

Leukocytes and complement in atherosclerosis Alipour, A.

Citation

Alipour, A. (2012, February 9). *Leukocytes and complement in atherosclerosis*. Retrieved from https://hdl.handle.net/1887/18459

Version:	Corrected Publisher's Version
License:	<u>Licence agreement concerning inclusion of doctoral</u> <u>thesis in the Institutional Repository of the University</u> <u>of Leiden</u>
Downloaded from:	https://hdl.handle.net/1887/18459

Note: To cite this publication please use the final published version (if applicable).

Mannose binding lectin 2 haplotypes do not affect the progression of coronary atherosclerosis in men with proven coronary artery disease treated with pravastatin

A. Alipour¹, M. Castro Cabezas¹, J.W.F. Elte¹, J.C. Vallvé², J. Ribalta², A.H. Zwinderman^{3,4}, J.C. Defesche⁵, J.W. Jukema^{4,6}

¹Department of Internal Medicine, Sint Franciscus Gasthuis, Rotterdam, The Netherlands ²Unitat de Recerca de Lípids i Arteriosclerosi, Facultat de Medicina, Universitat Rovira i Virgili, CIBER de Diabetes y Enfermedades Metabólicas Asociadas (CIBERDEM), Reus, Spain ³Department of Medical Statistics, Academic Medical Center, Amsterdam, The Netherlands ⁴Interuniversity Cardiology Institute of the Netherlands and Durrer Center for Cardiogenetic Research, Utrecht, The Netherlands

⁵Department of Vascular Medicine, Academic Medical Centre, University of Amsterdam, The Netherlands

⁶Department of Cardiology, Leiden University Medical Center, Leiden, The Netherlands

Atherosclerosis 2011;215:125-129

ABSTRACT

Introduction: Mannose binding lectin (MBL) is one of the three initiators of complement activation. Polymorphisms of the MBL2 gene and its promoter, and especially haplotypes, determine MBL plasma levels. MBL deficiency has been associated with the development of atherosclerosis. We evaluated whether the rate of angiographic progression of coronary atherosclerosis during pravastatin treatment was associated with MBL2 haplotypes in REGRESS, a placebo-controlled 2 years intervention study.

Methods: Three polymorphic sites in exon 1 (rs1800450, rs1800451, and rs5030737) of the MBL2 gene and 2 sites (rs7096206, and rs11003125) in the promoter region were genotyped in 398 subjects. Genotyping was performed using Applied Biosystems[®] TaqMan[®] Genotyping Assays. We divided the group in high, intermediate and low MBL2 secretor haplotypes. Quantitative coronary angiography was performed. Endpoints were mean segment diameter (MSD) and minimum obstruction diameter (MOD) established by quantitative coronary angiography. **Results:** At inclusion, 50.1, 31.7 and 17.6% of the patients in the REGRESS cohort carried the high, intermediate and low MBL2 secretor haplotype was not informative. There were no baseline differences between the MBL2 haplotypes for age, BMI, lipid levels, leukocyte counts, CRP, MSD and MOD. The intermediate MBL2 placebo group showed the greatest increase in MSD compared to the low MBL2 group (P=0.03). No difference was found for the change in MOD. No significant interaction between MBL2 haplotype groups and pravastatin therapy was observed.

Conclusions: In men with proven coronary artery disease, MBL2 secretor haplotypes are not associated to the rate of progression of coronary sclerosis nor does pravastatin treatment influence progression based on MBL2 haplotypes.

INTRODUCTION

Inflammation is closely associated to atherosclerosis (1-2). In the past few years several pro-inflammatory genes have been identified, which have been suggested to play a role in atherosclerosis. One of these genes is the Mannose Binding Lectin (MBL) gene. MBL deficiency has been associated to coronary artery disease (CAD), increased intima-media thickness in carotid arteries (3-5) and atherosclerosis in different clinical situations (6-8). Recent work from our laboratory shows that MBL deficiency may lead to a disturbed metabolism of postprandial VLDL1 lipoproteins (9), potentially favoring atherosclerosis.

