Cover Page



Universiteit Leiden



The handle <u>http://hdl.handle.net/1887/20982</u> holds various files of this Leiden University dissertation.

Author: Gierman, Lobke Marijn Title: Inflammation : a link between metabolic syndrome and osteoarthritis? Issue Date: 2013-06-18

1

General introduction to osteoarthritis



1.1 Introduction

In 1743 William Hunter stated; From Hippocrates to the present age it is universally allowed that ulcerated cartilage is a troublesome thing and that once destroyed, is not repaired (1). The fact that destroyed cartilage is the main feature of the disease osteoarthritis (OA), makes it that researchers since decades are aiming to find the holy grail how to interfere in the pathogenesis of OA. In general, OA is described as a heterogeneous joint disease characterized by a progressive loss of cartilage.

1.1.1 Facts and figures

Worldwide it is estimated that 9.6% of men and 18% of women aged >60 years have symptomatic OA (2). However, these numbers should be interpreted with caution as the definition of OA is unclear and its onset is difficult to determine. Based on radiographs, OA in the hand joints is most frequently followed by knee and hip (3, 4). In general, OA is more prevalent in Europe and the USA than in other parts of the world (figure 1). OA is a major cause of impaired mobility and it belongs to the top ten of leading causes of burden of disease in high-income countries (5). In the Netherlands, most recent data from General Practices indicate that the prevalence of OA is 29/1000 for males and 50/1000 for females in the adult population (6).

1.1.2 Symptoms

Osteoarthritis is a heterogeneous disorder which is diagnosed based on symptoms, joint pathology or a combination of these two. Symptoms attributable to OA include pain, cracking (crepitus) and stiffness in the affected joints. The presence of osteophytes, joint space narrowing, sclerosis and altered shape of the bone end can be assessed, e.g. by using radiograph, and are in general classified on behalf of the Kellgren-Lawrence grading system (7). The most used diagnostic criteria were developed by the American College of Rheumatology (ACR) (8-10). These criteria include a combination of symptoms (pain) with radiographs.

1.1.3 Risk factors

Several risk factors for the development and progression of OA have been determined. The increase of age is one of the major contributors (figure 1) and is seen in all joints. After the age of 50, the prevalence and incidence of OA in the female gender is significantly greater than in men (11). Other risk factors which are frequently shown to increase OA occurrence are e.g. obesity, sex hormones, ethnicity and race, genetic predisposition and joint trauma (12). The mechanisms by which these risk factors contribute to the development and progression of OA are far from understood.



Figure 1. Prevalence of osteoarthritis of the knee, by age group, sex and region in 2000 (World Health Organization). A regions=developed countries in North America, Western Europe, Japan, Australia and New Zealand. AF=countries in sub-Saharan Africa. AM BD=developing countries in the Americas. EM=countries in the Eastern Mediterranean and North African regions. EU BC=developing countries in Europe. SEA=countries in Southeast Asia. WP B=countries in the Western Pacific region (2).

1.1.4 Problems and challenges

Existing therapies are primarily aimed to reduce pain and no cure is available which can interfere in the pathology of OA. Based on different risk factors (and mechanisms involved) and frequently observed differences in disease progression (between patients but also between joints) it becomes increasingly clear that OA is a disease which involves multiple tissues (13). It is hypothesized that a variety of OA forms may exist that are similar with respect to outcome, however have a different underlying pathophysiological process. This may explain the variable outcomes of clinical trials, biomarker studies and genetic association studies, and therefore the difficulties observed when analyzing the efficacy of novel drugs. Without patient stratification, clinical trials may be 'contaminated' with patients that respond differently to interventions, which results in very large and costly clinical trials and prohibits the process of the development of new disease interfering therapies. To improve prediction of disease outcome, to optimize clinical trial efficiency and to analyze the efficacy of novel drugs, it is crucial to better understand the various mechanisms leading to the clinical outcome of OA.

Currently, OA management is directed toward patients in the latter phase of the disease. Although it is difficult to detect OA in an earlier stage, as radiographs are not sensitive enough, emphasis on early diagnosis and prevention could have more significance. Relevant biomarkers, objective measures that can be derived from body fluids such as blood or urine, are needed to diagnose and forecast OA in an earlier phase of the disease (14). To obtain representative biomarkers it is essential to better understand the role of local and systemic factors, which are involved in the pathogenesis of OA.

1.2 Local alterations; from cartilage to multiple tissue disease

OA has long been considered a wear and tear disease leading to loss of cartilage. During the past decades there have been significant developments in the scientific understanding of OA. These days OA is appreciated as a disease affecting the whole joint which involves complex interactions between several joint tissues (figure 2)(15, 16).

