathways in the human prostate: identification of intermediate phenotypes by keratin expression. J Histochem Cytochem 2001;49(2):271-8.
- 33. Wang X, Kruithof-de Julio M, Economides KD, Walker D, Yu H, Halili MV, et al. A luminal epithelial stem cell that is a cell of origin for prostate cancer. Nature 2009;461(7263):495-500.
- Germann M, Wetterwald A, Guzman-Ramirez N, van der Pluijm G, Culig Z, Cecchini MG, et al. Stem-like cells with luminal progenitor phenotype survive castration in human prostate cancer. Stem Cells 2012;30(6):1076-86.
- 35. Arnold JT, Isaacs JT. Mechanisms involved in the progression of androgen-independent prostate cancers: it is not only the cancer cell's fault. Endocr Relat Cancer 2002;9(1):61-73.
- 36. Tuxhorn JA, Ayala GE, Smith MJ, Smith VC, Dang TD, Rowley DR. Reactive stroma in human prostate cancer: induction of myofibroblast phenotype and extracellular matrix remodeling. Clin Cancer Res 2002;8(9):2912-23.
- 37. Nagle RB, Ahmann FR, McDaniel KM, Paquin ML, Clark VA, Celniker A. Cytokeratin characterization of human prostatic carcinoma and its derived cell lines. Cancer Res 1987;47(1):281-6.
- 38. Bonkhoff H, Stein U, Remberger K. The proliferative function of basal cells in the normal and hyperplastic human prostate. Prostate 1994;24(3):114-8.
- van den Hoogen C, van der Horst G, Cheung H, Buijs JT, Lippitt JM, Guzman-Ramirez N, et al. High aldehyde dehydrogenase activity identifies tumor-initiating and metastasis-initiating cells in human prostate cancer. Cancer Res 2010;70(12):5163-73.
- 40. Collins AT, Berry PA, Hyde C, Stower MJ, Maitland NJ. Prospective identification of tumorigenic prostate cancer stem cells. Cancer Res 2005;65(23):10946-51.
- 41. Lawson DA, Witte ON. Stem cells in prostate cancer initiation and progression. J Clin Invest 2007;117(8):2044-50.
- 42. Taylor RA, Cowin PA, Cunha GR, Pera M, Trounson AO, Pedersen J, et al. Formation of human prostate tissue from embryonic stem cells. Nat Methods 2006;3(3):179-81.
- 43. Noordzij MA, van Steenbrugge GJ, van der Kwast TH, Schroder FH. Neuroendocrine cells in the normal, hyperplastic and neoplastic prostate. Urol Res 1995;22(6):333-41.
- 44. Sciarra A, Mariotti G, Gentile V, Voria G, Pastore A, Monti S, et al. Neuroendocrine differentiation in human prostate tissue: is it detectable and treatable? BJU Int 2003;91(5):438-45.
- 45. di Sant'Agnese PA. Neuroendocrine differentiation in prostatic carcinoma: an update on recent developments. Ann Oncol 2001;12 Suppl 2:S135-40.
- 46. Potts JM. The four categories of prostatitis: a practical approach to treatment. Cleve Clin J Med 2001;68(5):389-90, 92-3, 97.
- 47. Isaacs JT, Coffey DS. Etiology and disease process of benign prostatic hyperplasia. Prostate Suppl 1989;2:33-50.
- 48. Lepor H. Medical therapy for benign prostatic hyperplasia. Urology 1993;42(5):483-501.
- 49. Elterman DS, Barkin J, Kaplan SA. Optimizing the management of benign prostatic hyperplasia. Ther Adv Urol 2012;4(2):77-83.

- 50. Bostwick DG, Cooner WH, Denis L, Jones GW, Scardino PT, Murphy GP. The association of benign prostatic hyperplasia and cancer of the prostate. Cancer 1992;70(1 Suppl):291-301.
- 51. SEER Stat Fact Sheets: Prostate Cancer. http://seer.cancer.gov/statfacts/html/prost.html.
- 52. Jemal A, Siegel R, Xu J, Ward E. Cancer statistics, 2010. CA Cancer J Clin 2010;60(5):277-300.
- 53. Yatani R, Kusano I, Shiraishi T, Hayashi T, Stemmermann GN. Latent prostatic carcinoma: pathological and epidemiological aspects. Jpn J Clin Oncol 1989;19(4):319-26.
- 54. McNeal JE, Bostwick DG. Intraductal dysplasia: a premalignant lesion of the prostate. Hum Pathol 1986;17(1):64-71.
- 55. Haggman MJ, Macoska JA, Wojno KJ, Oesterling JE. The relationship between prostatic intraepithelial neoplasia and prostate cancer: critical issues. J Urol 1997;158(1):12-22.
- 56. Bostwick DG. Prostatic intraepithelial neoplasia (PIN). Urology 1989;34(6 Suppl):16-22.
- 57. Dong JT. Chromosomal deletions and tumor suppressor genes in prostate cancer. Cancer Metastasis Rev 2001;20(3-4):173-93.
- 58. Lapointe J, Li C, Giacomini CP, Salari K, Huang S, Wang P, et al. Genomic profiling reveals alternative genetic pathways of prostate tumorigenesis. Cancer Res 2007;67(18):8504-10.
- 59. Taylor BS, Schultz N, Hieronymus H, Gopalan A, Xiao Y, Carver BS, et al. Integrative genomic profiling of human prostate cancer. Cancer Cell 2010;18(1):11-22.
- 60. Xu J, Zheng SL, Carpten JD, Nupponen NN, Robbins CM, Mestre J, et al. Evaluation of linkage and association of HPC2/ELAC2 in patients with familial or sporadic prostate cancer. Am J Hum Genet 2001;68(4):901-11.
- 61. Xu J, Zheng SL, Chang B, Smith JR, Carpten JD, Stine OC, et al. Linkage of prostate cancer susceptibility loci to chromosome 1. Hum Genet 2001;108(4):335-45.
- 62. Carracedo A, Pandolfi PP. The PTEN-PI3K pathway: of feedbacks and cross-talks. Oncogene 2008;27(41):5527-41.
- 63. Di Cristofano A, Pesce B, Cordon-Cardo C, Pandolfi PP. Pten is essential for embryonic development and tumour suppression. Nat Genet 1998;19(4):348-55.
- 64. Gurel B, Iwata T, Koh CM, Jenkins RB, Lan F, Van Dang C, et al. Nuclear MYC protein overexpression is an early alteration in human prostate carcinogenesis. Mod Pathol 2008;21(9):1156-67.
- 65. Ellwood-Yen K, Graeber TG, Wongvipat J, Iruela-Arispe ML, Zhang J, Matusik R, et al. Mycdriven murine prostate cancer shares molecular features with human prostate tumors. Cancer Cell 2003;4(3):223-38.
- Tomlins SA, Rhodes DR, Perner S, Dhanasekaran SM, Mehra R, Sun XW, et al. Recurrent fusion of TMPRSS2 and ETS transcription factor genes in prostate cancer. Science 2005;310(5748):644-8.
- 67. Schultz M, Parzinger H, Posdnjakov DV, Chikisheva TA, Schmidt-Schultz TH. Oldest known case of metastasizing prostate carcinoma diagnosed in the skeleton of a 2,700-year-old Scythian king from Arzhan (Siberia, Russia). Int J Cancer 2007;121(12):2591-5.
- Schmidt-Schultz TH, Schultz M. Bone protects proteins over thousands of years: extraction, analysis, and interpretation of extracellular matrix proteins in archeological skeletal remains. Am J Phys Anthropol 2004;123(1):30-9.
- 69. Schmidt-Schultz TH, Schultz M. Intact growth factors are conserved in the extracellular matrix of ancient human bone and teeth: a storehouse for the study of human evolution in health and disease. Biol Chem 2005;386(8):767-76.
- 70. Lilja H, Ulmert D, Vickers AJ. Prostate-specific antigen and prostate cancer: prediction, detection and monitoring. Nat Rev Cancer 2008;8(4):268-78.