MBL is an important activating factor of the lectin pathway of the complement system (10,11). The MBL2 gene codes for the active MBL protein and has three known mutations: allele B at codon 54 (G54D, rs1800450), allele C at codon 57 (G57E, rs1800450) and allele D at codon 52 (R52C, rs5030737) (11,12). These mutations lead to structural abnormalities and cause MBL deficiency (11). The wild-type codon is allele A. Moreover, there are two mutations in the promoter region at –550 (H/L, rs11003125) and –221 (Y/X, rs7096206), which result in a decreased synthesis of the protein (12-14). The alleles interact with each other to form MBL2 'scretor haplotypes' producing high, intermediate and low MBL levels (12,13).

The role of MBL in atherosclerosis is unsettled since several recent studies have suggested that not only low, but also high MBL levels are associated with atherosclerosis (15-19).

Statins have been shown to reduce the atherosclerotic burden in different groups of patients (20-22) and are able to modulate complement components in normolipidemic subjects with CAD (23). Currently, no data are available on the role of MBL in the progression of coronary atherosclerosis in prospective intervention studies with statins. In this study, we aimed to investigate the relation between the MBL2 secretor haplotypes and the progression of coronary atherosclerosis in patients treated with pravastatin compared to placebo in the REGRESS (Regression Growth Evaluation Statin Study) population (20).

MATERIALS AND METHODS

Subjects

Samples collected in the REGRESS cohort were used (20). The REGRESS protocol has been described in detail elsewhere (20). In brief, REGRESS was designed as a double-blind, placebocontrolled, multicenter study to assess the effect of pravastatin treatment on the progression and regression of coronary atherosclerosis. All patients were men of Caucasian descent; they were <70 years of age, and had angiographically documented CAD (>50% stenosis of 1 major vessel). Patients who had unstable angina or who suffered a myocardial infarction within the preceding 6 months of the study were excluded; angina pectoris classification was based on the Rose questionnaire. All patients had total cholesterol levels between 4 and 8 mmol/L and TG levels <4 mmol/L.

The present analysis was approved by the Local Ethical Committee of the Sint Franciscus Gasthuis and the Medical Ethical committee of the Leiden University Medical Center.

Analytical methods

All lipid laboratory tests were carried out at the Lipid Reference Laboratory (Atlanta, Ga). Serum lipids, blood cell counts and high sensitive CRP (hsCRP) were measured in fasting blood samples by standard techniques and LDL was calculated according to the Friedewald formula (20).

Quantitative Coronary Arteriography

The quantitative coronary arteriographic procedures have been described in detail elsewhere and include the mean segment diameter (MSD) and minimum obstruction diameter (MOD) on a per patient basis (20).

DNA analysis and assignment of haplotype status

Genomic DNA was isolated from white cells via standard procedures, dissolved in 10 mmol/L Tris, 1 mmol/L EDTA, pH 8.0, and stored at 4°C. 3 polymorphic sites in exon 1 (rs1800450, rs1800451, and rs5030737) of the MBL2 gene and 2 sites (rs7096206, and rs11003125) in the promoter region were genotyped in 398 subjects. Genomic DNA samples were genotyped using TaqMan^{*} SNP Genotyping Assays specifics for each one of the polymorphic sites and according to the manufacturer's instructions. The assays were performed on a GeneAmp PCR system 9700 with TaqMan^{*} master mix (Applied Biosystems^{*}) and allelic discrimination was done on a 7900HT fast real-time PCR system (Applied Biosystems^{*}). To ensure consistency between runs, samples of known genotypes were repeated in every analysis.

Patients were considered to have the high MBL2 secretor haplotypes if they had the HYA and LYA haplotypes. The haplotypes LXA, HYD, HYBD and HXA were characterised as the intermediate MBL2 secretors and the haplotypes LYB and LYC as the low MBL2 secretors (12,13).