1.2.1 Cartilage

Articular cartilage is a highly specialized avascular connective tissue which provides smooth articulation and bending of the joints during movement. The extracellular matrix of cartilage consists mainly of collagen type II. Collagen type II provides a network in which other constituents, such as proteoglycans and chondrocytes, are embedded. Collagens and proteoglycans give cartilage the capacity to absorb and distribute loads and to present a low-friction surface (18). Chondrocytes, the only cell type present in cartilage, have very low metabolic activity and are assumed to maintain the extracellular matrix by a low turnover replacement of matrix proteins. During OA development the chondrocytes become "activated", a process which is characterized by cell proliferation and cluster formation. A disturbed equilibrium develops in which the rate of loss of collagens and proteoglycans goes beyond the rate of the deposition of newly synthesized molecules (19).



Figure 2. Schematic overview of the normal and OA knee joint depicting the joint tissues affected in osteoarthritis (17)

1.2.2 Synovial tissue

Synovial tissue is in direct contact with articular cartilage and primarily maintains the synovial cavity and synthesizes synovial fluid. Synovial fluid has a lubricating function and facilitates a smooth movement between joints. Under normal conditions, synovial tissue consists of only 1-2 cell layers of macrophage-like (type A) synoviocytes and fibroblast-like (FLS or type B) synoviocytes (20). The inflammation of the synovium (synovitis) has been shown to occur in a number of OA patients and may produce proteases and cytokines that contribute to the disease (21). It has been suggested that activated synovial macrophages play a key role in the processes leading to synovial inflammation. This inflammation may act as a trigger for several symptoms of OA via release of soluble mediators by synovial tissue, thus contributing to the breakdown of cartilage by promoting destruction and impairing the ability of repair (22).

1.2.3 Infrapatellar fat pad

The knee joint contains a special form of adipose tissue named the infrapatellar fat pad (IPFP), which until recently did not receive a lot of attention as a contributor to the OA process. IPFP is located intra-capsularly and extra-synovially in the knee joint, and is in close contact with synovial layers and articular cartilage. Its main role is to facilitate the distribution of synovial fluid and to absorb forces through the knee (23). Considering its location and with regard to OA as a multiple tissue disease it is likely that IPFP is also involved in the pathogenesis of OA (24). Several soluble mediators are locally produced in the knee joint by the IPFP (25, 26). However, the precise role of the IPFP still needs to be elucidated.

1.2.4 Bone, meniscus and ligaments

Subchondral bone, meniscus and ligaments are described to be involved in OA pathogenesis. During OA, bone remodeling takes place as a result of altered joint homeostasis. Hereby, new bone at the joint margins (osteophytes) is formed. In addition, there is evidence available suggesting a link between bone sclerosis and modifications in bone mineralization and the progression of OA. A role for transforming growth factor (TGF)- β , which for example is produced by synoviocytes, is appreciated

in the development of osteophytes (27, 28). Whether subchondral bone sclerosis precedes the onset of OA or is a change that occurs parallel to cartilage degradation is unknown (16, 29). Furthermore, a role for the meniscus and surrounding ligaments is proposed. Meniscal damage occurs in 63% of adults with symptomatic knee OA (30), and it was shown that it leads to a 7.4 times higher chance to develop radiographic knee OA 30 months later (31). These data suggest that bone and meniscus need to be incorporated in the search for new OA targets.

1.2.5 Local Inflammation

Although OA is conventionally not considered as an inflammatory disease, the production of several inflammatory soluble mediators by different tissues in the knee joint suggests that inflammation has a more important role in affecting cartilage homeostasis than originally thought. Until now the most studied cytokines in OA are interleukin (IL)-1 β and tumor necrosis factor (TNF)- α . These cytokines are likely to be produced by articular chondrocytes, but synoviocytes may also very well be the source. In clinical trials attempting to block their activity, however, only minimal efficacy was found (32, 33). Both cytokines are mediated through the nuclear factor kappa B (NFKB) cascade, which is a pathway designated to have an important role in OA pathogenesis and a central regulator in catabolic actions in chondrocytes (34). Also, these cytokines are able to initiate other cytokines such as IL-8, IL-6, monocyte chemoattractant protein (MCP)-1 and RANTES (regulated on activation, normal T-cell expressed and secreted), which drive inflammation, inhibit matrix synthesis and promote cellular apoptosis by affecting, for example, aggrecanase (ADAMTS; a disintegrin and metalloproteinase with thrombospondin motifs) and collagenase activities (MMPs; matrix metalloproteinases) (22, 29, 35).

A special class of inflammatory mediators are oxylipins. These mediators are derived from fatty acids and can be found in all tissues in the body and may be interesting targets in relation to OA initiation and perpetuation. A balanced level of oxylipins is essential in maintaining joint homeostasis and alterations have widespread consequences. Prostaglandins are an example of such oxylipins and they have shown to be inhibited by non-steroidal anti-inflammatory drugs (NSAIDs), which leads to reduced OA pain. The precise role of other oxylipins, however, is unknown and merits more extensive research (36-38).

Exploring the role of the different oxylipins and soluble mediators involved and produced by several tissues in the joint, such as the synovial tissue and the IPFP, may lead to potential targets for disease-modifying interventions.