- 71. Bussemakers MJ, van Bokhoven A, Verhaegh GW, Smit FP, Karthaus HF, Schalken JA, et al. DD3: a new prostate-specific gene, highly overexpressed in prostate cancer. Cancer Res 1999;59(23):5975-9.
- 72. Dijkstra S, Mulders PF, Schalken JA. Clinical use of novel urine and blood based prostate cancer biomarkers: a review. Clin Biochem 2014;47(10-11):889-96.

- 73. Savblom C, Malm J, Giwercman A, Nilsson JA, Berglund G, Lilja H. Blood levels of free-PSA but not complex-PSA significantly correlates to prostate release of PSA in semen in young men, while blood levels of complex-PSA, but not free-PSA increase with age. Prostate 2005;65(1):66-72.
- 74. Qiu SD, Young CY, Bilhartz DL, Prescott JL, Farrow GM, He WW, et al. In situ hybridization of prostate-specific antigen mRNA in human prostate. J Urol 1990;144(6):1550-6.
- 75. Epstein JI, Allsbrook WC, Jr., Amin MB, Egevad LL, Committee IG. The 2005 International Society of Urological Pathology (ISUP) Consensus Conference on Gleason Grading of Prostatic Carcinoma. Am J Surg Pathol 2005;29(9):1228-42.
- 76. Mellinger GT, Gleason D, Bailar J, 3rd. The histology and prognosis of prostatic cancer. J Urol 1967;97(2):331-7.
- 77. Gleason DF, Mellinger GT. Prediction of prognosis for prostatic adenocarcinoma by combined histological grading and clinical staging. J Urol 1974;111(1):58-64.
- 78. Epstein JI. Update on the Gleason grading system. Ann Pathol 2011;31(5 Suppl):S20-6.
- 79. Humphrey PA. Gleason grading and prognostic factors in carcinoma of the prostate. Mod Pathol 2004;17(3):292-306.
- 80. Ohori M, Wheeler TM, Scardino PT. The New American Joint Committee on Cancer and International Union Against Cancer TNM classification of prostate cancer. Clinicopathologic correlations. Cancer 1994;74(1):104-14.
- 81. Godtman RA, Holmberg E, Khatami A, Stranne J, Hugosson J. Outcome following active surveillance of men with screen-detected prostate cancer. Results from the Goteborg randomised population-based prostate cancer screening trial. Eur Urol 2013;63(1):101-7.
- 82. Hayes JH, Ollendorf DA, Pearson SD, Barry MJ, Kantoff PW, Lee PA, et al. Observation versus initial treatment for men with localized, low-risk prostate cancer: a cost-effectiveness analysis. Ann Intern Med 2013;158(12):853-60.
- 83. Lund L, Svolgaard N, Poulsen MH. Prostate cancer: a review of active surveillance. Res Rep Urol 2014;6:107-12.
- 84. Cooperberg MR, Carroll PR, Klotz L. Active surveillance for prostate cancer: progress and promise. J Clin Oncol 2011;29(27):3669-76.
- 85. Adolfsson J. Watchful waiting and active surveillance: the current position. BJU Int 2008;102(1):10-4.
- 86. Bianco FJ, Jr., Scardino PT, Eastham JA. Radical prostatectomy: long-term cancer control and recovery of sexual and urinary function ("trifecta"). Urology 2005;66(5 Suppl):83-94.
- 87. Heidenreich A, Bastian PJ, Bellmunt J, Bolla M, Joniau S, van der Kwast T, et al. EAU guidelines on prostate cancer. part 1: screening, diagnosis, and local treatment with curative intent-update 2013. Eur Urol 2014;65(1):124-37.
- 88. Allen B. Systemic targeted alpha radiotherapy for cancer. J Biomed Phys Eng 2013;3(3):67-80.
- 89. Bauman G, Rumble RB, Chen J, Loblaw A, Warde P, Members of the IIEP. Intensity-modulated radiotherapy in the treatment of prostate cancer. Clin Oncol (R Coll Radiol) 2012;24(7):461-73.
- 90. Eton DT, Lepore SJ, Helgeson VS. Early quality of life in patients with localized prostate carcinoma: an examination of treatment-related, demographic, and psychosocial factors. Cancer 2001;92(6):1451-9.
- 91. Humphrey PA. Diagnosis of adenocarcinoma in prostate needle biopsy tissue. J Clin Pathol 2007;60(1):35-42.
- 92. Taylor RA, Toivanen R, Frydenberg M, Pedersen J, Harewood L, Australian Prostate Cancer B, et al. Human epithelial basal cells are cells of origin of prostate cancer, independent of CD133 status. Stem Cells 2012;30(6):1087-96.
- Hirano D, Okada Y, Minei S, Takimoto Y, Nemoto N. Neuroendocrine differentiation in hormone refractory prostate cancer following androgen deprivation therapy. Eur Urol 2004;45(5):586-92; discussion 92.

- 94. Kokubo H, Yamada Y, Nishio Y, Fukatsu H, Honda N, Nakagawa A, et al. Immunohistochemical study of chromogranin A in Stage D2 prostate cancer. Urology 2005;66(1):135-40.
- 95. Carles J, Castellano D, Climent MA, Maroto P, Medina R, Alcaraz A. Castration-resistant metastatic prostate cancer: current status and treatment possibilities. Clin Transl Oncol 2012;14(3):169-76.
- 96. Nathoo N, Caris EC, Wiener JA, Mendel E. History of the vertebral venous plexus and the significant contributions of Breschet and Batson. Neurosurgery 2011;69(5):1007-14; discussion 14.
- 97. Dodds PR, Caride VJ, Lytton B. The role of vertebral veins in the dissemination of prostatic carcinoma. J Urol 1981;126(6):753-5.
- 98. Honn KV, Tang DG. Adhesion molecules and tumor cell interaction with endothelium and subendothelial matrix. Cancer Metastasis Rev 1992;11(3-4):353-75.
- 99. Wang J, Loberg R, Taichman RS. The pivotal role of CXCL12 (SDF-1)/CXCR4 axis in bone metastasis. Cancer Metastasis Rev 2006;25(4):573-87.
- 100. Sun YX, Schneider A, Jung Y, Wang J, Dai J, Wang J, et al. Skeletal localization and neutralization of the SDF-1(CXCL12)/CXCR4 axis blocks prostate cancer metastasis and growth in osseous sites in vivo. J Bone Miner Res 2005;20(2):318-29.
- 101. Schneider JG, Amend SR, Weilbaecher KN. Integrins and bone metastasis: integrating tumor cell and stromal cell interactions. Bone 2011;48(1):54-65.
- 102. Farias E, Lu M, Li X, Schnapp LM. Integrin alpha8beta1-fibronectin interactions promote cell survival via PI3 kinase pathway. Biochem Biophys Res Commun 2005;329(1):305-11.
- 103. Weilbaecher KN, Guise TA, McCauley LK. Cancer to bone: a fatal attraction. Nat Rev Cancer 2011;11(6):411-25.
- 104. Ghotra VP, He S, van der Horst G, Nijhoff S, de Bont H, Lekkerkerker A, et al. SYK is a candidate kinase target for the treatment of advanced prostate cancer. Cancer Res 2015;75(1):230-40.
- 105. Paget S. The distribution of secondary growths in cancer of the breast. 1889. Cancer Metastasis Rev 1989;8(2):98-101.
- 106. Ozdemir BC, Hensel J, Secondini C, Wetterwald A, Schwaninger R, Fleischmann A, et al. The molecular signature of the stroma response in prostate cancer-induced osteoblastic bone metastasis highlights expansion of hematopoietic and prostate epithelial stem cell niches. PLoS One 2014;9(12):e114530.
- 107. Kaplan RN, Rafii S, Lyden D. Preparing the "soil": the premetastatic niche. Cancer Res 2006;66(23):11089-93.