Statistics

We first checked whether the genotype distributions of the five MBL2-gene SNPs were in Hardy-Weinberg equilibrium using one-degree of freedom Chi-square tests. Subsequently, baseline patient characteristics were compared between genotypes using ANOVA, Chi-square tests, or Kruskal-Wallis tests where appropriate. MSD and MOD changes during the trial were compared between genotypes using ANOVA with baseline values as covariates, and the occurrence of cardiac events during the trial were analysed with the log rank test. Patients with distinctive haplotypes in the low (LYC) and intermediate (HYD, HYBD, HXA) MBL2 groups represented only a small portion of the total group. Therefore, the statistical analysis was carried out for the MBL2 secretor haplotypes, rather than the distinctive haplotypes in the three groups. We estimated individual haplotypes and the association between haplotypes and patient characteristics using Phase (24) and the algorithms developed by Souverein et al (25). Concerning power analysis, the current sample size provided at least 80% power to detect a significant association between MBL2 genotypes and MSD/MOD change if the mean difference is larger than about 0.3 standard deviations. This was the case for rs7096206, rs11003125 and rs1800450. Because the minor allele frequencies were lower, the mean difference should be larger than 0.46 for rs5030737 and larger than 0.76 for rs1800451. The standardized mean difference (or Cohen's effect size) is considered moderate between 0.2 and 0.5 and large > 0.5.

RESULTS

MBL2 distribution and baseline characteristics (Tables 1-3)

The number of REGRESS patients of whom DNA was available and with at least one genotyped MBL2 SNP was 398. The relative frequency of the SNPs is listed in Table 1. Only the rs7096206 and rs1800450 SNPs were not in Hardy-Weinberg equilibrium (Table 1). The distribution of the genotypes was similar with those reported in other studies (12,13).

The MBL2 haplotype distribution was also comparable to other studies (12,13) (Table 2). The haplotype HXD with a relative frequency of only 0.6±0.4% could not be divided in a category corresponding to one of the three MBL2 secretor levels. Therefore, we did not include the HXD haplotype in our further analysis. There were no differences between the different SNPs and haplotypes for baseline cardiovascular risk factors and the progression of CAD (Table 3).

Promotor region polymorphism		Frequency % (no)	HWE				
rs7096206	Х	22.1 (156)	0.01*				
	Y	77.9 (554)					
rs11003125	Н	37.2 (270)	0.51				
	L	62.8 (448)					
Structurally encoding polymorphism							
rs1800450	В	15.2 (111)	0.00*				
	W	84.8 (448)					
rs1800451	C	3.1 (21)	0.55				
	W	96.9 (655)					
rs5030737	D	9.6 (62)	0.18				
	W	90.4 (584)					

Table 1. MBL2 allele frequencies in the REGRESS cohort

HWE, Hardy-Weinberg equilibrium. * *P* < 0.05.

Corresponding MBL	rs7096206	rs11003125	rs1800450	rs1800451	rs5030737	haplotype	Rel. freq.
	-221	-550	+54	+57	+52		(SEM)
High	Y	Н	А	А	А	HYA	27.3 (1.9)
	Y	L	Α	Α	Α	LYA	22.8 (1.9)
Intermediate	Х	L	Α	А	Α	LXA	22.8 (1.9)
	Y	н	Α	Α	D	HYD	6.5 (1.2)
	Y	н	В	Α	D	HYBD	1.8 (0.7)
	Х	н	Α	Α	Α	HXA	0.6 (0.4)
Low	Y	L	В	Α	Α	LYB	15.2 (1.7)
	Y	L	Α	С	Α	LYC	2.4 (0.7)
Undetermined	Х	Н	А	А	D	HXD	0.6 (0.4)

Table 2. Distribution according to extended haplotypes, and to high, intermediate and low MBL2 secretor haplotypes