1.3 Systemic alterations; from mechanical to systemic disease

For several decades OA was regarded as a joint disease complicated by mechanical factors and age-related modifications. Limited attention to other factors influencing the disease was given. With the current knowledge, it is widely accepted that OA is more than only a mechanical disease and may be seen as a systemic disorder of multifactorial origin wherein genetic, environmental, hormonal and metabolic factors interact and contribute to OA pathogenesis (39). The metabolic syndrome comprises a profile including a combination of being obese, hypertension, dyslipidemia and impaired glucose tolerance (40). Systemic alterations may be induced by components of the metabolic syndrome. Recently, various studies presented a relation between OA and the prevalence of metabolic syndrome is over twofold higher in the OA population (41). In addition, having OA is associated with an over 5 times increased risk of having metabolic syndrome (42). These results have led investigators to contemplate common underlying pathologies in OA and metabolic syndrome related diseases (39, 43, 44).

1.3.1 Obesity

Obesity is an important and strong risk factor for OA and one of the components of the metabolic syndrome (45, 46). Obesity is becoming an increasing problem in the western world. In the Netherlands, 11 % of the adult population is obese (body mass index (BMI) >30) (47). For long, it was thought that the mechanical forces induced by obesity could explain the association between OA and obesity. However, it has been demonstrated that obesity is also associated with hand OA (48). As we do not walk on our hands, this suggests that systemic factors induced by obesity contribute considerably to the initiation and progression of OA (49). Obesity is associated with a mild chronic inflammation and the adipokines secreted by adipocytes and

macrophages within adipose tissue are suggested to be a metabolic link between obesity and OA (50). The relative contribution of these processes in the onset and progression of OA, however, remains unclear.

The association between obesity and the development of OA has been studied in several animal models. In the early 50's it was already discovered that mice receiving a high fat diet (HFD) developed features of OA twice as fast compared to mice fed a normal diet (figure 3) (51). In addition, mice of the STR/ort strain, which are susceptible to develop spontaneous obesity and certain aspects of the human metabolic syndrome, develop OA in a short period of time (52). Furthermore, it has been demonstrated that mice receiving HFD showed more OA cartilage degeneration than those fed a normal diet in a post-traumatic mouse model (53). A very HFD in mice induces OA, but when animals are placed on a wheel-running exercise plan progression of knee OA is inhibited without reduced body fat (54). Furthermore, leptin-deficient (ob/ob) and leptin receptor-deficient (db/db) mice, which have extremely high body weight on a normal diet, are protected from the development of OA (55), which excludes a role for mechanical factors. The fact that adipose tissue of mice can become inflamed under conditions of metabolic stress (e.g. HFD feeding) and secrete a broad spectrum of inflammatory mediators (56, 57), suggests that HFDfed mice constitute a suitable model to study the role of (metabolic stress-induced) inflammation on the development of OA.



Figure 3. Male C57/Bl6 mice receiving a high fat diet developed osteoarthritis (OA) twice as fast as mice receiving a control diet. The similar slopes indicate that a high fat diet accelerates the onset but not the progression of OA (51).

1.3.2 Cholesterol

Hypercholesterolemia is associated with the metabolic syndrome. Increased levels of cholesterol in the plasma have shown to be associated with generalized OA (58). Furthermore, it has been demonstrated that genes regulating cholesterol efflux have a diminished expression in OA cartilage compared to normal cartilage (59). The underlying mechanism explaining the link between cholesterol and OA features are therefore very interesting. This is supported by the fact that atherosclerosis, which is an atheromatous vascular disease caused by elevated cholesterol intake, is independently associated with OA in women (60, 61). Whether atherosclerosis and OA have common underlying mechanisms or are causative for each other is uncertain.

With respect to animal models, the effects of hypercholesterolemia on OA development is difficult to examine, as wild type mice have a different lipoprotein metabolism compared to humans. The intake of a cholesterol-containing diet by wild type mice leads to elevated plasma cholesterol levels. However, as a result of high levels of anti-atherosclerotic high density lipoprotein (HDL) ('good' cholesterol) and low levels of very low (VL)DL and low (L)DL ('bad' cholesterol) no atherosclerosis develops. Experimental data on investigating the role of cholesterol, and the mechanism behind it in relation to OA development, are therefore scarce and more research needs to be done to elaborate on this association.

1.3.3 Systemic inflammation

The metabolic syndrome induces systemic inflammatory responses. This mechanism of action may also be held responsible for the association of metabolic syndrome with OA, either by a direct effect on articular cartilage, or by the modulation of several tissues in and around the joint. The precise role of inflammation in OA development is, as mentioned earlier, uncertain (chapter 1.2.4) (22, 62). Among the components of the metabolic syndrome, obesity was thought to be key initiator for OA. However, recently published epidemiologic data demonstrated that the metabolic syndrome rather than obesity in itself has the greatest impact on the severity of OA (63). OA donors showed several changed systemic mediators in their serum compared to those obtained from healthy persons (64, 65). Of these