- 108. Kaplan RN, Psaila B, Lyden D. Bone marrow cells in the 'pre-metastatic niche': within bone and beyond. Cancer Metastasis Rev 2006;25(4):521-9.
- 109. Hiratsuka S, Watanabe A, Aburatani H, Maru Y. Tumour-mediated upregulation of chemoattractants and recruitment of myeloid cells predetermines lung metastasis. Nat Cell Biol 2006;8(12):1369-75.
- 110. Kitamura T, Qian BZ, Pollard JW. Immune cell promotion of metastasis. Nat Rev Immunol 2015;15(2):73-86.
- 111. Erler JT, Bennewith KL, Cox TR, Lang G, Bird D, Koong A, et al. Hypoxia-induced lysyl oxidase is a critical mediator of bone marrow cell recruitment to form the premetastatic niche. Cancer Cell 2009;15(1):35-44.
- 112. Hirakawa S, Brown LF, Kodama S, Paavonen K, Alitalo K, Detmar M. VEGF-C-induced lymphangiogenesis in sentinel lymph nodes promotes tumor metastasis to distant sites. Blood 2007;109(3):1010-7.
- 113. Alderton GK. Metastasis. Exosomes drive premetastatic niche formation. Nat Rev Cancer 2012;12(7):447.
- 114. Sanchez CA, Andahur EI, Valenzuela R, Castellon EA, Fulla JA, Ramos CG, et al. Exosomes from bulk and stem cells from human prostate cancer have a differential microRNA content that contributes cooperatively over local and pre-metastatic niche. Oncotarget 2015.

- 115. Torsvik A, Bjerkvig R. Mesenchymal stem cell signaling in cancer progression. Cancer Treat Rev 2013;39(2):180-8.
- 116. Mueller MM, Fusenig NE. Friends or foes bipolar effects of the tumour stroma in cancer. Nat Rev Cancer 2004;4(11):839-49.
- 117. Stetler-Stevenson WG, Yu AE. Proteases in invasion: matrix metalloproteinases. Semin Cancer Biol 2001;11(2):143-52.
- 118. Zhu ML, Kyprianou N. Role of androgens and the androgen receptor in epithelialmesenchymal transition and invasion of prostate cancer cells. FASEB J 2010;24(3):769-77.
- 119. Sun Y, Wang BE, Leong KG, Yue P, Li L, Jhunjhunwala S, et al. Androgen deprivation causes epithelial-mesenchymal transition in the prostate: implications for androgen-deprivation therapy. Cancer Res 2012;72(2):527-36.
- 120. van der Pluijm G. Epithelial plasticity, cancer stem cells and bone metastasis formation. Bone 2011;48(1):37-43.
- 121. Puhr M, Hoefer J, Schafer G, Erb HH, Oh SJ, Klocker H, et al. Epithelial-to-mesenchymal transition leads to docetaxel resistance in prostate cancer and is mediated by reduced expression of miR-200c and miR-205. Am J Pathol 2012;181(6):2188-201.
- 122. Bonnomet A, Brysse A, Tachsidis A, Waltham M, Thompson EW, Polette M, et al. Epithelial-tomesenchymal transitions and circulating tumor cells. J Mammary Gland Biol Neoplasia 2010;15(2):261-73.
- 123. Brabletz T. To differentiate or not--routes towards metastasis. Nat Rev Cancer 2012;12(6):425-36.
- 124. Kienast Y, von Baumgarten L, Fuhrmann M, Klinkert WE, Goldbrunner R, Herms J, et al. Realtime imaging reveals the single steps of brain metastasis formation. Nat Med 2010;16(1):116-22.
- 125. Psaila B, Lyden D. The metastatic niche: adapting the foreign soil. Nat Rev Cancer 2009;9(4):285-93.
- 126. Sleeman JP. The metastatic niche and stromal progression. Cancer Metastasis Rev 2012;31(3-4):429-40.
- 127. Shiozawa Y, Pedersen EA, Havens AM, Jung Y, Mishra A, Joseph J, et al. Human prostate cancer metastases target the hematopoietic stem cell niche to establish footholds in mouse bone marrow. J Clin Invest 2011;121(4):1298-312.
- 128. Sneddon JB, Werb Z. Location, location: the cancer stem cell niche. Cell Stem Cell 2007;1(6):607-11.
- 129. Hensel J, Thalmann GN. Biology of Bone Metastases in Prostate Cancer. Urology 2016.
- 130. Sosa MS, Bragado P, Aguirre-Ghiso JA. Mechanisms of disseminated cancer cell dormancy: an awakening field. Nat Rev Cancer 2014;14(9):611-22.
- 131. Aguirre-Ghiso JA. Models, mechanisms and clinical evidence for cancer dormancy. Nat Rev Cancer 2007;7(11):834-46.
- 132. Barkan D, El Touny LH, Michalowski AM, Smith JA, Chu I, Davis AS, et al. Metastatic growth from dormant cells induced by a col-I-enriched fibrotic environment. Cancer Res 2010;70(14):5706-16.
- 133. Mundy GR. Metastasis to bone: causes, consequences and therapeutic opportunities. Nat Rev Cancer 2002;2(8):584-93.
- 134. Kusumbe AP, Ramasamy SK, Adams RH. Coupling of angiogenesis and osteogenesis by a specific vessel subtype in bone. Nature 2014;507(7492):323-8.
- 135. Wang L, Benedito R, Bixel MG, Zeuschner D, Stehling M, Savendahl L, et al. Identification of a clonally expanding haematopoietic compartment in bone marrow. EMBO J 2013;32(2):219-30.
- 136. Logothetis CJ, Lin SH. Osteoblasts in prostate cancer metastasis to bone. Nat Rev Cancer 2005;5(1):21-8.
- 137. Theriault RL, Theriault RL. Biology of bone metastases. Cancer Control 2012;19(2):92-101.

- 138. Dai J, Keller J, Zhang J, Lu Y, Yao Z, Keller ET. Bone morphogenetic protein-6 promotes osteoblastic prostate cancer bone metastases through a dual mechanism. Cancer Res 2005;65(18):8274-85.
- 139. Schwaninger R, Rentsch CA, Wetterwald A, van der Horst G, van Bezooijen RL, van der Pluijm G, et al. Lack of noggin expression by cancer cells is a determinant of the osteoblast response in bone metastases. Am J Pathol 2007;170(1):160-75.
- 140. Rubin J, Chung LW, Fan X, Zhu L, Murphy TC, Nanes MS, et al. Prostate carcinoma cells that have resided in bone have an upregulated IGF-I axis. Prostate 2004;58(1):41-9.
- 141. Yang YQ, Tan YY, Wong R, Wenden A, Zhang LK, Rabie AB. The role of vascular endothelial growth factor in ossification. Int J Oral Sci 2012;4(2):64-8.
- 142. Milat F, Ng KW. Is Wnt signalling the final common pathway leading to bone formation? Mol Cell Endocrinol 2009;310(1-2):52-62.
- 143. Lynch CC, Hikosaka A, Acuff HB, Martin MD, Kawai N, Singh RK, et al. MMP-7 promotes prostate cancer-induced osteolysis via the solubilization of RANKL. Cancer Cell 2005;7(5):485-96.
- 144. Secondini C, Wetterwald A, Schwaninger R, Thalmann GN, Cecchini MG. The role of the BMP signaling antagonist noggin in the development of prostate cancer osteolytic bone metastasis. PLoS One 2011;6(1):e16078.
- 145. Roodman GD. Mechanisms of bone metastasis. N Engl J Med 2004;350(16):1655-64.
- 146. Liao J, Schneider A, Datta NS, McCauley LK. Extracellular calcium as a candidate mediator of prostate cancer skeletal metastasis. Cancer Res 2006;66(18):9065-73.
- 147. Sanders JL, Chattopadhyay N, Kifor O, Yamaguchi T, Butters RR, Brown EM. Extracellular calcium-sensing receptor expression and its potential role in regulating parathyroid hormone-related peptide secretion in human breast cancer cell lines. Endocrinology 2000;141(12):4357-64.