Table 3. Baseline characteristics according to extended MBL-2 haplotypes

	HYA	LYA	LXA	HYD	HYBD	HXA	LYB	LYC	P-value
Age (yrs)	55.7	56.8	56.4	55.1	59.7	62.4	55.2	58.4	0.15
	(0.5)	(0.5)	(0.7)	(1.2)	(2.1)	(2.5)	(0.8)	(1.5)	
BMI (kg/m ²)	59.9	26.1	25.8	26.3	26.3	24.9	26.2	25.2	0.7
	(0.2)	(0.2)	(0.2)	(0.4)	(0.8)	(1.2)	(0.3)	(0.5)	
LDL (mmol/L)	4.23	4.26	4.28	4.33	4.10	4.36	4.28	3.98	0.82
	(0.05)	(0.06)	(0.07)	(0.11)	(0.17)	(0.32)	(0.08)	(0.16)	
HDL (mmol/L)	0.94	0.94	0.95	0.93	0.93	1.02	0.93	0.95	0.99
	(0.01)	(0.02)	(0.02)	(0.03)	(0.05	(0.1)	(0.02)	(0.05)	
TG (mmol/L)	1.73	1.77	1.82	1.86	1.74	1.35	1.78	1.78	0.85
	(0.05)	(0.05)	(0.06)	(0.11)	(0.25)	(0.10)	(0.08)	(0.18)	
CRP (mg/L)	0.90	0.98	0.98	0.82	1.62	ND	1.02	0.90	0.89
	(0.09)	(0.10)	(0.12)	(0.21)	(0.7)		(0.17)	(0.34)	
WBC count (*10 ⁹ /L)	7.12	7.15	7.01	7.07	7.74	6.85	7.30	7.80	0.73
	(0.13)	(0.14)	(0.16)	(0.24)	(0.61)	(0.67)	(0.21)	(0.67)	
Family history for CVD (%)	51	46	44	51	30	50	49	52	0.84
History of myocardial	47	47	48	49	70	75	48	38	0.77
infarction (%)									
Multivessel disease (%)	56.2	57.3	59.4	60.8	50.0	50.0	64.1	61.9	0.92
MSD (millimeters)	2.74	2.72	2.76	2.79	2.76	2.58	2.69	2.85	0.55
	(0.02)	(0.03)	(0.03)	(0.06)	(0.11)	(0.19)	(0.04)	(0.07)	
MOD (millimeters)	1.79	1.77	1.78	1.83	1.81	1.70	1.77	1.80	0.97
	(0.02)	(0.02)	(0.03)	(0.05)	(0.11)	(0.22)	(0.04)	(0.06)	

Data are mean±SEM. WBC count; white blood cell count. P-value by ANOVA.

Baseline characteristics of the REGRESS patients population divided into different MBL2 secretor haplotypes are listed in Table 4 and did not show any differences between the groups. There were also no differences in the use of medication between the three groups (data not shown).

	High	Intermediate	Low	P-value
Age (yrs)	56.2 (0.36)	56.3 (0.56)	55.7 (0.73)	0.77
BMI (kg/m ²)	26.0 (0.1)	25.9 (0.2)	26.0 (0.3)	1.0
LDL (mmol/L)	4.24 (0.04)	4.28 (0.05)	4.23 (0.07)	0.78
HDL (mmol/L)	0.94 (0.01)	0.95 (0.01)	0.93 (0.02)	0.82
TG (mmol/L)	1.75 (0.04)	1.82 (0.05)	1.78 (0.07)	0.54
CRP (mg/L)	0.94 (0.07)	0.96 (0.10)	0.99 (0.15)	0.94
WBC count (*10 ⁹ /L)	7.14 (0.10)	7.06 (0.13)	7.39 (0.21)	0.35
Family history CVD (%)	49	45	50	0.64
History of myocardial infarction (%)	47	50	46	0.71
Multivessel disease (%)	56.70	59.10	63.70	0.37
MSD (millimeters)	2.73 (0.02)	2.76 (0.03)	2.71 (0.03)	0.45
MOD (millimeters)	1.78 (0.02)	1.79 (0.02)	1.77 (0.03)	0.89

Table 4. Baseline characteristics according to high, intermediate and low MBL2 secretor haplotypes

Data are mean±SEM. WBC count; white blood cell count. P-value by ANOVA.