mediators the adipokines, such as leptin, adiponectin, resistin and visfatin, have been extensively studied for their pro- and anti-inflammatory capacities in OA (50). It is found that serum adiponectin, leptin, and resistin concentrations are associated with OA severity and progression and with local synovial tissue inflammation (66-68). The effect of adipokines on the OA process is controversial. Leptin is thought to play a role in TGF- β activation (69) and, as mentioned, extremely obese leptin-impaired mice are protected from the development of OA (55). In addition, leptin induces collagen release from bovine cartilage explants and upregulates MMP-1 and MMP-13 in bovine chondrocytes (70). It has been demonstrated that adiponectin alters the balance by downregulating MMPs and upregulating TIMPs. However, it is also believed that it may act protective against OA by reducing the production of proinflammatory cytokines (71). Adiponectin-treated chondrocytes produce IL-6, MMP-3 and MMP-9 and this adipokine induces MMP-1 and IL-6 production in synovial fibroblasts (72, 73). A relatively newly discovered adipokine, visfatin, has been suggested as a promising target for treating OA. Visfatin synthesis is increased by IL-1β treatment in an *in vitro* culture of human chondrocytes. Furthermore, visfatin increases the synthesis and release of MMP-3, MMP-13, ADAMTS-4, and ADAMTS-5 and decreases aggrecan production in chondrocytes, suggesting a pro-inflammatory function in cartilage (74, 75). With regard to cholesterol, it is postulated that oxidized low density lipoprotein triggers pro-inflammatory cytokine production (76). It should be noted that these mediators are not solely originating from the blood and that synovial tissue (21) and IPFP (24) secrete a number of these inflammatory factors as well, which may contribute to OA. Whether targeting the inflammatory response induced by components of the metabolic syndrome, such as adipokines, will reduce OA development is unknown. The integration of knowledge of the metabolic syndrome and related diseases in OA research provides new leads to tackle the underlying mechanisms responsible for OA pathogenesis.

1.4 Outline of the thesis

The aim of this thesis is to provide insight in the role of multiple local and systemic factors contributing to the pathogenesis of OA. With the current knowledge, more advanced technologies, and novel animal models, we should be able to shed a different light on the tissues involved in OA pathogenesis. Hopefully, this will ultimately lead to the identification of mechanisms that provide targets for disease-modifying therapies and novel biomarkers to detect OA in an earlier phase. With respect to the future, we try to contribute to a rationale for a better stratification of OA patients and consequently to more personalized medicine.

The first three chapters include a description of local factors that possibly contribute to OA development. By investigating the secretion of several mediators by synovial tissue, IPFP and synovial fluid, a profile of OA and normal (non-OA) donors is provided.

In **Chapter 2** we aimed to get insight in the role of synovial tissue. We hypothesized that synovial tissue derived from end-stage OA patients is more inflamed than that from normal donors (21). The synovial tissue of OA donors may therefore secrete a wide array of factors that can alter cartilage degradation, which is not the case for synovial tissues derived from normal donors. To test this hypothesis, we used a multiplex approach (an advanced technology including multiplex bead-based immunoassays (77)). In addition, the ability of OA and normal synovial tissues to initiate degeneration of healthy cartilage was assessed by culturing synovial tissues and healthy cartilage explants in a complex *in vitro* co-culture transwell system.

The purpose of **Chapter 3** was to explore the role of IPFP in OA. Oxylipins, important signaling molecules involved in the modulation of inflammatory responses (78, 79), were detected using a mass spectrometry method to get an overview of what is secreted by the IPFP. Furthermore, differences in IPFP secretion between OA and normal donors were evaluated.

We profiled synovial fluid samples as representatives of the secretion of soluble mediators by different tissues involved in the joint (**Chapter 4**). Again we used a multiplex approach to get a wide and comprehensive overview of OA and normal donors. Data were further explored using multi-variate statistics to identify clusters of interrelated mediators.

Besides the involvement of local factors we examined the development of OA with regard to alterations on a systemic level. Considering the hypothesis that obesity might lead to metabolic stress, which possibly contributes to the development of OA (62), we used a HFD-induced OA mouse model which is described in **Chapter 5**. More specifically, we used the human C-reactive protein (hCRP) transgenic mice (80). CRP is an acute phase protein and an established marker for systemic inflammation in humans (81). Consequently, we were able to monitor the inflammatory state of the animals, using a human marker, during the development of OA. To perceive whether medicines applied for other targets in the metabolic syndrome were able to counteract OA-related changes, we included a prophylactic intervention with statins (cholesterol-lowering drug) and a peroxisome proliferator activated receptor (PPAR) agonist (anti-diabetic drug).

In **Chapter 6** we emphasized on lipid and systemic alterations induced by elevated cholesterol intake instead of HFD. We investigated whether a cholesterol-containing diet and, consequently, the development of atherosclerosis was sufficient to induce OA in APOE*3 Leiden.CETP transgenic mice. This is a well-established model for hyperlipidemia and atherosclerosis and resembles the human lipoprotein metabolism in contrary to wild type mice (82-84). Furthermore, we included cholesterol-lowering interventions, with a statin and ezetimibe, in groups receiving a high cholesterol diet, to assess their effects on OA development. Both drugs diminish cholesterol levels to a same extent but have a different mode of action, which allowed us to investigate possible additional effects of statin beyond its cholesterol-lowering capacities.