- 148. Sasaki A, Boyce BF, Story B, Wright KR, Chapman M, Boyce R, et al. Bisphosphonate risedronate reduces metastatic human breast cancer burden in bone in nude mice. Cancer Res 1995;55(16):3551-7.
- 149. van der Pluijm G, Que I, Sijmons B, Buijs JT, Lowik CW, Wetterwald A, et al. Interference with the microenvironmental support impairs the de novo formation of bone metastases in vivo. Cancer Res 2005;65(17):7682-90.
- 150. Yuen KK, Shelley M, Sze WM, Wilt T, Mason MD. Bisphosphonates for advanced prostate cancer. Cochrane Database Syst Rev 2006(4):CD006250.
- 151. Huggins C, Hodges CV. Studies on prostatic cancer. I. The effect of castration, of estrogen and androgen injection on serum phosphatases in metastatic carcinoma of the prostate. CA Cancer J Clin 1972;22(4):232-40.
- 152. Heidenreich A, Bastian PJ, Bellmunt J, Bolla M, Joniau S, van der Kwast T, et al. EAU guidelines on prostate cancer. Part II: Treatment of advanced, relapsing, and castration-resistant prostate cancer. Eur Urol 2014;65(2):467-79.
- 153. Tran C, Ouk S, Clegg NJ, Chen Y, Watson PA, Arora V, et al. Development of a secondgeneration antiandrogen for treatment of advanced prostate cancer. Science 2009;324(5928):787-90.
- 154. Uchio EM, Aslan M, Wells CK, Calderone J, Concato J. Impact of biochemical recurrence in prostate cancer among US veterans. Arch Intern Med 2010;170(15):1390-5.
- 155. Visakorpi T, Hyytinen E, Koivisto P, Tanner M, Keinanen R, Palmberg C, et al. In vivo amplification of the androgen receptor gene and progression of human prostate cancer. Nat Genet 1995;9(4):401-6.
- 156. Koivisto P, Kononen J, Palmberg C, Tammela T, Hyytinen E, Isola J, et al. Androgen receptor gene amplification: a possible molecular mechanism for androgen deprivation therapy failure in prostate cancer. Cancer Res 1997;57(2):314-9.

- 157. Linja MJ, Savinainen KJ, Saramaki OR, Tammela TL, Vessella RL, Visakorpi T. Amplification and overexpression of androgen receptor gene in hormone-refractory prostate cancer. Cancer Res 2001;61(9):3550-5.
- 158. Taplin ME, Bubley GJ, Shuster TD, Frantz ME, Spooner AE, Ogata GK, et al. Mutation of the androgen-receptor gene in metastatic androgen-independent prostate cancer. N Engl J Med 1995;332(21):1393-8.
- 159. Zhao XY, Malloy PJ, Krishnan AV, Swami S, Navone NM, Peehl DM, et al. Glucocorticoids can promote androgen-independent growth of prostate cancer cells through a mutated androgen receptor. Nat Med 2000;6(6):703-6.
- 160. Robzyk K, Oen H, Buchanan G, Butler LM, Tilley WD, Mandal AK, et al. Uncoupling of hormonedependence from chaperone-dependence in the L701H mutation of the androgen receptor. Mol Cell Endocrinol 2007;268(1-2):67-74.
- 161. Brooke GN, Parker MG, Bevan CL. Mechanisms of androgen receptor activation in advanced prostate cancer: differential co-activator recruitment and gene expression. Oncogene 2008;27(21):2941-50.
- 162. Steinkamp MP, O'Mahony OA, Brogley M, Rehman H, Lapensee EW, Dhanasekaran S, et al. Treatment-dependent androgen receptor mutations in prostate cancer exploit multiple mechanisms to evade therapy. Cancer Res 2009;69(10):4434-42.
- 163. Dehm SM, Schmidt LJ, Heemers HV, Vessella RL, Tindall DJ. Splicing of a novel androgen receptor exon generates a constitutively active androgen receptor that mediates prostate cancer therapy resistance. Cancer Res 2008;68(13):5469-77.
- 164. Guo Z, Yang X, Sun F, Jiang R, Linn DE, Chen H, et al. A novel androgen receptor splice variant is up-regulated during prostate cancer progression and promotes androgen depletionresistant growth. Cancer Res 2009;69(6):2305-13.
- 165. Hu R, Dunn TA, Wei S, Isharwal S, Veltri RW, Humphreys E, et al. Ligand-independent androgen receptor variants derived from splicing of cryptic exons signify hormone-refractory prostate cancer. Cancer Res 2009;69(1):16-22.
- 166. Titus MA, Schell MJ, Lih FB, Tomer KB, Mohler JL. Testosterone and dihydrotestosterone tissue levels in recurrent prostate cancer. Clin Cancer Res 2005;11(13):4653-7.
- 167. Stanbrough M, Bubley GJ, Ross K, Golub TR, Rubin MA, Penning TM, et al. Increased expression of genes converting adrenal androgens to testosterone in androgen-independent prostate cancer. Cancer Res 2006;66(5):2815-25.
- 168. Locke JA, Guns ES, Lubik AA, Adomat HH, Hendy SC, Wood CA, et al. Androgen levels increase by intratumoral de novo steroidogenesis during progression of castration-resistant prostate cancer. Cancer Res 2008;68(15):6407-15.
- 169. Montgomery RB, Mostaghel EA, Vessella R, Hess DL, Kalhorn TF, Higano CS, et al. Maintenance of intratumoral androgens in metastatic prostate cancer: a mechanism for castration-resistant tumor growth. Cancer Res 2008;68(11):4447-54.
- 170. Tannock IF, de Wit R, Berry WR, Horti J, Pluzanska A, Chi KN, et al. Docetaxel plus prednisone or mitoxantrone plus prednisone for advanced prostate cancer. N Engl J Med 2004;351(15):1502-12.
- 171. de Bono JS, Oudard S, Ozguroglu M, Hansen S, Machiels JP, Kocak I, et al. Prednisone plus cabazitaxel or mitoxantrone for metastatic castration-resistant prostate cancer progressing after docetaxel treatment: a randomised open-label trial. Lancet 2010;376(9747):1147-54.
- 172. de Bono JS, Logothetis CJ, Molina A, Fizazi K, North S, Chu L, et al. Abiraterone and increased survival in metastatic prostate cancer. N Engl J Med 2011;364(21):1995-2005.
- 173. Fizazi K, Scher HI, Molina A, Logothetis CJ, Chi KN, Jones RJ, et al. Abiraterone acetate for treatment of metastatic castration-resistant prostate cancer: final overall survival analysis of the COU-AA-301 randomised, double-blind, placebo-controlled phase 3 study. Lancet Oncol 2012;13(10):983-92.

- 174. Scher HI, Fizazi K, Saad F, Taplin ME, Sternberg CN, Miller K, et al. Increased survival with enzalutamide in prostate cancer after chemotherapy. N Engl J Med 2012;367(13):1187-97.
- 175. van Soest RJ, de Wit R. Irrefutable evidence for the use of docetaxel in newly diagnosed metastatic prostate cancer: results from the STAMPEDE and CHAARTED trials. BMC Med 2015;13(1):304.
- 176. Fizazi K, Faivre L, Lesaunier F, Delva R, Gravis G, Rolland F, et al. Androgen deprivation therapy plus docetaxel and estramustine versus androgen deprivation therapy alone for high-risk localised prostate cancer (GETUG 12): a phase 3 randomised controlled trial. Lancet Oncol 2015;16(7):787-94.
- 177. Grisanzio C, Signoretti S. p63 in prostate biology and pathology. J Cell Biochem 2008;103(5):1354-68.
- 178. Richardson GD, Robson CN, Lang SH, Neal DE, Maitland NJ, Collins AT. CD133, a novel marker for human prostatic epithelial stem cells. J Cell Sci 2004;117(Pt 16):3539-45.