Change of MSD and MOD after 2 years of therapy (Figure 1)

Of the total group 194 and 204 patients received placebo or pravastatin, respectively. After 2 years of treatment, there were no significant differences for the occurrence of cardiovascular events between the low, intermediate and high MBL2 secretor haplotypes in the placebo (10.2% vs. 21.6% vs. 19.2% respectively, P=0.17) and pravastatin (6.2% vs. 6.5% vs. 7.2% respectively, P=0.96) treated groups.

In the placebo group, the subjects with the intermediate MBL2 secretor haplotypes showed the highest increase of MSD (0.142±0.025 mm) compared to the lowest MBL2 group (0.037±0.040 mm) (P=0.03 by ANOVA, Figure 1A). Furthermore, the intermediate MBL2 group showed a trend to a higher increase of MOD (0.154±0.026 mm) compared to the low (0.113±0.056) (P=0.08 by ANOVA; Figure 1B). In the pravastatin treated groups these differences were absent for both MSD and MOD.

Subsequently, we compared the effects of placebo and pravastatin treatments on the 2 years change of MSD and MOD. Here, the test for interaction between the different MBL2 secretor haplotypes and therapy in both placebo and pravastatin groups showed that there was no relation between the change in MSD (P=0.16) and MOD (P=0.20) and MBL2 secretor groups.



Figure 1. Mean ± SEM progression of mean segement diameter (MSD) (Figure 1A) and minimal obstruction diameter (MOD) (Figure 1B) over 2 years by placebo and pravastatin in subjects with proven coronary artery disease divided in 3 groups of MBL2 secretor haplotypes (high, intermediate and low).

DISCUSSION

The present study suggests that there is no interaction between MBL2 haplotypes and pravastatin therapy in regard to the progression of CAD. In the past few years the relationship between MBL and atherosclerosis has not been settled in the literature. Both, low (3-9) and high (15-19) MBL2 secretor haplotypes, have been associated with the generation of atherosclerosis. This is the first study showing that the progression of coronary sclerosis does not depend on the MBL2 genotypes and that the treatment effect of pravastatin is not influenced by MBL2 haplotypes. If anything, the present data suggest that low MBL may protect from progression of coronary atherosclerosis compared to intermediate haplotypes. It is the intermediate group that seemed to benefit most from intervention. Furthermore, we have shown that MBL2 haplotypes do not predict cardiovascular events in subjects with proven CAD in a 2 years follow up program.

The baseline MBL2 haplotype distribution was comparable with other studies conducted in Caucasians (12,13). This finding is interesting since all subjects in the REGRESS cohort had significant coronary atherosclerosis. If MBL2 secretor haplotypes were truly risk factors for atherogenesis, we should have found a higher frequency of one these secretor haplotypes in our group. Moreover, the fact that there were no differences in the history of myocardial infarction or for the family history for cardio-vascular disease strengthens the view that in the REGRESS cohort the MBL2 haplotypes do not represent a risk factor for atherosclerosis. However, due to the study design in which no subjects without coronary atherosclerosis were included, we were not able to compare baseline values of the REGRESS patients with non-atherosclerotic controls in a prospective study.