To elaborate more on the outcomes of chapters 5 and 6, in **Chapter 7** we extrapolated the HFD-induced OA mouse model to the APOE*3 Leiden.CETP mouse. We included an early ('prophylactic') and late ('therapeutic') intervention with a statin. Furthermore, we included a fenofibrate (cholesterol-lowering drug) intervention, since another study indicated that this type of drug has beneficial effects on OA development. To gain more insight into the mechanisms behind the HFD-induced OA model, the effect of a caspase-1 inhibitor, responsible for the conversion of pro-IL-1 to IL-1 (one of the key cytokines believed to be involved in OA), was studied. In addition, a group that started on HFD, but returned to control chow diet halfway the study, was included to evaluate the effect of a diet switch on OA development.

In the final chapters, **Chapter 8** and **Chapter 9** (in Dutch), a comprehensive overview of the performed work is given and discussed.

References

- 1. Buchanan WW. William Hunter (1718-1783). Rheumatology 2003;42:1260-1.
- 2. Woolf AD, Pfleger B. Burden of major musculoskeletal conditions. Bull World Health Organ 2003;81:646-56.
- van Saase JL, van Romunde LK, Cats A, Vandenbroucke JP, Valkenburg HA. Epidemiology of osteoarthritis: Zoetermeer survey. Comparison of radiological osteoarthritis in a Dutch population with that in 10 other populations. Ann Rheum Dis 1989;48:271-80.
- Lawrence RC, Hochberg MC, Kelsey JL, McDuffie FC, Medsger TA, Jr, Felts WR, et al. Estimates of the prevalence of selected arthritic and musculoskeletal diseases in the United States. J Rheumatol 1989;16:427-41.
- Lopez AD, Mathers CD, Ezzati M, Jamison DT, Murray CJ. Global and regional burden of disease and risk factors, 2001: systematic analysis of population health data. Lancet 2006;367:1747-57.
- Poos MJJC, Gommer AM, Zantinge EM, Uiters E. Hoe vaak komt artrose voor en hoeveel mensen sterven eraan?. Volksgezondheid Toekomst Verkenning, Nationaal Kompas Volksgezondheid 2009.
- 7. Kellgren JH, Lawrence JS. Radiological assessment of osteo-arthrosis. Ann Rheum Dis 1957;16:494-502.
- Altman R, Alarcon G, Appelrouth D, Bloch D, Borenstein D, Brandt K, et al. The American College of Rheumatology criteria for the classification and reporting of osteoarthritis of the hip. Arthritis Rheum 1991;34:505-14.
- Altman R, Alarcon G, Appelrouth D, Bloch D, Borenstein D, Brandt K, et al. The American College of Rheumatology criteria for the classification and reporting of osteoarthritis of the hand. Arthritis Rheum 1990;33:1601-10.
- 10. Altman R, Asch E, Bloch D, Bole G, Borenstein D, Brandt K, et al. Development of criteria for the classification and reporting of osteoarthritis. Classification of osteoarthritis of the knee. Diagnostic and Therapeutic Criteria Committee of the American Rheumatism Association. Arthritis Rheum 1986;29:1039-49.
- 11. Oliveria SA, Felson DT, Reed JI, Cirillo PA, Walker AM. Incidence of symptomatic hand, hip, and knee osteoarthritis among patients in a health maintenance organization. Arthritis Rheum 1995;38:1134-41.
- 12. Arden N, Nevitt MC. Osteoarthritis: epidemiology. Best Pract Res Clin Rheumatol 2006;20:3-25.
- 13. Hunter DJ, Felson DT. Osteoarthritis. BMJ 2006;332:639-42.
- Bauer DC, Hunter DJ, Abramson SB, Attur M, Corr M, Felson D, et al. Classification of osteoarthritis biomarkers: a proposed approach. Osteoarthritis Cartilage 2006;14:723-7.
- 15. Brandt KD, Radin EL, Dieppe PA, van de Putte L. Yet more evidence that osteoarthritis is not a cartilage disease. Ann Rheum Dis 2006;65:1261-4.
- 16. Loeser RF, Goldring SR, Scanzello CR, Goldring MB. Osteoarthritis: a disease of the joint as an organ. Arthritis Rheum 2012;64:1697-707.
- 17. Hunter DJ. Osteoarthritis. Best Pract Res Clin Rheumatol 2011;25:801-14.
- Ulrich-Vinther M, Maloney MD, Schwarz EM, Rosier R, O'Keefe RJ. Articular cartilage biology. J Am Acad Orthop Surg 2003;11:421-30.
- 19. Goldring MB, Marcu KB. Cartilage homeostasis in health and rheumatic diseases. Arthritis Res Ther 2009;11:224.
- 20. Iwanaga T, Shikichi M, Kitamura H, Yanase H, Nozawa-Inoue K. Morphology and functional roles of synoviocytes in the joint. Arch Histol Cytol 2000;63:17-31.