- 179. Maitland NJ, Frame FM, Polson ES, Lewis JL, Collins AT. Prostate cancer stem cells: do they have a basal or luminal phenotype? Horm Cancer 2011;2(1):47-61.
- 180. Hurt EM, Kawasaki BT, Klarmann GJ, Thomas SB, Farrar WL. CD44+ CD24(-) prostate cells are early cancer progenitor/stem cells that provide a model for patients with poor prognosis. Br J Cancer 2008;98(4):756-65.
- 181. Goldstein AS, Lawson DA, Cheng D, Sun W, Garraway IP, Witte ON. Trop2 identifies a subpopulation of murine and human prostate basal cells with stem cell characteristics. Proc Natl Acad Sci U S A 2008;105(52):20882-7.
- 182. Liu AY, Roudier MP, True LD. Heterogeneity in primary and metastatic prostate cancer as defined by cell surface CD profile. Am J Pathol 2004;165(5):1543-56.
- 183. Karthaus WR, Iaquinta PJ, Drost J, Gracanin A, van Boxtel R, Wongvipat J, et al. Identification of multipotent luminal progenitor cells in human prostate organoid cultures. Cell 2014;159(1):163-75.
- 184. Eaton CL, Colombel M, van der Pluijm G, Cecchini M, Wetterwald A, Lippitt J, et al. Evaluation of the frequency of putative prostate cancer stem cells in primary and metastatic prostate cancer. Prostate 2010;70(8):875-82.
- 185. Colombel M, Eaton CL, Hamdy F, Ricci E, van der Pluijm G, Cecchini M, et al. Increased expression of putative cancer stem cell markers in primary prostate cancer is associated with progression of bone metastases. Prostate 2012;72(7):713-20.
- 186. Maitland NJ, Collins AT. Prostate cancer stem cells: a new target for therapy. J Clin Oncol 2008;26(17):2862-70.
- 187. Dean M, Fojo T, Bates S. Tumour stem cells and drug resistance. Nat Rev Cancer 2005;5(4):275-84.
- 188. Kypta RM, Waxman J. Wnt/beta-catenin signalling in prostate cancer. Nat Rev Urol 2012;9(8):418-28.
- 189. Verras M, Sun Z. Roles and regulation of Wnt signaling and beta-catenin in prostate cancer. Cancer Lett 2006;237(1):22-32.
- 190. Terry S, Yang X, Chen MW, Vacherot F, Buttyan R. Multifaceted interaction between the androgen and Wnt signaling pathways and the implication for prostate cancer. J Cell Biochem 2006;99(2):402-10.
- 191. Robinson DR, Zylstra CR, Williams BO. Wnt signaling and prostate cancer. Curr Drug Targets 2008;9(7):571-80.
- 192. Wang G, Wang J, Sadar MD. Crosstalk between the androgen receptor and beta-catenin in castrate-resistant prostate cancer. Cancer Res 2008;68(23):9918-27.
- 193. Semenov M, Tamai K, He X. SOST is a ligand for LRP5/LRP6 and a Wnt signaling inhibitor. J Biol Chem 2005;280(29):26770-5.

- 194. Rentsch CA, Cecchini MG, Thalmann GN. Loss of inhibition over master pathways of bone mass regulation results in osteosclerotic bone metastases in prostate cancer. Swiss Med Wkly 2009;139(15-16):220-5.
- 195. Wang Y. Wnt/Planar cell polarity signaling: a new paradigm for cancer therapy. Mol Cancer Ther 2009;8(8):2103-9.
- 196. Veeman MT, Axelrod JD, Moon RT. A second canon. Functions and mechanisms of betacatenin-independent Wnt signaling. Dev Cell 2003;5(3):367-77.
- 197. Davies G, Jiang WG, Mason MD. Cell-cell adhesion molecules and signaling intermediates and their role in the invasive potential of prostate cancer cells. J Urol 2000;163(3):985-92.
- 198. Carvalho FL, Simons BW, Eberhart CG, Berman DM. Notch signaling in prostate cancer: a moving target. Prostate 2014;74(9):933-45.
- 199. Domingo-Domenech J, Vidal SJ, Rodriguez-Bravo V, Castillo-Martin M, Quinn SA, Rodriguez-Barrueco R, et al. Suppression of acquired docetaxel resistance in prostate cancer through depletion of notch- and hedgehog-dependent tumor-initiating cells. Cancer Cell 2012;22(3):373-88.
- 200. Zoni E, van der Pluijm G, Gray PC, Kruithof-de Julio M. Epithelial Plasticity in Cancer: Unmasking a MicroRNA Network for TGF-beta-, Notch-, and Wnt-Mediated EMT. J Oncol 2015;2015:198967.
- 201. Collu GM, Hidalgo-Sastre A, Brennan K. Wnt-Notch signalling crosstalk in development and disease. Cell Mol Life Sci 2014;71(18):3553-67.
- 202. Han L, Diehl A, Nguyen NK, Korangath P, Teo W, Cho S, et al. The Notch pathway inhibits TGFbeta signaling in breast cancer through HEYL-mediated crosstalk. Cancer Res 2014;74(22):6509-18.
- 203. Villaronga MA, Bevan CL, Belandia B. Notch signaling: a potential therapeutic target in prostate cancer. Curr Cancer Drug Targets 2008;8(7):566-80.
- 204. Belandia B, Powell SM, Garcia-Pedrero JM, Walker MM, Bevan CL, Parker MG. Hey1, a mediator of notch signaling, is an androgen receptor corepressor. Mol Cell Biol 2005;25(4):1425-36.
- 205. Hales EC, Taub JW, Matherly LH. New insights into Notch1 regulation of the PI3K-AKT-mTOR1 signaling axis: targeted therapy of gamma-secretase inhibitor resistant T-cell acute lymphoblastic leukemia. Cell Signal 2014;26(1):149-61.
- 206. Grishina IB, Kim SY, Ferrara C, Makarenkova HP, Walden PD. BMP7 inhibits branching morphogenesis in the prostate gland and interferes with Notch signaling. Dev Biol 2005;288(2):334-47.
- 207. Ghosh S, Lau H, Simons BW, Powell JD, Meyers DJ, De Marzo AM, et al. PI3K/mTOR signaling regulates prostatic branching morphogenesis. Dev Biol 2011;360(2):329-42.
- 208. Whelan JT, Kellogg A, Shewchuk BM, Hewan-Lowe K, Bertrand FE. Notch-1 signaling is lost in prostate adenocarcinoma and promotes PTEN gene expression. J Cell Biochem 2009;107(5):992-1001.
- 209. Carver BS, Chapinski C, Wongvipat J, Hieronymus H, Chen Y, Chandarlapaty S, et al. Reciprocal feedback regulation of PI3K and androgen receptor signaling in PTEN-deficient prostate cancer. Cancer Cell 2011;19(5):575-86.
- 210. Pourmand G, Ziaee AA, Abedi AR, Mehrsai A, Alavi HA, Ahmadi A, et al. Role of PTEN gene in progression of prostate cancer. Urol J 2007;4(2):95-100.
- 211. Santagata S, Demichelis F, Riva A, Varambally S, Hofer MD, Kutok JL, et al. JAGGED1 expression is associated with prostate cancer metastasis and recurrence. Cancer Res 2004;64(19):6854-7.
- 212. Sethi N, Kang Y. Notch signalling in cancer progression and bone metastasis. Br J Cancer 2011;105(12):1805-10.
- 213. Sethi N, Dai X, Winter CG, Kang Y. Tumor-derived JAGGED1 promotes osteolytic bone metastasis of breast cancer by engaging notch signaling in bone cells. Cancer Cell 2011;19(2):192-205.

- 214. Cui D, Dai J, Keller JM, Mizokami A, Xia S, Keller ET. Notch Pathway Inhibition Using PF-03084014, a gamma-Secretase Inhibitor (GSI), Enhances the Antitumor Effect of Docetaxel in Prostate Cancer. Clin Cancer Res 2015;21(20):4619-29.