Activation of the complement pathways results in cleavage of complement component 3 (C3), which is central in these pathways (10). When MBL binds to its target (usually a microorganism), MBL-associated serine protease (MASP) functions as a convertase to clip C3 into C3a and C3b. Recent studies have described that elevated C3 is associated with CAD, insulin resistance, obesity, elevated fasting and postprandial triglycerides and disturbed postprandial free fatty acid handling (26-29). Furthermore, it has been shown that there is deposition of complement colocalized with CRP in atherosclerotic plaques (30,31) and that complement activation also plays a role in the induction of tissue damage after myocardial infarction (32). Therefore, the role of complement within atherosclerosis is well established. The role of MBL, however, is still subject of debate in the literature (3-9,15-19). Our results suggest that intermediate MBL2 secretor haplotypes are associated to increased progression and that these subjects might benefit the most from pravastatin treatment. However, the test for interaction between the MBL2 secretor haplotypes and therapy in both placebo and pravastatin groups showed that there was no relation for therapy and the change in progression of coronary atherosclerosis. One has to realize that the formation of activated C3 does not only depend on the MBL route, but can also be achieved by the alternative and classical pathways (10). All these data suggest that progression and generation of atherosclerosis may not be associated to MBL, but involves the complex system of complement with all its pathways and components. Future studies are needed to explore these mechanisms.

Our study has several limitations. Firstly, the REGRESS cohort only consisted of men. Keller et al found that increased serum MBL levels were a risk factor for future CAD in men but not in women (17). We did not measure MBL activity directly. However, it is known that the interplay between the promoter and structural gene variants controls the basal serum level and therefore the activity of MBL (12,13). An additional advantage was that genotyping also allowed us to study the mutations and their contribution to the progression of atherosclerosis. The second limitation in our study is the fact that rs7096206 and rs1800450 were not in Hardy-Weinberg equilibrium. This may be explained in two ways. Firstly, the locus being under selection,

embreeding or being analyzed in a population of multiracial origin. The latter is not the case since all the subjects were Caucasians. However, we can not rule out the possibility of the loci being under selection or embreeding. Secondly, there is a possibility of genotyping errors. Also this can not be completely ruled out, but the fact that the samples of known genotypes were included in every analysis as control and that the allele frequencies were fairly similar to the published data in all cases, is reassuring.

The final limitation of the study is the relative short length of follow-up of two years, which might be too short to detect subtle changes. Unfortunately, the original REGRESS study was designed as a two year trial. However, since the power of our study was above 80% for moderate effect-sizes of at least three of the MBL2 SNPs, we may conclude that there is no association between MBL2 SNPs and the MSD/MOD changes in 2 years of follow-up.

In conclusion, in men with proven coronary artery disease, the low, intermediate and high MBL2 secretor haplotypes do not evidently predict the progression of atherosclerotic burden and cardiovascular events in 2 years of follow up. Furthermore, there is no significant interaction between pravastatin therapy and MBL2 secretor haplotypes in regard to progression of coronary atherosclerosis.

ACKNOWLEDGEMENTS

We are grateful to all the patients, doctors and co-workers who made DNA samples available to us.

Funding

The financial support for this study was provided by Research Foundation Internal Medicine of the Sint Franciscus Gasthuis in Rotterdam, The Netherlands. The REGRESS study was supported by Bristol-Myers Squibb Co, Princeton, NJ. CIBER de Diabetes y Enfermedades Metabólicas Asociadas is an ISCIII project.

Disclosures None declared.

REFERENCES:

- 1. Ross R. Atherosclerosis--an inflammatory disease. N Engl J Med 1999;340:115-26.
- Danesh JP, Whincup P, Walker M, Lennon L, Thomson A, Appleby P, Gallimore JR, Pepys MB. Low grade inflammation and coronary heart disease: prospective study and update meta-analysis. BMJ 2000; 321:199-204.
- 3. Madsen HO, Videm V, Svejgaard A, Svennevig JL, Garred P. Association of mannose-binding lectin deficiency with severe atherosclerosis. The Lancet 1998;352:959-60.
- 4. Best LG, Davidson M, North KE, MacCluer JW, Zhang Y, Lee ET, Howard BV, DeCroo S, Ferrell RE. Prospective analysis of mannose-binding lectin genotypes and coronary artery disease in American Indians. Circulation 2004;109:471-5.
- 5. Hegele RA, Ban MR, Anderson CM, Spence JD. Infection-susceptibility alleles of mannose-binding lectin are associated with increased carotid plaque area. J Invest Med 2000;48:198-202.
- Ohlenschlaeger T, Garred P, Madsen HO, Jacobsen S. Mannose-binding lectin variant alleles and the risk of arterial thrombosis in systemic lupus erythematosus. N Engl J Med 2004;351;260-7.
- Biezeveld MH, Kuipers IM, Geissler J, Lam J, Ottenkamp JJ, Hack CE, Kuijpers TW. Association of mannose-binding lectin genotype with cardiovascular abnormalities in Kawasaki disease. Lancet 2003;361;1268-70.
- Fiane AE. Ueland T, Simonsen S, Scott H, Endresen K, Gullestad L, Geiran OR, Haraldsen G, Heggelund L, Andreassen AK, Wergeland R, Froland S, Aukrust P, Mollnes TE. Low mannose-binding lectin and increased complement activation correlate to allograft vasculopathy, ischaemia, and rejection after human heart transplantation. Eur Heart J 2005;26:1660-5.
- Alipour A, van Oostrom AJHHM, Van Wijk JPH, Verseyden C, Plokker HWM, Jukema JW, Rabelink AJ, Castro Cabezas M. Mannose binding lectin deficiency and triglyceride-rich lipoprotein metabolism in normolipidemic subjects. Atherosclerosis 2009;206:444-50.
- 10. Walport JW. Complement; first of two parts. N Engl J Med 2001;344:1058-1065.
- 11. Turner MW. The role of mannose-binding lectin in health and disease. Mol Immunol 2003;40:423-9.
- 12. Madsen HO, Garred P, Thiel S, Kurtzhals JAL, Lamm LU, Ryder LP, Svejgaard A. Interplay between promotor and structural gene variants control basal serum level of mannan-binding protein. J Immunol 1995;155:3013-20.
- Crosdale DJ, Ollier WE, Thomson W, Dyer PA, Jensenious J, Johnson RWG, Poulton KV. Mannose binding lectin (MBL) genotype distributions with relation to serum levels in UK Caucasoids. Eur J Immunogenet 2000;27:111-7.
- 14. Salimans MM, Bax WA, Stegemans F, van Deuren M, Bartelink AK, van Dijk H. Association between familial deficiency of mannose-binding lectin and mutations in the corresponding gene and promoter region. Clin Diagn Lab Immun 2004;11:806-7.
- Aittoniemi J, Fan YM, Laaksonen R, Janatuinen T, Vesalainen R, Nuutila P, Knuuti J, Hulkkonen J, Hurme M, Lehtimaki T. The effect of mannan-binding lectin variant alleles on coronary artery reactivity in healthy young men. Int J Cardiol 2004;97:317-8.
- Hansen TK, Tarnow L, Thiel S, Steffensen R, Stehouwer CD, Schalkwijk CG, Parving HH, Flyvbjerg A. Association between mannose-binding lectin and vascular complications in type 1 diabetes. Diabetes 2004;53:1570-6.
- 17. Keller TT, van Leuven SI, Meuwese MC, Wareham NJ, Luben R, Stroes ES, Hack CE, Levi M, Khaw KT, Boekholdt SM. Serum levels of mannose-binding lectin and the risk of future coronary artery disease in apparently healthy men and women. Arterioscler Thromb Vasc Biol 2006;26:2345-50.