- 21. de Lange-Brokaar BJ, Ioan-Facsinay A, van Osch GJ, Zuurmond AM, Schoones J, Toes RE, et al. Synovial inflammation, immune cells and their cytokines in osteoarthritis: a review. Osteoarthritis Cartilage 2012;20:1484-99.
- 22. Goldring MB, Otero M. Inflammation in osteoarthritis. Curr Opin Rheumatol 2011;23:471-8.
- 23. Gallagher J, Tierney P, Murray P, O'Brien M. The infrapatellar fat pad: anatomy and clinical correlations. Knee Surg Sports Traumatol Arthrosc 2005;13:268-72.
- 24. Clockaerts S, Bastiaansen-Jenniskens YM, Runhaar J, Van Osch GJ, Van Offel JF, Verhaar JA, et al. The infrapatellar fat pad should be considered as an active osteoarthritic joint tissue: a narrative review. Osteoarthritis Cartilage 2010;18:876-82.
- 25. Klein-Wieringa IR, Kloppenburg M, Bastiaansen-Jenniskens YM, Yusuf E, Kwekkeboom JC, El-Bannoudi H, et al. The infrapatellar fat pad of patients with osteoarthritis has an inflammatory phenotype. Ann Rheum Dis 2011;70:851-7.
- 26. Ushiyama T, Chano T, Inoue K, Matsusue Y. Cytokine production in the infrapatellar fat pad: another source of cytokines in knee synovial fluids. Ann Rheum Dis 2003;62:108-12.
- Bakker AC, van de Loo FA, van Beuningen HM, Sime P, van Lent PL, van der Kraan PM, et al. Overexpression of active TGF-beta-1 in the murine knee joint: evidence for synoviallayer-dependent chondro-osteophyte formation. Osteoarthritis Cartilage 2001;9:128-36.
- 28. Uchino M, Izumi T, Tominaga T, Wakita R, Minehara H, Sekiguchi M, et al. Growth factor expression in the osteophytes of the human femoral head in osteoarthritis. Clin Orthop Relat Res 2000;377:119-25.
- 29. Abramson SB, Attur M. Developments in the scientific understanding of osteoarthritis. Arthritis Res Ther 2009;11:227.
- Englund M, Guermazi A, Gale D, Hunter DJ, Aliabadi P, Clancy M, et al. Incidental meniscal findings on knee MRI in middle-aged and elderly persons. N Engl J Med 2008;359:1108-15.
- Englund M, Guermazi A, Roemer FW, Aliabadi P, Yang M, Lewis CE, et al. Meniscal tear in knees without surgery and the development of radiographic osteoarthritis among middle-aged and elderly persons: The Multicenter Osteoarthritis Study. Arthritis Rheum 2009;60:831-9.
- Magnano MD, Chakravarty EF, Broudy C, Chung L, Kelman A, Hillygus J, et al. A pilot study of tumor necrosis factor inhibition in erosive/inflammatory osteoarthritis of the hands. J Rheumatol 2007;34:1323-7.
- 33. Cohen SB, Proudman S, Kivitz AJ, Burch FX, Donohue JP, Burstein D, et al. A randomized, double-blind study of AMG 108 (a fully human monoclonal antibody to IL-1R1) in patients with osteoarthritis of the knee. Arthritis Res Ther 2011;13:R125.
- 34. Marcu KB, Otero M, Olivotto E, Borzi RM, Goldring MB. NF-kappaB signaling: multiple angles to target OA. Curr Drug Targets 2010;11:599-613.
- 35. Pelletier JP, Martel-Pelletier J, Abramson SB. Osteoarthritis, an inflammatory disease: potential implication for the selection of new therapeutic targets. Arthritis Rheum 2001;44:1237-47.
- 36. Laufer S. Role of eicosanoids in structural degradation in osteoarthritis. Curr Opin Rheumatol 2003;15:623-7.
- 37. Haeggstrom JZ, Funk CD. Lipoxygenase and leukotriene pathways: biochemistry, biology, and roles in disease. Chem Rev 2011;111:5866-98.
- 38. Molloy ES, McCarthy GM. Eicosanoids, osteoarthritis, and crystal deposition diseases. Curr Opin Rheumatol 2005;17:346-50.
- Katz JD, Agrawal S, Velasquez M. Getting to the heart of the matter: osteoarthritis takes its place as part of the metabolic syndrome. Curr Opin Rheumatol 2010;22:512-9.