- 215. Jones E, Pu H, Kyprianou N. Targeting TGF-beta in prostate cancer: therapeutic possibilities during tumor progression. Expert Opin Ther Targets 2009;13(2):227-34.
- 216. Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. J Clin Invest 2009;119(6):1420-8.
- 217. Massague J, Blain SW, Lo RS. TGFbeta signaling in growth control, cancer, and heritable disorders. Cell 2000;103(2):295-309.
- 218. Massague J. TGFbeta in Cancer. Cell 2008;134(2):215-30.
- 219. Wu MY, Hill CS. Tgf-beta superfamily signaling in embryonic development and homeostasis. Dev Cell 2009;16(3):329-43.
- 220. Chen D, Zhao M, Mundy GR. Bone morphogenetic proteins. Growth Factors 2004;22(4):233-41.
- 221. Franzen P, ten Dijke P, Ichijo H, Yamashita H, Schulz P, Heldin CH, et al. Cloning of a TGF beta type I receptor that forms a heteromeric complex with the TGF beta type II receptor. Cell 1993;75(4):681-92.
- 222. ten Dijke P, Ichijo H, Franzen P, Schulz P, Saras J, Toyoshima H, et al. Activin receptor-like kinases: a novel subclass of cell-surface receptors with predicted serine/threonine kinase activity. Oncogene 1993;8(10):2879-87.
- 223. ten Dijke P, Yamashita H, Sampath TK, Reddi AH, Estevez M, Riddle DL, et al. Identification of type I receptors for osteogenic protein-1 and bone morphogenetic protein-4. J Biol Chem 1994;269(25):16985-8.
- 224. Yamashita H, ten Dijke P, Huylebroeck D, Sampath TK, Andries M, Smith JC, et al. Osteogenic protein-1 binds to activin type II receptors and induces certain activin-like effects. J Cell Biol 1995;130(1):217-26.
- 225. Liu F, Ventura F, Doody J, Massague J. Human type II receptor for bone morphogenic proteins (BMPs): extension of the two-kinase receptor model to the BMPs. Mol Cell Biol 1995;15(7):3479-86.
- 226. Nohno T, Ishikawa T, Saito T, Hosokawa K, Noji S, Wolsing DH, et al. Identification of a human type II receptor for bone morphogenetic protein-4 that forms differential heteromeric complexes with bone morphogenetic protein type I receptors. J Biol Chem 1995;270(38):22522-6.
- 227. Ebisawa T, Tada K, Kitajima I, Tojo K, Sampath TK, Kawabata M, et al. Characterization of bone morphogenetic protein-6 signaling pathways in osteoblast differentiation. J Cell Sci 1999;112 (Pt 20):3519-27.
- 228. Aoki H, Fujii M, Imamura T, Yagi K, Takehara K, Kato M, et al. Synergistic effects of different bone morphogenetic protein type I receptors on alkaline phosphatase induction. J Cell Sci 2001;114(Pt 8):1483-9.
- 229. Rosenzweig BL, Imamura T, Okadome T, Cox GN, Yamashita H, ten Dijke P, et al. Cloning and characterization of a human type II receptor for bone morphogenetic proteins. Proc Natl Acad Sci U S A 1995;92(17):7632-6.
- 230. Scharpfenecker M, van Dinther M, Liu Z, van Bezooijen RL, Zhao Q, Pukac L, et al. BMP-9 signals via ALK1 and inhibits bFGF-induced endothelial cell proliferation and VEGF-stimulated angiogenesis. J Cell Sci 2007;120(Pt 6):964-72.
- 231. David L, Mallet C, Mazerbourg S, Feige JJ, Bailly S. Identification of BMP9 and BMP10 as functional activators of the orphan activin receptor-like kinase 1 (ALK1) in endothelial cells. Blood 2007;109(5):1953-61.
- 232. Wu Y, Zhou BP. New insights of epithelial-mesenchymal transition in cancer metastasis. Acta Biochim Biophys Sin (Shanghai) 2008;40(7):643-50.
- 233. Shi Y, Massague J. Mechanisms of TGF-beta signaling from cell membrane to the nucleus. Cell 2003;113(6):685-700.

- 234. Moustakas A, Heldin CH. Non-Smad TGF-beta signals. J Cell Sci 2005;118(Pt 16):3573-84.
- 235. Pardali K, Moustakas A. Actions of TGF-beta as tumor suppressor and pro-metastatic factor in human cancer. Biochim Biophys Acta 2007;1775(1):21-62.
- 236. Zhang YE. Non-Smad pathways in TGF-beta signaling. Cell Res 2009;19(1):128-39.
- 237. Lonn P, Moren A, Raja E, Dahl M, Moustakas A. Regulating the stability of TGFbeta receptors and Smads. Cell Res 2009;19(1):21-35.
- 238. Danen EH, Sonnenberg A. Integrins in regulation of tissue development and function. J Pathol 2003;200(4):471-80.
- 239. Danen EH. Integrins: regulators of tissue function and cancer progression. Curr Pharm Des 2005;11(7):881-91.
- 240. Hynes RO. Integrins: bidirectional, allosteric signaling machines. Cell 2002;110(6):673-87.
- 241. Pontes-Junior J, Reis ST, Dall'Oglio M, Neves de Oliveira LC, Cury J, Carvalho PA, et al. Evaluation of the expression of integrins and cell adhesion molecules through tissue microarray in lymph node metastases of prostate cancer. J Carcinog 2009;8:3.
- 242. Kikkawa H, Kaihou M, Horaguchi N, Uchida T, Imafuku H, Takiguchi A, et al. Role of integrin alpha(v)beta3 in the early phase of liver metastasis: PET and IVM analyses. Clin Exp Metastasis 2002;19(8):717-25.
- 243. Enns A, Korb T, Schluter K, Gassmann P, Spiegel HU, Senninger N, et al. Alphavbeta5-integrins mediate early steps of metastasis formation. Eur J Cancer 2005;41(7):1065-72.
- 244. Margadant C, Sonnenberg A. Integrin-TGF-beta crosstalk in fibrosis, cancer and wound healing. EMBO Rep 2010;11(2):97-105.
- 245. Xiong J, Balcioglu HE, Danen EH. Integrin signaling in control of tumor growth and progression. Int J Biochem Cell Biol 2013;45(5):1012-5.
- 246. Truong H, Danen EH. Integrin switching modulates adhesion dynamics and cell migration. Cell Adh Migr 2009;3(2):179-81.
- 247. Yamada KM, Even-Ram S. Integrin regulation of growth factor receptors. Nat Cell Biol 2002;4(4):E75-6.
- 248. Chen Q, Lin TH, Der CJ, Juliano RL. Integrin-mediated activation of MEK and mitogenactivated protein kinase is independent of Ras [corrected]. J Biol Chem 1996;271(30):18122-7.
- 249. Figel S, Gelman IH. Focal adhesion kinase controls prostate cancer progression via intrinsic kinase and scaffolding functions. Anticancer Agents Med Chem 2011;11(7):607-16.
- 250. Keller ET, Fu Z, Yeung K, Brennan M. Raf kinase inhibitor protein: a prostate cancer metastasis suppressor gene. Cancer Lett 2004;207(2):131-7.
- 251. Fornaro M, Manes T, Languino LR. Integrins and prostate cancer metastases. Cancer Metastasis Rev 2001;20(3-4):321-31.
- 252. Ren B, Yu YP, Tseng GC, Wu C, Chen K, Rao UN, et al. Analysis of integrin alpha7 mutations in prostate cancer, liver cancer, glioblastoma multiforme, and leiomyosarcoma. J Natl Cancer Inst 2007;99(11):868-80.
- 253. Bonkhoff H, Stein U, Remberger K. Differential expression of alpha 6 and alpha 2 very late antigen integrins in the normal, hyperplastic, and neoplastic prostate: simultaneous demonstration of cell surface receptors and their extracellular ligands. Hum Pathol 1993;24(3):243-8.