- Rugonfalvi-Kiss S, Dósa E, Madsen HO, Endrész V, Prohászka Z, Laki J, Karádi I, Gönczöl E, Selmeci L, Romics L, Füst G, Entz L, Garred P. High rate of early restenosis after cartotid eversion endarteriectomy in homozygous carriers of the normal mannose-binding lectin deficiency. Stroke 2005;36:944-8.
- Collard CD, Shernan SK, Fox AA, Bernig T, Chanock SJ, Vaughn WK, Takahashi K, Ezekowitz AB, Jarolim P, Body SC. The MBL2 'LYQA secretor' haplotype is an independent predictor of postoperative myocardial infarction in whites undergoing coronary artery bypass graft surgery. Circulation 2007;116(11 Suppl):1106-12.
- 20. Jukema JW, Bruschke AV, van Boven AJ, Reiber JH, Bal ET, Zwinderman AH, Jansen H, Boerma GJ, van Rappard FM, Lie KI; on behalf of the REGRESS Study Group, Interuniversity Cardiology Institute Utrecht, Netherlands. Effects of lipid lowering by pravastatin on progression and regression of coronary artery disease in symptomatic men with normal to moderately elevated serum cholesterol levels: the Regression Growth Evaluation Statin Study (REGRESS). Circulation 1995;91:2528–40.
- LaRosa JC, Grundy SM, Waters DD, Shear C, Barter P, Fruchart JC, Gotto AM, Greten H, Kastelein JJ, Shepherd J, Wenger NK; Treating to New Targets (TNT) Investigators. Intensive lipid lowering with atorvastatin in patients with stable coronary disease. N Engl J Med 2005;352:1425-35.
- Smilde TJ, van Wissen S, Wollersheim H, Trip MD, Kastelein JJ, Stalenhoef AF. Effect of aggressive versus conventional lipid lowering on atherosclerosis progression in familial hypercholesterolaemia (ASAP): a prospective, randomised, double-blind trial. The Lancet 2001;357:577-81.
- 23. Halkes CJ, van Dijk H, de Jaegere PP, Plokker HW, van der Helm Y, Erkelens DW, Castro Cabezas M. Postprandial increase of complement component 3 in normolipidemic patients with coronary artery disease: effects of expanded-dose simvastatin. Arterioscler Thromb Vasc Biol 2001;21:1526-30.
- 24. Stephens M, Smith N, Donnelly, P. A new statistical method for haplotype reconstruction from population data. Am J Hum Genet 2001;68:978-89.
- Souverein OW, Zwinderman AH, Tanck MW. Estimating haplotype effects on dichotomous outcome for unphased genotype data using a weighted penalized log-likelihood approach. Hum Hered 2006; 61:104-10.
- 26. Muscari A, Bozzoli C, Puddu GM, Sangiorgi Z, Dormi A, Rovinetti C, Descovich GC, Puddu P. Association of serum C3 levels with the risk of myocardial infarction. Am J Med 1995;98:357-64.
- 27. Verseyden C, Meijssen S, van Dijk H, Jansen H, Castro Cabezas M. Effects of atorvastatin on fasting and postprandial complement component 3 response in familial combined hyperlipidemia. J Lipid Res 2003;44:2100-08.
- Van Oostrom AJHHM, Alipour A, Plokker HWM, Sniderman AD, Castro Cabezas M. The metabolic syndrome in relation to complement component 3 and postprandial lipemia in patients from an outpatient lipid clinic and healthy volunteers. Atherosclerosis 2007;90:167-73.
- Muscari A, Massarelli G, Bastagli L, Poggiopollini G, Tomassetti V, Drago G, martignani C, Pacilli P, Boni P, Puddu P. Relationship of serum C3 to fasting insulin, risk factors and previous ischaemic events in middle-aged men. Eur Heart J 2000;21:1081-90.
- 30. Oksjoki R, Kovanen PT, Pentikainen MO. Role of complement activation in atherosclerosis. Curr Opin Lipidol 2003;14:477-82.
- Torzewski J, Torzewski M, Bowyer DE, Fröhlich M, Koenig W, Waltenberger J, Fitzsimmons C, Hombach V.. C-reactive protein frequently colocalizes with the terminal complement complex in the intima of early atherosclerotic lesions of human coronary arteries. Arterioscler Thromb Vasc Biol 1998;18:1386-92.
- 32. Griselli M, Herbert J, Hutchinson WL, Taylor KM, Sohail M, Krausz T, Pepys MB. C-reactive protein and complement are important mediators of tissue damage in acute myocardial infarction. J Exp Med 1999;190:1733-40.