- 40. Day C. Metabolic syndrome, or What you will: definitions and epidemiology. Diab Vasc Dis Res 2007;4:32-8.
- 41. Sowers M, Karvonen-Gutierrez CA, Palmieri-Smith R, Jacobson JA, Jiang Y, Ashton-Miller JA. Knee osteoarthritis in obese women with cardiometabolic clustering. Arthritis Rheum 2009;61:1328-36.
- 42. Puenpatom RA, Victor TW. Increased prevalence of metabolic syndrome in individuals with osteoarthritis: an analysis of NHANES III data. Postgrad Med 2009;121:9-20.
- 43. Velasquez MT, Katz JD. Osteoarthritis: another component of metabolic syndrome?. Metab Syndr Relat Disord 2010;8:295-305.
- 44. Masuko K, Murata M, Suematsu N, Okamoto K, Yudoh K, Nakamura H, et al. A metabolic aspect of osteoarthritis: lipid as a possible contributor to the pathogenesis of cartilage degradation. Clin Exp Rheumatol 2009;27:347-53.
- 45. Felson DT, Anderson JJ, Naimark A, Walker AM, Meenan RF. Obesity and knee osteoarthritis. The Framingham Study. Ann Intern Med 1988;109:18-24.
- 46. Blagojevic M, Jinks C, Jeffery A, Jordan KP. Risk factors for onset of osteoarthritis of the knee in older adults: a systematic review and meta-analysis. Osteoarthritis Cartilage 2010;18:24-33.
- 47. Rijksinstituut voor Volksgezondheid en Milieu (Dutch National Institute for Public Health and the Environment). Zorgbalans 2010.
- 48. Yusuf E, Nelissen RG, Ioan-Facsinay A, Stojanovic-Susulic V, DeGroot J, van Osch G, et al. Association between weight or body mass index and hand osteoarthritis: a systematic review. Ann Rheum Dis 2010;69:761-5.
- 49. Yusuf E. Metabolic factors in osteoarthritis: obese people do not walk on their hands. Arthritis Res Ther 2012;14:123.
- Gomez R, Conde J, Scotece M, Gomez-Reino JJ, Lago F, Gualillo O. What's new in our understanding of the role of adipokines in rheumatic diseases?. Nat Rev Rheumatol 2011;7:528-36.
- 51. Silberberg M, Silberberg R. Degenerative joint disease in mice fed a high-fat diet at various ages. Exp Med Surg 1952;10:76-87.
- 52. Sokoloff L, Mickelsen O, Silverstein E, Jay GE,Jr, Yamamoto RS. Experimental obesity and osteoarthritis. Am J Physiol 1960;198:765-70.
- 53. Louer CR, Furman BD, Huebner JL, Kraus VB, Olson SA, Guilak F. Diet-induced obesity significantly increases the severity of posttraumatic arthritis in mice. Arthritis Rheum 2012;64:3220-30.
- 54. Griffin TM, Huebner JL, Kraus VB, Yan Z, Guilak F. Induction of osteoarthritis and metabolic inflammation by a very high-fat diet in mice: effects of short-term exercise. Arthritis Rheum 2012;64:443-53.
- 55. Griffin TM, Huebner JL, Kraus VB, Guilak F. Extreme obesity due to impaired leptin signaling in mice does not cause knee osteoarthritis. Arthritis Rheum 2009;60:2935-44.
- 56. Weisberg SP, Hunter D, Huber R, Lemieux J, Slaymaker S, Vaddi K, et al. CCR2 modulates inflammatory and metabolic effects of high-fat feeding. J Clin Invest 2006;116:115-24.
- 57. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW, Jr. Obesity is associated with macrophage accumulation in adipose tissue. J Clin Invest 2003;112:1796-808.
- Sturmer T, Sun Y, Sauerland S, Zeissig I, Gunther KP, Puhl W, et al. Serum cholesterol and osteoarthritis. The baseline examination of the Ulm Osteoarthritis Study. J Rheumatol 1998;25:1827-32.
- 59. Tsezou A, Iliopoulos D, Malizos KN, Simopoulou T. Impaired expression of genes regulating cholesterol efflux in human osteoarthritic chondrocytes. J Orthop Res 2010;28:1033-9.