- 254. King TE, Pawar SC, Majuta L, Sroka IC, Wynn D, Demetriou MC, et al. The role of alpha 6 integrin in prostate cancer migration and bone pain in a novel xenograft model. PLoS One 2008;3(10):e3535.
- 255. van den Hoogen C, van der Horst G, Cheung H, Buijs JT, Pelger RC, van der Pluijm G. Integrin alphav expression is required for the acquisition of a metastatic stem/progenitor cell phenotype in human prostate cancer. Am J Pathol 2011;179(5):2559-68.
- 256. van der Horst G, van den Hoogen C, Buijs JT, Cheung H, Bloys H, Pelger RC, et al. Targeting of alpha(v)-integrins in stem/progenitor cells and supportive microenvironment impairs bone metastasis in human prostate cancer. Neoplasia 2011;13(6):516-25.

- 257. Zoni E, van der Horst G, van de Merbel AF, Chen L, Rane JK, Pelger RC, et al. miR-25 Modulates Invasiveness and Dissemination of Human Prostate Cancer Cells via Regulation of alphav- and alpha6-Integrin Expression. Cancer Res 2015;75(11):2326-36.
- 258. Pechkovsky DV, Scaffidi AK, Hackett TL, Ballard J, Shaheen F, Thompson PJ, et al. Transforming growth factor beta1 induces alphavbeta3 integrin expression in human lung fibroblasts via a beta3 integrin-, c-Src-, and p38 MAPK-dependent pathway. J Biol Chem 2008;283(19):12898-908.
- 259. Munger JS, Huang X, Kawakatsu H, Griffiths MJ, Dalton SL, Wu J, et al. The integrin alpha v beta 6 binds and activates latent TGF beta 1: a mechanism for regulating pulmonary inflammation and fibrosis. Cell 1999;96(3):319-28.
- 260. Sun H, Hu K, Wu M, Xiong J, Yuan L, Tang Y, et al. Contact by melanoma cells causes malignant transformation of human epithelial-like stem cells via alpha V integrin activation of transforming growth factor beta1 signaling. Exp Biol Med (Maywood) 2011;236(3):352-65.
- 261. Coleman RE, Major P, Lipton A, Brown JE, Lee KA, Smith M, et al. Predictive value of bone resorption and formation markers in cancer patients with bone metastases receiving the bisphosphonate zoledronic acid. J Clin Oncol 2005;23(22):4925-35.
- 262. Chin SL, Johnson SA, Quinn J, Mirosavljevic D, Price JT, Dudley AC, et al. A role for alphaV integrin subunit in TGF-beta-stimulated osteoclastogenesis. Biochem Biophys Res Commun 2003;307(4):1051-8.
- 263. Klauzinska M, Castro NP, Rangel MC, Spike BT, Gray PC, Bertolette D, et al. The multifaceted role of the embryonic gene Cripto-1 in cancer, stem cells and epithelial-mesenchymal transition. Semin Cancer Biol 2014;29:51-8.
- 264. Shani G, Fischer WH, Justice NJ, Kelber JA, Vale W, Gray PC. GRP78 and Cripto form a complex at the cell surface and collaborate to inhibit transforming growth factor beta signaling and enhance cell growth. Mol Cell Biol 2008;28(2):666-77.
- 265. Shen MM. Nodal signaling: developmental roles and regulation. Development 2007;134(6):1023-34.
- 266. Cheng SK, Olale F, Bennett JT, Brivanlou AH, Schier AF. EGF-CFC proteins are essential coreceptors for the TGF-beta signals Vg1 and GDF1. Genes Dev 2003;17(1):31-6.
- 267. Chen C, Ware SM, Sato A, Houston-Hawkins DE, Habas R, Matzuk MM, et al. The Vg1-related protein Gdf3 acts in a Nodal signaling pathway in the pre-gastrulation mouse embryo. Development 2006;133(2):319-29.
- 268. Gray PC, Harrison CA, Vale W. Cripto forms a complex with activin and type II activin receptors and can block activin signaling. Proc Natl Acad Sci U S A 2003;100(9):5193-8.
- 269. Kelber JA, Shani G, Booker EC, Vale WW, Gray PC. Cripto is a noncompetitive activin antagonist that forms analogous signaling complexes with activin and nodal. J Biol Chem 2008;283(8):4490-500.
- 270. Kelber JA, Panopoulos AD, Shani G, Booker EC, Belmonte JC, Vale WW, et al. Blockade of Cripto binding to cell surface GRP78 inhibits oncogenic Cripto signaling via MAPK/PI3K and Smad2/3 pathways. Oncogene 2009;28(24):2324-36.
- 271. Gray PC, Shani G, Aung K, Kelber J, Vale W. Cripto binds transforming growth factor beta (TGFbeta) and inhibits TGF-beta signaling. Mol Cell Biol 2006;26(24):9268-78.
- 272. Shukla A, Ho Y, Liu X, Ryscavage A, Glick AB. Cripto-1 alters keratinocyte differentiation via blockade of transforming growth factor-beta1 signaling: role in skin carcinogenesis. Mol Cancer Res 2008;6(3):509-16.
- 273. Yan YT, Liu JJ, Luo Y, E C, Haltiwanger RS, Abate-Shen C, et al. Dual roles of Cripto as a ligand and coreceptor in the nodal signaling pathway. Mol Cell Biol 2002;22(13):4439-49.
- 274. Nagaoka T, Karasawa H, Castro NP, Rangel MC, Salomon DS, Bianco C. An evolving web of signaling networks regulated by Cripto-1. Growth Factors 2012;30(1):13-21.
- 275. Strizzi L, Bianco C, Normanno N, Salomon D. Cripto-1: a multifunctional modulator during embryogenesis and oncogenesis. Oncogene 2005;24(37):5731-41.

- 276. Tao Q, Yokota C, Puck H, Kofron M, Birsoy B, Yan D, et al. Maternal wnt11 activates the canonical wnt signaling pathway required for axis formation in Xenopus embryos. Cell 2005;120(6):857-71.
- 277. Watanabe K, Nagaoka T, Lee JM, Bianco C, Gonzales M, Castro NP, et al. Enhancement of Notch receptor maturation and signaling sensitivity by Cripto-1. J Cell Biol 2009;187(3):343-53.
- 278. Bianco C, Strizzi L, Normanno N, Khan N, Salomon DS. Cripto-1: an oncofetal gene with many faces. Curr Top Dev Biol 2005;67:85-133.
- 279. Nagaoka T, Karasawa H, Turbyville T, Rangel MC, Castro NP, Gonzales M, et al. Cripto-1 enhances the canonical Wnt/beta-catenin signaling pathway by binding to LRP5 and LRP6 coreceptors. Cell Signal 2013;25(1):178-89.
- 280. Postovit LM, Seftor EA, Seftor RE, Hendrix MJ. Targeting Nodal in malignant melanoma cells. Expert Opin Ther Targets 2007;11(4):497-505.
- 281. Terry S, El-Sayed IY, Destouches D, Maille P, Nicolaiew N, Ploussard G, et al. CRIPTO overexpression promotes mesenchymal differentiation in prostate carcinoma cells through parallel regulation of AKT and FGFR activities. Oncotarget 2015;6(14):11994-2008.
- 282. Cai X, Hagedorn CH, Cullen BR. Human microRNAs are processed from capped, polyadenylated transcripts that can also function as mRNAs. RNA 2004;10(12):1957-66.
- 283. Lee Y, Kim M, Han J, Yeom KH, Lee S, Baek SH, et al. MicroRNA genes are transcribed by RNA polymerase II. EMBO J 2004;23(20):4051-60.
- 284. Borchert GM, Lanier W, Davidson BL. RNA polymerase III transcribes human microRNAs. Nat Struct Mol Biol 2006;13(12):1097-101.
- 285. Lee Y, Jeon K, Lee JT, Kim S, Kim VN. MicroRNA maturation: stepwise processing and subcellular localization. EMBO J 2002;21(17):4663-70.