- Hoeven TA, Kavousi M, Clockaerts S, Kerkhof HJ, van Meurs JB, Franco O, et al. Association of atherosclerosis with presence and progression of osteoarthritis: the Rotterdam Study. Ann Rheum Dis 2013;72:646-51.
- 61. Jonsson H, Helgadottir GP, Aspelund T, Eiriksdottir G, Sigurdsson S, Ingvarsson T, et al. Hand osteoarthritis in older women is associated with carotid and coronary atherosclerosis: the AGES Reykjavik study. Ann Rheum Dis 2009;68:1696-700.
- 62. Berenbaum F. Osteoarthritis as an inflammatory disease (osteoarthritis is not osteoarthrosis!). Osteoarthritis Cartilage 2013;21:16-21
- 63. Yoshimura N, Muraki S, Oka H, Tanaka S, Kawaguchi H, Nakamura K, et al. Accumulation of metabolic risk factors such as overweight, hypertension, dyslipidaemia, and impaired glucose tolerance raises the risk of occurrence and progression of knee osteoarthritis: a 3-year follow-up of the ROAD study. Osteoarthritis Cartilage 2012;20:1217-26.
- 64. Attur M, Krasnokutsky-Samuels S, Samuels J, Abramson SB. Prognostic biomarkers in osteoarthritis. Curr Opin Rheumatol 2013;25:136-44.
- 65. Fernandez-Puente P, Mateos J, Fernandez-Costa C, Oreiro N, Fernandez-Lopez C, Ruiz-Romero C, et al. Identification of a panel of novel serum osteoarthritis biomarkers. J Proteome Res 2011;10:5095-101.
- 66. Filkova M, Liskova M, Hulejova H, Haluzik M, Gatterova J, Pavelkova A, et al. Increased serum adiponectin levels in female patients with erosive compared with non-erosive osteoarthritis. Ann Rheum Dis 2009;68:295-6.
- 67. de Boer TN, van Spil WE, Huisman AM, Polak AA, Bijlsma JW, Lafeber FP, et al. Serum adipokines in osteoarthritis; comparison with controls and relationship with local parameters of synovial inflammation and cartilage damage. Osteoarthritis Cartilage 2012;20:846-53.
- Yusuf E, Ioan-Facsinay A, Bijsterbosch J, Klein-Wieringa I, Kwekkeboom J, Slagboom PE, et al. Association between leptin, adiponectin and resistin and long-term progression of hand osteoarthritis. Ann Rheum Dis 2011;70:1282-4.
- Kumpers P, Gueler F, Rong S, Mengel M, Tossidou I, Peters I, et al. Leptin is a coactivator of TGF-beta in unilateral ureteral obstructive kidney disease. Am J Physiol Renal Physiol 2007;293:F1355-62.
- 70. Hui W, Litherland GJ, Elias MS, Kitson GI, Cawston TE, Rowan AD, et al. Leptin produced by joint white adipose tissue induces cartilage degradation via upregulation and activation of matrix metalloproteinases. Ann Rheum Dis 2012;71:455-62.
- 71. Chen TH, Chen L, Hsieh MS, Chang CP, Chou DT, Tsai SH. Evidence for a protective role for adiponectin in osteoarthritis. Biochim Biophys Acta 2006;1762:711-8.
- 72. Lago R, Gomez R, Otero M, Lago F, Gallego R, Dieguez C, et al. A new player in cartilage homeostasis: adiponectin induces nitric oxide synthase type II and pro-inflammatory cytokines in chondrocytes. Osteoarthritis Cartilage 2008;16:1101-9.
- 73. Tang CH, Chiu YC, Tan TW, Yang RS, Fu WM. Adiponectin enhances IL-6 production in human synovial fibroblast via an AdipoR1 receptor, AMPK, p38, and NF-kappa B pathway. J Immunol 2007;179:5483-92.
- 74. Gosset M, Berenbaum F, Salvat C, Sautet A, Pigenet A, Tahiri K, et al. Crucial role of visfatin/pre-B cell colony-enhancing factor in matrix degradation and prostaglandin E2 synthesis in chondrocytes: possible influence on osteoarthritis. Arthritis Rheum 2008;58:1399-409.
- 75. Hu PF, Bao JP, Wu LD. The emerging role of adipokines in osteoarthritis: a narrative review. Mol Biol Rep 2011;38:873-8.
- 76. Tedgui A, Mallat Z. Cytokines in atherosclerosis: pathogenic and regulatory pathways. Physiol Rev 2006;86:515-81.
- 77. Leng SX, McElhaney JE, Walston JD, Xie D, Fedarko NS, Kuchel GA. ELISA and multiplex technologies for cytokine measurement in inflammation and aging research. J Gerontol A Biol Sci Med Sci 2008;63:879-84.

- 78. Lewis RA. Interactions of eicosanoids and cytokines in immune regulation. Adv Prostaglandin Thromboxane Leukot Res 1990;20:170-8.
- 79. Tilley SL, Coffman TM, Koller BH. Mixed messages: modulation of inflammation and immune responses by prostaglandins and thromboxanes. J Clin Invest 2001;108:15-23.
- Verschuren L, Wielinga PY, van Duyvenvoorde W, Tijani S, Toet K, van Ommen B, et al. A dietary mixture containing fish oil, resveratrol, lycopene, catechins, and vitamins E and C reduces atherosclerosis in transgenic mice. J Nutr 2011;141:863-9.
- 81. Ciliberto G, Arcone R, Wagner EF, Ruther U. Inducible and tissue-specific expression of human C-reactive protein in transgenic mice. EMBO J 1987;6:4017-22.
- Westerterp M, van der Hoogt CC, de Haan W, Offerman EH, Dallinga-Thie GM, Jukema JW, et al. Cholesteryl ester transfer protein decreases high-density lipoprotein and severely aggravates atherosclerosis in APOE*3-Leiden mice. Arterioscler Thromb Vasc Biol 2006;26:2552-9.
- van der Hoorn JW, de Haan W, Berbee JF, Havekes LM, Jukema JW, Rensen PC, et al. Niacin increases HDL by reducing hepatic expression and plasma levels of cholesteryl ester transfer protein in APOE*3Leiden.CETP mice. Arterioscler Thromb Vasc Biol 2008;28:2016-22.
- 84. Kuhnast S, van der Hoorn JWA, van den Hoek AM, Havekes LM, Liau G, Jukema JW, et al. Aliskiren inhibits atherosclerosis development and improves plaque stability in APOE*3Leiden.CETP transgenic mice with or without treatment with atorvastatin. J Hypertens 2012;30:107-16.