- 286. Ozsolak F, Poling LL, Wang Z, Liu H, Liu XS, Roeder RG, et al. Chromatin structure analyses identify miRNA promoters. Genes Dev 2008;22(22):3172-83.
- 287. Monteys AM, Spengler RM, Wan J, Tecedor L, Lennox KA, Xing Y, et al. Structure and activity of putative intronic miRNA promoters. RNA 2010;16(3):495-505.
- 288. Ha M, Kim VN. Regulation of microRNA biogenesis. Nat Rev Mol Cell Biol 2014;15(8):509-24.
- 289. Lee Y, Ahn C, Han J, Choi H, Kim J, Yim J, et al. The nuclear RNase III Drosha initiates microRNA processing. Nature 2003;425(6956):415-9.
- 290. Bohnsack MT, Czaplinski K, Gorlich D. Exportin 5 is a RanGTP-dependent dsRNA-binding protein that mediates nuclear export of pre-miRNAs. RNA 2004;10(2):185-91.
- 291. Lund E, Guttinger S, Calado A, Dahlberg JE, Kutay U. Nuclear export of microRNA precursors. Science 2004;303(5654):95-8.
- 292. Yi R, Qin Y, Macara IG, Cullen BR. Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. Genes Dev 2003;17(24):3011-6.
- 293. Bernstein E, Caudy AA, Hammond SM, Hannon GJ. Role for a bidentate ribonuclease in the initiation step of RNA interference. Nature 2001;409(6818):363-6.
- 294. Hutvagner G, McLachlan J, Pasquinelli AE, Balint E, Tuschl T, Zamore PD. A cellular function for the RNA-interference enzyme Dicer in the maturation of the let-7 small temporal RNA. Science 2001;293(5531):834-38.
- 295. Ketting RF, Fischer SE, Bernstein E, Sijen T, Hannon GJ, Plasterk RH. Dicer functions in RNA interference and in synthesis of small RNA involved in developmental timing in C. elegans. Genes Dev 2001;15(20):2654-9.
- 296. Macrae IJ, Zhou K, Li F, Repic A, Brooks AN, Cande WZ, et al. Structural basis for doublestranded RNA processing by Dicer. Science 2006;311(5758):195-8.
- 297. Park JE, Heo I, Tian Y, Simanshu DK, Chang H, Jee D, et al. Dicer recognizes the 5' end of RNA for efficient and accurate processing. Nature 2011;475(7355):201-5.
- 298. Tian Y, Simanshu DK, Ma JB, Park JE, Heo I, Kim VN, et al. A phosphate-binding pocket within the platform-PAZ-connector helix cassette of human Dicer. Mol Cell 2014;53(4):606-16.

- 299. Hessels D, Schalken JA. Urinary biomarkers for prostate cancer: a review. Asian J Androl 2013;15(3):333-9.
- 300. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, et al. Circulating microRNAs as stable blood-based markers for cancer detection. Proc Natl Acad Sci U S A 2008;105(30):10513-8.
- 301. Yaman Agaoglu F, Kovancilar M, Dizdar Y, Darendeliler E, Holdenrieder S, Dalay N, et al. Investigation of miR-21, miR-141, and miR-221 in blood circulation of patients with prostate cancer. Tumour Biol 2011;32(3):583-8.
- 302. Brase JC, Wuttig D, Kuner R, Sultmann H. Serum microRNAs as non-invasive biomarkers for cancer. Mol Cancer 2010;9:306.
- 303. Bryant RJ, Pawlowski T, Catto JW, Marsden G, Vessella RL, Rhees B, et al. Changes in circulating microRNA levels associated with prostate cancer. Br J Cancer 2012;106(4):768-74.
- 304. Huang X, Yuan T, Liang M, Du M, Xia S, Dittmar R, et al. Exosomal miR-1290 and miR-375 as prognostic markers in castration-resistant prostate cancer. Eur Urol 2015;67(1):33-41.
- 305. Lin HM, Castillo L, Mahon KL, Chiam K, Lee BY, Nguyen Q, et al. Circulating microRNAs are associated with docetaxel chemotherapy outcome in castration-resistant prostate cancer. Br J Cancer 2014;110(10):2462-71.
- 306. Rane JK, Scaravilli M, Ylipaa A, Pellacani D, Mann VM, Simms MS, et al. MicroRNA expression profile of primary prostate cancer stem cells as a source of biomarkers and therapeutic targets. Eur Urol 2015;67(1):7-10.
- 307. Lewis BP, Burge CB, Bartel DP. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. Cell 2005;120(1):15-20.
- 308. Gu S, Jin L, Zhang F, Sarnow P, Kay MA. Biological basis for restriction of microRNA targets to the 3' untranslated region in mammalian mRNAs. Nat Struct Mol Biol 2009;16(2):144-50.
- 309. Garzon R, Marcucci G, Croce CM. Targeting microRNAs in cancer: rationale, strategies and challenges. Nat Rev Drug Discov 2010;9(10):775-89.
- 310. Okamura K, Liu N, Lai EC. Distinct mechanisms for microRNA strand selection by Drosophila Argonautes. Mol Cell 2009;36(3):431-44.
- 311. Hu HY, Yan Z, Xu Y, Hu H, Menzel C, Zhou YH, et al. Sequence features associated with microRNA strand selection in humans and flies. BMC Genomics 2009;10:413.
- 312. Beilharz TH, Humphreys DT, Clancy JL, Thermann R, Martin DI, Hentze MW, et al. microRNAmediated messenger RNA deadenylation contributes to translational repression in mammalian cells. PLoS One 2009;4(8):e6783.
- 313. Mathonnet G, Fabian MR, Svitkin YV, Parsyan A, Huck L, Murata T, et al. MicroRNA inhibition of translation initiation in vitro by targeting the cap-binding complex eIF4F. Science 2007;317(5845):1764-7.
- 314. Wang B, Yanez A, Novina CD. MicroRNA-repressed mRNAs contain 40S but not 60S components. Proc Natl Acad Sci U S A 2008;105(14):5343-8.
- 315. Petersen CP, Bordeleau ME, Pelletier J, Sharp PA. Short RNAs repress translation after initiation in mammalian cells. Mol Cell 2006;21(4):533-42.
- 316. Nottrott S, Simard MJ, Richter JD. Human let-7a miRNA blocks protein production on actively translating polyribosomes. Nat Struct Mol Biol 2006;13(12):1108-14.
- 317. Liu J, Valencia-Sanchez MA, Hannon GJ, Parker R. MicroRNA-dependent localization of targeted mRNAs to mammalian P-bodies. Nat Cell Biol 2005;7(7):719-23.
- 318. Eulalio A, Huntzinger E, Izaurralde E. Getting to the root of miRNA-mediated gene silencing. Cell 2008;132(1):9-14.
- 319. Fan X, Chen X, Deng W, Zhong G, Cai Q, Lin T. Up-regulated microRNA-143 in cancer stem cells differentiation promotes prostate cancer cells metastasis by modulating FNDC3B expression. BMC Cancer 2013;13:61.
- 320. Li K, Liu C, Zhou B, Bi L, Huang H, Lin T, et al. Role of EZH2 in the growth of prostate cancer stem cells isolated from LNCaP cells. Int J Mol Sci 2013;14(6):11981-93.

- 321. Josson S, Gururajan M, Hu P, Shao C, Chu GY, Zhau HE, et al. miR-409-3p/-5p promotes tumorigenesis, epithelial-to-mesenchymal transition, and bone metastasis of human prostate cancer. Clin Cancer Res 2014;20(17):4636-46.
- 322. Liu C, Kelnar K, Vlassov AV, Brown D, Wang J, Tang DG. Distinct microRNA expression profiles in prostate cancer stem/progenitor cells and tumor-suppressive functions of let-7. Cancer Res 2012;72(13):3393-404.
- 323. Liu C, Kelnar K, Liu B, Chen X, Calhoun-Davis T, Li H, et al. The microRNA miR-34a inhibits prostate cancer stem cells and metastasis by directly repressing CD44. Nat Med 2011;17(2):211-5.